

1-1-2017

Proteoglycan neofunctions: regulation of inflammation and autophagy in cancer biology.

Liliana Schaefer
Klinikum der Goethe-Universiti

Claudia Tredup
Klinikum der Goethe-Universiti

Maria A. Gubbiotti
Thomas Jefferson University

Renato V. Iozzo
Thomas Jefferson University

Follow this and additional works at: <https://jdc.jefferson.edu/pacbfp>

 Part of the [Oncology Commons](#), and the [Pathology Commons](#)

[Let us know how access to this document benefits you](#)

Recommended Citation

Schaefer, Liliana; Tredup, Claudia; Gubbiotti, Maria A.; and Iozzo, Renato V., "Proteoglycan neofunctions: regulation of inflammation and autophagy in cancer biology." (2017). *Department of Pathology, Anatomy, and Cell Biology Faculty Papers*. Paper 230.
<https://jdc.jefferson.edu/pacbfp/230>

This Article is brought to you for free and open access by the Jefferson Digital Commons. The Jefferson Digital Commons is a service of Thomas Jefferson University's [Center for Teaching and Learning \(CTL\)](#). The Commons is a showcase for Jefferson books and journals, peer-reviewed scholarly publications, unique historical collections from the University archives, and teaching tools. The Jefferson Digital Commons allows researchers and interested readers anywhere in the world to learn about and keep up to date with Jefferson scholarship. This article has been accepted for inclusion in Department of Pathology, Anatomy, and Cell Biology Faculty Papers by an authorized administrator of the Jefferson Digital Commons. For more information, please contact: JeffersonDigitalCommons@jefferson.edu.



Published in final edited form as:

FEBS J. 2017 January ; 284(1): 10–26. doi:10.1111/febs.13963.

Proteoglycan neofunctions: Regulation of inflammation and autophagy in cancer biology

Liliana Schaefer¹, Claudia Tredup¹, Maria A. Gubbiotti², and Renato V. Iozzo²

¹Pharmazentrum Frankfurt/ZAFES, Institut für Allgemeine Pharmakologie und Toxikologie, Klinikum der Goethe-Universität Frankfurt am Main, Frankfurt am Main, Germany

²Department of Pathology, Anatomy and Cell Biology, and the Cancer Cell Biology and Signaling Program, Kimmel Cancer Center, Thomas Jefferson University, Philadelphia, USA

Abstract

Inflammation and autophagy have emerged as prominent issues in the context of proteoglycan signaling. In particular, two small, leucine-rich proteoglycans, biglycan and decorin, play pivotal roles in the regulation of these vital cellular pathways and, as such, are intrinsically involved in cancer initiation and progression. In this minireview we will address novel functions of biglycan and decorin in inflammation and autophagy, and analyze new emerging signaling events triggered by these proteoglycans, which directly or indirectly modulate these processes. We will critically discuss the dual role of proteoglycan-driven inflammation and autophagy in tumor biology, and delineate the potential mechanisms through which soluble ECM constituents affect the microenvironment associated with inflammatory and neoplastic diseases.

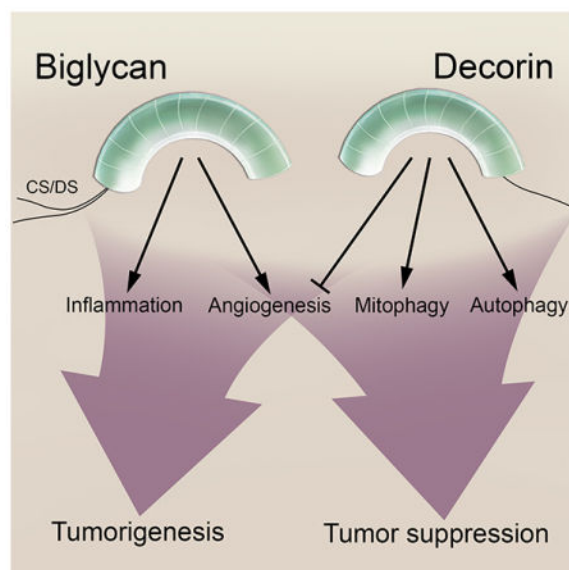
Graphical Abstract

We critically assess the role of two small, leucine-rich proteoglycans, biglycan and decorin, in inflammation and autophagy. The signaling axes of these two proteoglycans are operative during various stages of cancer initiation and progression. Moreover, we discuss the dual role of proteoglycan-driven inflammation and autophagy in tumor biology, and offer unique roles for these molecules as potential therapeutic agents.

Correspondence to: Liliana Schaefer: Pharmazentrum Frankfurt, Institut für Allgemeine Pharmakologie und Toxikologie, Klinikum der Goethe-Universität Frankfurt am Main, Haus 74, Z. 3.108a, Theodor-Stern-Kai 7, 60590 Frankfurt am Main, Germany. schaefer@med.uni-frankfurt.de. Renato V. Iozzo: Department of Pathology, Anatomy and Cell Biology, Sidney Kimmel medical College at Thomas Jefferson University, 1020 Locust Street, Jefferson Alumni Hall Suite 336, Philadelphia, PA 19107, USA, renato.iozzo@jefferson.edu.

Author contributions

L.S., C.T., M.A.G. and R.V.I. conceived the ideas, wrote the manuscript and created the figures.



Keywords

Small leucine-rich proteoglycans; decorin; biglycan; receptor tyrosine kinase; Toll-like receptors; LC3

Introduction

For decades, the two closely related small, leucine-rich proteoglycans (SLRPs), biglycan and decorin, have been considered as rather structural elements of the extracellular matrix (ECM). For example, biglycan has long been regarded as an important factor for bone mineralization and decorin originally gained recognition for its role in the regulation of collagen fibrillogenesis [1]. However, this paradigm shifted when biglycan and decorin were identified as functional signaling molecules, particularly in the context of inflammation and autophagy regulation. As these processes play a crucial role both in the initiation and progression of tumorigenesis, and as these proteoglycans distinguish themselves further as dynamic players in cell signaling, it has become apparent that biglycan and decorin, and their downstream signaling effectors, might represent druggable therapeutic targets for the treatment of malignant disease. The role of syndecan and heparanase, two key players in cancer progression, is also covered in this thematic minireview series [2,3].

In this review, we provide an inclusive and critical assessment of the roles of biglycan and decorin in autophagy as well as in inflammatory signaling pathways, with a strong emphasis on their functions in the setting of cancer. We will further evaluate the intricate connection between these endogenous matrix constituents and both normal and pathological cellular activity to provide insight into future options for the treatment of cancer and other diseases where inflammation is predominant.

Matrix regulation of endothelial cell autophagy and tumor cell mitophagy

Neovascularization is a long-standing hallmark in the context of cancer progression as the growth of new blood vessels allows the tumor proper to acquire necessary nutrients to enlarge and metastasize. As such, the advent of anti-angiogenic compounds has opened a new era of cancer treatment and the development of next-generation angiogenesis inhibitors remains a popular basis for chemotherapeutic drug design. Though a variety of pharmacologic agents have been engineered to target this process, an important discovery revealed that the ECM harbors an endogenous angiostatic factor in the class I small, leucine-rich proteoglycan, decorin [4]. Initial studies have demonstrated the ability of decorin to regulate collagen fibrillogenesis, thereby establishing a function for decorin in the preservation of the structural integrity of ECM [5–9]. Subsequent work has revealed that decorin plays a much more active role by regulating key cellular processes through its aptitude for binding to different receptor tyrosine kinases (RTKs) and modifying downstream signaling pathways [10–20]. In tumor cells, decorin acts primarily through the hepatocyte growth factor (HGF/Met) and epidermal growth factor receptor (EGFR) signaling axes to decrease the expression of the pro-angiogenic factors, vascular endothelial growth factor A (VEGFA) and hypoxia-inducible factor 1- α (HIF-1 α), while simultaneously increasing the expression of thrombospondin-1 [11,21], a powerful anti-angiogenic protein [22], and tissue inhibitor of metalloproteinases-3 (TIMP3) [11,21], a key regulator of ECM turnover [11,21] (Fig. 1). The cumulative effect of these changes of the tumor secretome creates an anti-angiogenic and, subsequently less metastatic, tumor microenvironment.

Though decorin can alter the cellular milieu by controlling the secretion of angiogenic factors from tumor cells themselves, more recent work has uncovered a novel mechanism where decorin attenuates tumor progression by directly targeting the tumor microenvironment rather than acting solely on the tumor proper. A unique screen of orthologous tumor xenografts capable of differentiating between human and mouse gene expression revealed an increase in the tumor suppressor gene, paternally expressed gene 3 (*Peg3*), in decorin-treated animals compared to vehicle-treated controls [23]. Interestingly, this decorin-evoked gene induction occurs only in the tumor stroma and not in the tumor itself [23]. Given this information, a more detailed analysis of *Peg3* expression in tumor-supporting cells upon decorin treatment revealed that *Peg3* plays a role in mediating endothelial cell autophagy, the accumulation of cytoplasmic contents in double-membrane structures, which are eventually degraded via lysosomal machinery [24,25]. Mechanistically, decorin acts as a partial agonist for vascular endothelial growth factor receptor 2 (VEGFR2), the main receptor responsible for facilitating the angiogenic response in endothelial cells, resulting in a signaling cascade culminating in *Peg3*-dependent canonical autophagy involving the standard autophagic intermediates, Beclin 1 and microtubule-associated light chain protein 3 (LC3) [13,17] (Fig. 1). Decorin also competes with VEGFA, the canonical ligand of VEGFR2, to hinder angiogenesis [26]. As VEGFR2 is specifically expressed on endothelial cells, it is plausible that the anti-angiogenic property of decorin is linked to its induction of autophagy exclusively in the tumor vasculature. These findings are the first to link a soluble extracellular proteoglycan to the induction of the catabolic process of

autophagy and expose a novel function for matrix molecules to regulate the lysosomal degradation pathway.

Regarding the aforementioned decorin-mediated alteration of thrombospondin-1 secretion by tumor cells, it appears that thrombospondin-1 is surfacing as an important autophagic signaling molecule. For example, thrombospondin-1 inhibits RAS-positive tumor cell growth via autophagic induction [27]. On the contrary, studies blocking the function of CD47, the main thrombospondin-1 receptor, show a paradoxical increase in autophagic activity [28]. Thus, the ability of thrombospondin-1 to modulate autophagy appears to be quite complex, but may be important for putting together more pieces of the puzzle to understand better the ability of decorin to inhibit neovascularization.

Also of importance is the recent discovery that decorin is an autophagy-inducible proteoglycan [12,16] where nutrient deprivation and chemical inhibition of mammalian target of rapamycin (mTOR) signaling augment its expression both *in vitro* in fibroblasts as well as *in vivo* in cardiac tissue. In addition, *Dcn*^{-/-} mice exhibit diminished cardiac autophagy vis-à-vis wild-type mice following a fasting challenge [12], validating the importance of decorin as a pivotal control center for the regulation of this catabolic process [16]. As pro-autophagic stimuli increase decorin expression, it is possible that the ability to modulate decorin levels in the tumor stroma may be a novel method for targeting neovascularization in the setting of cancer. Furthermore, in order to connect the processes of tumor progression and autophagy, a worthy hypothesis may be that, in the absence of decorin, lower levels of autophagy promote a pro-tumorigenic microenvironment.

In contrast, biglycan, another class I SLRP that shares the most structural homology with decorin [29,30], possesses pro-angiogenic properties through its ability to bind VEGFA and activate VEGFR2 signaling [31]. Additionally, when overexpressed in colon cancer cells, biglycan upregulates VEGFA leading to increased angiogenesis and tumor growth through activation of the extracellular signal-regulated kinase (ERK) pathway [32]. Therefore, the capacity to modulate angiogenesis appears to be specific to each respective proteoglycan, even within closely related members of the same family. Whether biglycan-mediated angiogenesis is related to autophagy remains to be seen. Interestingly, biglycan inhibits the angiostatic effects of endostatin [30]. As endostatin is a potent autophagic inducer in endothelial cells through the modulation of $\alpha 5\beta 1$ integrin signaling culminating in increased Beclin 1 expression [30], it is possible that biglycan may inhibit autophagy by diminishing endostatin activity.

Like biglycan, perlecan, a large, basement membrane proteoglycan, exhibits pro-angiogenic properties [33–45]. Specifically, through its heparan sulfate side chains, the N-terminal domain I of perlecan sequesters and presents VEGFA and fibroblast growth factors (FGFs) to their respective receptors resulting in enhanced angiogenesis [46–51]. Importantly, mice that have been genetically designed without the heparan sulfate attachment site display diminished angiogenesis and compromised wound healing, presumably due to impairment in the juxtaposition of growth factors with their signaling receptors [52,53]. Along with its pro-angiogenic properties, perlecan also appears to be anti-autophagic as a recent notable discovery illustrates that conditional *Heparan sulfate proteoglycan 2*^{-/-} (*Hspg2*^{-/-}) mice

display enhanced autophagy in skeletal muscle [54] where they exhibit less mTOR activity than wild-type cohorts.

We note that perlecan is a multi-modular proteoglycan, where different domains serve different functions [55]. For example, while its N-terminal domain is pro-angiogenic, its C-terminal fragment, endorepellin, embraces properties very similar to those of decorin, where it is capable of restricting endothelial cell migration, capillary morphogenesis, and angiogenesis [35]. Indeed, previous studies demonstrate that endorepellin exclusively targets the tumor vasculature, resulting in decreased tumor growth [56]. In addition, endorepellin is able to initiate autophagy in endothelial cells utilizing a mechanism comparable to that of decorin, as both work through the Peg3-signaling axis to induce the pro-autophagic intermediates Beclin 1 and LC3 [57]. As both endorepellin and decorin exhibit the capability to induce autophagy as well as inhibit tumorigenesis through their actions in the tumor microenvironment, it is again likely that one mechanism by which this occurs is via suppression of angiogenesis through autophagic degradation of the vasculature. In fact, recent work suggests that the angiostatic mechanism of endorepellin is directly linked to autophagic induction [58]. Specifically, endorepellin inhibits vessel sprouting in *ex vivo* aortic ring assays, which can be blocked by inhibiting autophagy via the adenosine monophosphate kinase (AMPK) inhibitor, compound C [58]. Therefore, as decorin follows this same mechanistic paradigm, it is likely that it too reduces tumor neovascularization via an autophagic mechanism. Interestingly, endorepellin interacts with endostatin [35], the aforementioned potent pro-autophagic matrix protein. As this interaction diminishes endostatin's ability to reduce angiogenesis in endothelial cells, it may also prevent autophagic induction by either protein via a competitive binding mechanism.

Taken together, it appears that a distinct network is arising in the ECM that is important for the regulation of normal cell homeostasis as well as to counteract dysregulation of homeostasis in the tumor microenvironment. *In vivo*, it seems that there exist intricate feedback loops to allow for the induction and suppression of both autophagy and angiogenesis in response to environmental cues. Preliminary evidence for this stems from the observation that anti-angiogenic proteoglycans, like decorin [4], seem to be pro-autophagic and vice versa. As an aberration of either process can be detrimental to normal physiology, these pathways may become deregulated following some sort of insult or environmental stressor, leading to the onset of cancer. Continued deregulation may contribute to the progression of cancer and eventually to metastasis. As we continue to examine the signaling capabilities of these proteoglycans, we will gain a better perspective about the changes in the tumor microenvironment that lead to metastasis and how they are related to proteoglycan function or dysfunction. Furthermore, this knowledge will provide new management options for cancer where mimetics of endogenously occurring matrix components can be utilized for a safer, more efficacious treatment regimen than existing options.

Curtailing cancer progression: Targeting the tumor proper

Though we have focused mainly on detailing the actions of decorin and related proteoglycans in the tumor stroma, we must acknowledge that decorin, which is biologically

active as a monomer [59], also engages directly with the tumor proper via its interaction with a variety of RTKs abundant on the surface of different types of cancer cells [60–62]. One of the best-known decorin/RTK interactions is that between decorin and the EGFR resulting in p21-mediated cell cycle arrest, apoptosis, and diminished angiogenesis [63–65] (Fig. 1). Furthermore, decorin allows for EGFR to be degraded via caveolin-mediated endocytosis [66]. In addition, decorin antagonizes ErbB2/4 and platelet-derived growth factor (PDGF) signaling, both resulting in decreased tumor growth [67–69].

As described above, decorin interacts with the HGF/Met receptor to reduce angiogenesis by modifying the tumor secretome [70] (Fig. 1). In the tumor parenchyma, this same interaction results in an attenuation of β -catenin and Myc signaling, thereby directly inhibiting tumor growth [71] (Fig. 1). Notably, the decorin/Met interaction in triple-negative breast carcinoma cells evokes tumor cell mitophagy via the tumor suppressor, mitostatin [72] (Fig. 1). This decorin-evoked mitophagy likely comes as a consequence of mitochondrial depolarization and is the first documentation that decorin can regulate catabolism in cancer cells. Thus, it is likely that some of decorin's anti-tumorigenic activity comes through its interference with proper oxidative phosphorylation resulting in an energy imbalance in the setting of cancer. Furthermore, loss of mitostatin results in the inability of decorin to suppress VEGFA [72], coupling the ability of decorin to act simultaneously in the tumor microenvironment and the tumor proper.

Canonical autophagy in tumor cells in response to decorin stimulation has been recently reported in glioma cells, where decorin-mediated autophagic induction contributes to a reduction in cellular migration [73]. Though this is the first report of decorin-induced autophagy directly in cancer cells, it is likely that tumor cells that possess the proper receptors and subsequent signaling machinery also undergo autophagy in response to decorin treatment. For example, recent studies posit that inhibition of EGFR signaling results in autophagic induction [74]. As decorin is a potent ligand of EGFR, it is probable that tumor cells over-expressing this receptor, such as A431 squamous carcinoma cells or A549 lung carcinoma cells, respond with canonical autophagy instead of, or in concert with, mitophagy when challenged with decorin. Nevertheless, as our lab has demonstrated the necessity of Peg3 for decorin-mediated autophagy in endothelial cells, it would be unsurprising if decorin induces autophagy only in few tumor types as many cancers possess little to no endogenous Peg3 [75,76].

Again, despite being in the same family, biglycan takes on different functions in the context of tumor signaling. In fact, several tumor types, including colon and pancreatic cancer, express biglycan at higher levels than normal tissue substantiating the idea that biglycan is pro-tumorigenic as well as pro-angiogenic [77–79]. For instance, upregulation of biglycan in gastric cancer mechanistically promotes invasion and migration of these cells via phosphorylation and consequent activation of the focal adhesion kinase (FAK) signaling pathway [80]. In colon cancer cells, depletion of biglycan results in decreased proliferation and p21-mediated cell cycle arrest [32]. In addition, biglycan is capable of inhibiting apoptosis in these colon cancer cells by regulating p38 MAPK signaling [32]. Moreover, though its ability to augment Wnt/ β -catenin signaling has only been explored in bone, some of the pro-tumorigenic effects of biglycan may also be attributed to its ability to amplify

Wnt/ β -catenin signaling as well as its ability to interact with LDL-receptor related protein 6 (LRP6) [81]. Interestingly, autophagy attenuates Wnt-signaling, suggesting a potential link between biglycan and this catabolic process in cancer [82]. Also, though direct alterations of signaling pathways via biglycan are largely responsible for increased tumorigenesis, a secondary mechanism may be the ability of biglycan to modify the biophysical properties of the matrix itself. For example, as biglycan modifies FAK-signaling, it is likely that some of the increased invasiveness and tumor progression resulting from biglycan overexpression may come as a result of increased matrix stiffness, thereby facilitating the migration of cancer cells to increase metastasis.

Though the function of autophagy in cancer is controversial, it is emerging as an important therapeutic target. Whether it should be enhanced or inhibited appears to be context-dependent as in some cases autophagy is beneficial for reducing tumor growth and in others it seems necessary for cancer cell survival. Regardless, as both decorin and biglycan are major players in the inhibition and progression of tumorigenesis, parsing out the mechanism behind modulation of the autophagic pathway by them and other matrix constituents is critical for comprehending how to best fine-tune novel treatment strategies. Though the role of decorin in autophagy as it pertains to cancer is much more well-established than that of biglycan, it is very probable that biglycan will develop into a remarkable player in this signaling pathway in the future. Thus, we must focus our efforts on discovering more of the intricacies of these signaling regimes to hone our understanding of matrix regulation of tumor initiation and metastasis.

Complexity of biglycan signaling: Pro-inflammatory and danger signals

Besides its function as a structural molecule, the small leucine-rich proteoglycan biglycan can be either proteolytically released from the ECM upon tissue stress and injury or *de novo* synthesized by activated macrophages and resident cells [83]. In its soluble form, biglycan acts as a signaling molecule and endogenous ligand of the Toll-like receptors (TLR)-2 and -4 on the surface of macrophages. This biological interaction autonomously triggers sterile inflammation and potentiates pathogen-mediated inflammation via a second TLR that is not involved in pathogen sensing [83–85]. Mechanistically, by engaging TLR2 and TLR4, biglycan rapidly activates p38, ERK, and NF- κ B signaling pathways. This leads to the synthesis and secretion of pro-inflammatory cytokines and chemokines, such as interleukin (IL)-1 β , tumor necrosis factor (TNF)- α , chemokine (C-C motif) ligand (CCL)2, chemokine (C-X-C motif) ligand (CXCL)1, CXCL-2, CXCL13, and CCL5 [83,84,86,87] (Fig. 2). This process initiates modulation of the immune environment and facilitates the recruitment of leukocytes to the site of inflammation.

The use of a transient transgenic mouse model in which biglycan is overexpressed in the liver under the albumin promoter and then released into the circulation has been instrumental for the deeper understanding of biglycan signaling *in vivo* [87]. This model has elucidated that soluble biglycan uses TLR2/4 signaling pathways, involving the adaptor molecule myeloid differentiation primary response 88 (MyD88), for the recruitment of neutrophils and macrophages while it regulates the infiltration of T cells exclusively through TLR4 and the

adaptor molecule TIR-domain-containing adapter-inducing interferon- β (TRIF) [87,88] (Fig. 2).

Moreover, biglycan organizes a multi-receptor crosstalk between TLR2/4 and the purinergic receptor, P2X₇ [85]. By clustering TLR2/4 and P2X₇, biglycan triggers the activation of caspase-1 through the NOD-like receptor protein 3 (NLRP3) inflammasome with subsequent release of mature IL-1 β and IL-18 in a reactive oxygen species (ROS)- and heat shock protein (HSP)-90-dependent manner [85]. As a consequence, biglycan-deficient mice show improved survival in LPS-induced sepsis and less severe inflammation in both models of sterile inflammatory renal injury and LPS-induced sepsis, as monitored by decreased levels of active caspase-1 and therefore of mature IL-1 β [83,85,89] (Fig. 2).

Thus, under pathological conditions soluble biglycan in its intact form, containing protein core and glycosaminoglycan (GAG) chain(s), and some biglycan fragments act as potent danger signals that mimic response to Gram-positive (via TLR2) and negative (via TLR4) pathogens [90]. While it is well established that the interaction of biglycan with TLR2 and the TLR4/MD2 complex occurs via protein core [83,84] the exact binding motifs are still under investigation. Interestingly, for the induction of TLR-dependent signaling the protein core of biglycan and at least one GAG side chain are required. Neither biglycan protein core nor GAG chains alone are capable of inducing TLR-dependent signaling and a proinflammatory response [83]. Therefore, it is conceivable that GAG chains of biglycan might be involved in the interaction with co-receptors and adaptor molecules of TLR2 and TLR4.

Proteolytic release of biglycan ensures a rapid inflammatory response to tissue stress or injury without the need for the *de novo* synthesis of biglycan. Several proteolytic enzymes such as bone morphogenetic protein-1 (BMP-1) [91], various matrix metalloproteinases (MMPs) [92] and a disintegrin and metalloproteinase with thrombospondin motifs (ADAMTS) and serine protease Granzyme B (GrB) have been shown to cleave biglycan. For details on biglycan degrading enzymes and cleavage of biglycan please refer to the recent review [93]. The fragments of biglycan acting as danger signals and proteinases involved in their generation are not identified yet. As biglycan protein core together with GAG side chain(s) trigger a proinflammatory response, it is tempting to speculate that not all of the generated biglycan fragments will be capable of binding to TLR2 and TLR4. Therefore, a proper inflammatory signal is secured by a very fast, *de novo* synthesis of biglycan in stimulated macrophages followed by additional synthesis in resident cells.

Biglycan tightly controls the severity of the inflammatory response

A novel mechanism has emerged wherein biglycan fine-tunes IL-1 β production and restores immune homeostasis [89]. By using TLR2/4, biglycan triggers ROS generation in macrophages through NADPH oxidases (NOX) 1 and 4, thereby enhancing the expression and secretion of IL-1 β . To impede this self-induced pro-inflammatory response, biglycan also triggers the synthesis and activation of NOX2 through selective TLR4 signaling pathways. While biglycan-mediated TLR4/TRIF signaling leads to the induction of *Nox2* mRNA expression, induction of TLR4/MyD88 signaling is required for activation of NOX2.

Activated NOX2 inhibits biglycan-mediated IL-1 β synthesis [89]. It is tempting to speculate that these mechanisms of self-limiting inflammation are involved under physiological conditions to fine-tune IL-1 β levels and could be therapeutically harnessed to treat inflammatory diseases.

However, there is strong evidence for beneficial effects of biglycan deficiency and pro-inflammatory effects of biglycan overexpression in various inflammatory diseases [93,94]. In fact, under pathological conditions biglycan stimulates the expression of HSP70 by selective utilization of TLR2. HSP70 binds to NOX2 thereby inducing its proteasomal degradation. This interaction consequently impairs the inhibitory function of NOX2 on IL-1 β expression, promoting pro-inflammatory effects of biglycan (Fig. 2). Accordingly, in a mouse model of sterile renal inflammation, the lack of biglycan in *Nox2*-deficient mice is protective of hyperinflammation as well as massive macrophage infiltration, leading to less severe kidney damage [89].

Thus, these data describe how danger signals control the severity of the inflammatory response, by utilizing multi-receptor signaling, selective induction of TLR2 and TLR4 signaling pathways, adaptor molecules and downstream events such as ROS production. The discovery of the dual role of biglycan in inflammation will open new prospects for therapeutic intervention. It is likely that selective inhibition of biglycan-TLR2- or biglycan-TLR4 signaling could be a new therapeutic approach in inflammatory diseases. Alternatively, the stabilization of biglycan-mediated NOX2 expression or activation might represent a novel pharmacological avenue to overcome inflammation without interfering with physiological TLR signaling.

Biglycan stabilizes HIF-2 α and evokes erythropoiesis

Recently, a novel function of biglycan has been discovered in a transgenic mouse model in which biglycan was constitutively overexpressed. Surprisingly, chronic increase in circulating biglycan leads to elevated hemoglobin concentrations, hematocrit values, and enhanced iron binding capacity, leading essentially to a clinical picture identical to secondary polycythemia [95]. Mechanistically, soluble biglycan induces polycythemia by triggering the hepatic and renal expression of erythropoietin (Epo), a key regulator of erythropoiesis. This occurs exclusively via TLR2 and involves biglycan-mediated stabilization of HIF-2 α , most likely through a hypoxia-independent pathway [95].(Fig. 3). This information provides a new link for biglycan in the regulation of inflammation via stabilizing HIF-2 α , a crucial trigger of pro-inflammatory cytokines, in macrophages [96]. Furthermore, these findings are in line with previous reports that biglycan selectively interacts with TLRs and their adapter molecules to achieve diverse biological outcomes [87,89]. While the clinical relevance of these findings has not been shown yet, it is conceivable that biglycan may critically impact tumorigenesis and cardiovascular disease by enhancing HIF-2 α abundance and triggering secondary polycythemia.

Inflammatory biglycan signaling in tumor cell biology

Despite growing evidence for a relationship among innate immunity, inflammation, and tumor development, the mechanisms responsible for this association are not well-defined [97,98]. Although inflammation is a beneficial and essential mechanism in tissue injury and pathogen defense, it requires a rapid clearance of inflammatory cells [99]. Imbalance of inflammation resolution leads to infiltration of immune cells, which may generate large amounts of cytokines, chemokines, and ROS resulting in tissue damage and chronic inflammation, thus contributing to tumor initiation and progression [100]. Furthermore, the inflammatory microenvironment is considered a hallmark in each stage of tumor development [98].

Taking into account the intimate involvement of biglycan in inflammation, it is not surprising that the overexpression of biglycan is associated with a variety of human malignancies, such as pancreatic adenocarcinoma [101,102], colon cancer [102], ovarian cancer [103] intrahepatic cholangiocarcinoma [104], gastric cancer [80], esophageal squamous cell carcinoma [105], and melanoma [106]. Moreover, biglycan is involved as a regulator of tumorigenesis [60], a prognostic marker for cancer progression and survival [107], and a target for colon cancer diagnostics and treatment [108]. It is conceivable that biglycan triggers tumorigenesis directly via TLR2/4-NF- κ B- and P2X₇-NLRP3-caspase-1-signaling or indirectly via downstream mediators such as pro-inflammatory cytokines, NOX enzymes, ROS, HIF-2 α , Epo and VEGF [30,77,83,85,86,89,95] (Figs. 2 and 3). Indeed, there is mounting evidence that activation of TLR2 and TLR4 leads to cancer progression due to increased tumor cell proliferation, resistance to apoptosis, increased production of growth factors (e.g. TGF- β , VEGF) and of pro-inflammatory cytokines [109–111]. Downstream of TLR2/4, the adaptor molecule MyD88 and its oncogenically-active mutant forms, are considered mediators of tumorigenesis in various cancers [112]. Beyond that, there is no doubt that constitutive activation of NF- κ B and MAPK pathways cause chronic inflammation which would favor cancer progression. The complex regulation of inflammation by biglycan is a result of signaling crosstalk between TLR2/4- and P2X₇-NLRP3-caspase-1-pathways [88,113,114]. While direct data on biglycan-NLRP3-caspase-1-signaling in carcinoma cells are not available yet, there is strong evidence for pro-tumorigenic effects of the NLRP3-caspase-1 signaling pathway [115,116]. Enhanced expression and activation of NLRP3 and caspase-1 is considered a poor prognostic sign for oncologic patients [115]. Therefore the development of inhibitors selectively targeting the NLRP3 inflammasome could be of potential therapeutic value.

Besides directly triggering pro-inflammatory TLR- and inflammasome-signaling, biglycan stimulates the generation of pro-inflammatory cytokines, NOX enzymes, ROS, HIF-2 α , Epo and VEGF, which are crucial mediators of inflammation and angiogenesis in cancer development. Accordingly, enhanced generation of NOX-derived ROS in inflammatory tumors causes chromosomal DNA alteration and genomic instability, as well as induces tumor cell proliferation, survival, and metastasis [117,118]. Furthermore, enhanced HIF-2 α , which might be due to biglycan-TLR2-dependent HIF-2 α stabilization [95], has been detected in a variety of tumors with diverse histogenetic background [119,120]. Interestingly, there is growing evidence that generation of HIF-2 α in cancer is restricted to

tumor-associated macrophages (TAMs) [119,121]. By regulating the expression of VEGF, VEGF receptors 1/2 and angiopoietins in TAMs, HIF-2 α induces tumor angiogenesis and triggers the expression of genes involved in tumor cell proliferation and tumor invasion [122–124]. On the other hand, biglycan has been described to trigger endothelial cell migration and neovascularization of tumors due to its ability to induce VEGF synthesis in a TLR2/4-dependent manner [32,77,111]. Additionally, biglycan can generate a reservoir for VEGF, which in turn can be released during tumor-associated ECM-degradation, thus promoting angiogenesis [125]. Moreover, soluble biglycan is expressed in highly metastatic tumor endothelial cells, where it promotes tumor cell migration [126]. Thus, it is conceivable that during cancer development enhanced biglycan levels could promote stabilization of TAM-derived HIF-2 α (Fig. 3) and trigger VEGF production and sequestration. Both bioactivities would ultimately trigger tumor angiogenesis.

During tumorigenesis a cross-talk between tumor and stromal cells modifies the microenvironment and the ECM [127]. The expression of biglycan by stromal fibroblasts is induced by TGF- β , a multi-purpose growth factor [128,129] which might then use the TLR2/4 on stromal macrophages to further modulate the tumor microenvironment and to induce the recruitment of tumor cells through chemokine production. In addition, biglycan is able to increase the affinity of TGF- β to its receptor and therefore promote cancer progression. Thus, we presume, that biglycan-triggered TGF- β signaling might create an environment, which is prone to tumor development, by triggering the expression of matrix proteins and enzymes involved in matrix remodeling [130,131]. Moreover, TGF- β activates fibroblast-to-myofibroblast differentiation [132,133], as well as epithelial-to-mesenchymal transition (EMT) [134] thus increasing the potential for metastasis. Furthermore, decorin interacts with TGF- β [62], to inhibit its signaling pathway, resulting in the reduction of fibrosis [135]. Interestingly, TGF- β inhibits decorin transcription [136], a finding opposite to the TGF- β -mediated induction of biglycan. These disparate functions of decorin and biglycan in the context of TGF- β signaling solidify the contrasting effects seen on tumorigenesis by these related proteoglycans, where one diminishes tumor progression while the other promotes disease.

Enhanced Epo production based on biglycan-TLR2-HIF-2 α signaling [95] provides an additional indirect mechanism of biglycan promoted tumorigenesis. Upon binding to erythropoietin receptors present on carcinoma cells, Epo promotes tumor growth, local invasion, metastasis, angiogenesis and lymphangiogenesis [137–140] (Fig. 3). Thus, there is a plethora of direct and indirect mechanisms through which biglycan promotes inflammation and angiogenesis resulting in tumor progression.

Besides strong indications for pro-oncogenic functions of biglycan, there are also some reports describing tumor suppressive effects of biglycan. For example, biglycan induces cell growth arrest in pancreatic cancer cell lines *in vitro* [78] and inhibits bladder cancer cell proliferation [141]. Interestingly, although elevated levels of biglycan correlate with high-grade human bladder cancer and muscle invasiveness, enhanced expression of tumor-associated biglycan is linked to a better survival rate [141]. Accordingly, knockdown of biglycan leads to increased proliferation of bladder cancer cells, indicating biglycan as an endogenous inhibitor of cancer cell growth in urothelial neoplasms [141]. In addition,

biglycan induces a high intratumoral inflammatory reaction as well as an enhanced autologous tumor response in diffuse large B-cell lymphomas, leading to improved therapeutic outcome and survival [142]. Presumably, this dual role of biglycan in tumorigenesis depends on the tumor cell type and differentiation stage (Fig. 2). According to our current knowledge, a biglycan-mediated chronic inflammatory milieu should promote tumor growth. However, acute inflammation caused by administration of biglycan into established tumors actually reduces malignant growth, analogous to beneficial effects of decorin-TLR2/4 signaling [143] (Fig. 2).

Taken together, the multifunctional involvement of biglycan in the regulation of inflammation makes it a potentially druggable target for cancer remedies as well as a novel anti-oncogenic therapeutic agent on its own.

Pro-inflammatory signaling of decorin

Analogous to biglycan, decorin induces a TLR2/4-dependent signaling cascade in macrophages leading to the activation of the MAPK, ERK, stress-activated protein kinase (SAPK) and p38 signaling pathways, and subsequently to the synthesis and secretion of CCL-2, TNF- α and IL-12p70 [143,144]. Moreover, decorin possesses the ability to further drive the cytokine profile toward a pro-inflammatory phenotype by impeding the production of the anti-inflammatory cytokine, IL-10, by macrophages. By sequestering active TGF- β , decorin reduces the abundance of TGF- β -driven oncogenic microRNA (miR)-21. miR-21 is a translational inhibitor of the tumor suppressor programmed cell death protein 4 (PDCD4), which itself post-translationally suppresses IL-10. Consequently, increased PDCD4 production causes reduction of IL-10 protein abundance [143]. Indeed, due to creating an inflammatory milieu in the tumor microenvironment together with reduction of the oncogene miR-21 and enhancement of the tumor suppressor PDCD4, decorin inhibits growth of established tumors [143]. Thus, decorin, as an important modulator of malignant growth, metastasis and tumor microenvironment [67,68,145–148] might be a useful tool for anti-tumor interventions.

Conclusions and perspectives

The functions of decorin in tumor biology continue to surface as more sophisticated research tools become available [149,150]. Though decorin has been touted as a “guardian from the matrix” for many years, this designation was primarily given due to its ability to alter RTK-signaling, resulting in reduction of tumor growth via cell cycle arrest. More recent work demonstrates the incredible versatility of this SLRP as decorin has now acquired a new role as a potent autophagic inducer in endothelial cells and a pro-mitophagic agent in tumor cells. The ability for decorin to regulate these signaling pathways offers a new perspective on matrix-regulation of both the tumor microenvironment as well as the tumor proper. Specifically, the capacity of decorin to both inhibit angiogenesis as well as act directly on the tumor proper makes it an ideal candidate for drug design. Furthermore, as decorin is a naturally-occurring proteoglycan, the side effects of using purified decorin or its biologically active fragments will likely be minimal, making it an attractive option for chemotherapy.

Moving to biglycan, research of the last two years has exponentially improved our knowledge regarding the role of this SLRP in inflammation and angiogenesis. There is now evidence that, besides mediating inflammation by biglycan, TLR2/4 are required for biglycan-dependent synthesis of VEGF and, therefore, regulation of angiogenesis. Furthermore, it became clear that biglycan is not only a pro-inflammatory stimulus but is also capable of self-limiting inflammation. The underlying mechanism of how biglycan tightly balances pro- and anti-inflammatory responses under physiological conditions has been provided [89]. Hopefully, these data will encourage further research on the mechanisms regarding anti-inflammatory signaling of other ECM-derived danger signals.

Beyond the well-known biglycan-dependent orchestration of various receptor signaling, another level of complexity in biglycan-dependent regulation of inflammation has recently been described [89]. It is now obvious that besides common signaling via TLR2 and TLR4, biglycan selectively binds to only one TLR. After choosing the receptor, the next step of signaling is achieved at the level of TLR adaptor molecules, finally resulting in a very specific downstream outcome. This might explain why, in certain cell types and under certain conditions, biglycan could actually suppress tumorigenesis. The mechanisms of how this selection of receptors and adaptors is regulated are not known yet. It is conceivable, however, that various co-receptors might help biglycan to “make that specific choice”.

One example for a specific outcome of biglycan-TLR2 signaling is the newly-identified function of biglycan to stabilize HIF-2 α and increase Epo production [95]. The exact mechanisms for biglycan-mediated HIF-2 α stabilization still need to be elucidated. Although little is known about the biglycan-HIF-2 α -Epo signaling axis during tumorigenesis, it is likely that the blockade of biglycan/TLR2-binding could protect against malignant growth. However, given that TLRs have crucial physiological functions, future studies are essential to unravel co-receptors and downstream mechanisms of biglycan/TLR-signaling for selective druggability in the context of cancer.

In conclusion, both decorin and biglycan are important mediators of tumor initiation and progression involving the cellular pathways of angiogenesis, inflammation, and autophagy. As the tumor microenvironment is a complex amalgam of different cell types and signaling molecules, it is probable that these critical cellular processes are intertwined in an intricate system to alter cell growth, metastasis, and nutrient availability for both normal and cancer cells. Therefore, it is important to focus on these pathways and their regulation by these and other matrix constituents in order to gain a closer understanding of the timeline of events involved in tumor initiation as well as to define which pathways are deregulated leading to tumor progression. Future therapeutic modalities will materialize as the nuances of these signaling pathways are elucidated, and thus it is essential to continue our efforts of unravelling the role of these proteoglycans in normal and aberrant physiology.

Acknowledgments

The original research in the authors' laboratory was supported by the German Research Council (SFB 815, project A5, SFB 1039, project B2, SFB 1177, project C2, SCHA 1082/6-1 all for LS) and LOEWE program Ub-Net (LS), and by the National Institutes of Health grants RO1 CA39481, RO1 CA47282, and RO1 CA164462 (to RVI). M.A. Gubbiotti was supported in part by NIH training grant T32 AA07463.

Abbreviations

AMPK	adenosine monophosphate kinase
CCL	chemokine (C–C motif) ligand
CS/DS	chondroitin sulfate/dermatan sulfate
CXCL	chemokine (C-X-C motif) ligand
ECM	ECM
EGFR	epidermal growth factor receptor
Epo	erythropoietin
ERK	extracellular signal-regulated kinases
GAG	glycosaminoglycan
HGF/Met	hepatocyte growth factor
HIF	hypoxia inducible factor
HSP	heat shock protein
HSPG2	heparan sulfate proteoglycan 2
IL	interleukin
LC3	microtubule-associated protein light chain 3
LRP6	LDL-receptor related protein 6
MyD88	Myeloid differentiation primary response protein
NOX	NADPH oxidase
NF-κB	nuclear factor kappa-light-chain-enhancer of activated B cells
NLRP3	NOD-like receptor protein 3
PDGF	platelet-derived growth factor
Peg3	paternally expressed gene 3
ROS	reactive oxygen species
SLRP	small, leucine-rich proteoglycan
TAM	tumor associated macrophages
TLR	Toll-like receptor
TGF-β	transforming growth factor β
TRIF	Toll/IL-1R domain-containing adaptor inducing IFN- β

VEGF vascular endothelial growth factor

VEGFR2 vascular endothelial growth factor receptor 2

References

1. Iozzo RV, Schaefer L. Proteoglycan form and function: A comprehensive nomenclature of proteoglycans. *Matrix Biol.* 2015; 42:11–55. [PubMed: 25701227]
2. Sanderson RD, Elkin M, Rapraeger AC, Ilan N, Vlodavsky I. Heparanase regulation of cancer, autophagy and inflammation: New mechanisms and targets for therapy. *FEBS J.* 2016
3. Afratis N, Mulhaupt HAB, Nikitovic D, Theocharis AD, Couchman JR, Karamanos NK. Syndecans: key regulators of cell signaling and biological functions. *FEBS J.* 2016 In press.
4. Järveläinen H, Puolakkainen P, Pakkanen S, Brown EL, Höök M, Iozzo RV, Sage H, Wight TN. A role for decorin in cutaneous wound healing and angiogenesis. *Wound Rep Reg.* 2006; 14:443–452.
5. Keene DR, San Antonio JD, Mayne R, McQuillan DJ, Sarris G, Santoro SA, Iozzo RV. Decorin binds near the C terminus of type I collagen. *J Biol Chem.* 2000; 275:21801–21804. [PubMed: 10823816]
6. Reed CC, Iozzo RV. The role of decorin in collagen fibrillogenesis and skin homeostasis. *Glycoconj J.* 2002; 19:249–255. [PubMed: 12975602]
7. Rühland C, Schönherr E, Robenek H, Hansen U, Iozzo RV, Bruckner P, Seidler DG. The glycosaminoglycan chain of decorin plays an important role in collagen fibril formation at the early stages of fibrillogenesis. *FEBS J.* 2007; 274:4246–4255. [PubMed: 17651433]
8. Sanches JCT, Jones CJP, Aplin JD, Iozzo RV, Zorn TMT, Oliveira SF. Collagen fibril organization in the pregnant endometrium of decorin-deficient mice. *J Anat.* 2010; 216:144–155. [PubMed: 19900179]
9. Weber IT, Harrison RW, Iozzo RV. Model structure of decorin and implications for collagen fibrillogenesis. *J Biol Chem.* 1996; 271:31767–31770. [PubMed: 8943211]
10. Järveläinen H, Sainio A, Wight TN. Pivotal role for decorin in angiogenesis. *Matrix Biol.* 2015; 43:15–26. [PubMed: 25661523]
11. Neill T, Painter H, Buraschi S, Owens RT, Lisanti MP, Schaefer L, Iozzo RV. Decorin antagonizes the angiogenic network. Concurrent inhibition of Met, hypoxia inducible factor-1 α and vascular endothelial growth factor A and induction of thrombospondin-1 and TIMP3. *J Biol Chem.* 2012; 287:5492–5506. [PubMed: 22194599]
12. Gubbiotti MA, Neill T, Frey H, Schaefer L, Iozzo RV. Decorin is an autophagy-inducible proteoglycan and is required for proper in vivo autophagy. *Matrix Biol.* 2015; 48:14–25. [PubMed: 26344480]
13. Neill T, Torres AT, Buraschi S, Iozzo RV. Decorin has an appetite for endothelial cell autophagy. *Autophagy.* 2013; 9:1626–1628. [PubMed: 23989617]
14. Bocian C, Urbanowitz AK, Owens RT, Iozzo RV, Gotte M, Seidler DG. Decorin potentiates interferon-gamma activity in a model of allergic inflammation. *J Biol Chem.* 2013; 288:12699–12711. [PubMed: 23460644]
15. Borges MC, Narayanan V, Iozzo RV, Ludwig MS. Deficiency of decorin induces expression of Foxp3 in CD4(+) CD25(+) T cells in a murine model of allergic asthma. *Respirology.* 2015; 20:904–911. [PubMed: 25712878]
16. Gubbiotti MA, Iozzo RV. Proteoglycans regulate autophagy via outside-in signaling: An emerging new concept. *Matrix Biol.* 2015; 48:6–13. [PubMed: 26462577]
17. Buraschi S, Neill T, Goyal A, Poluzzi C, Smythies J, Owens RT, Schaefer L, Torres A, Iozzo RV. Decorin causes autophagy in endothelial cells via Peg3. *Proc Natl Acad Sci U S A.* 2013; 110:E2582–E2591. [PubMed: 23798385]
18. Goyal A, Neill T, Owens RT, Schaefer L, Iozzo RV. Decorin activates AMPK, an energy sensor kinase, to induce autophagy in endothelial cells. *Matrix Biol.* 2014; 34:46–54. [PubMed: 24472739]

19. Mimura T, Han KY, Onguchi T, Chang J-H, Kim T-I, Kojima T, Zhou Z, Azar DT. MT1-MMP-mediated cleavage of decorin in corneal angiogenesis. *J Vasc Res.* 2008; 46:541–550.
20. Neill T, Schaefer L, Iozzo RV. Decorin as a multivalent therapeutic agent against cancer. *Adv Drug Deliv Rev.* 2016; 97:174–185. [PubMed: 26522384]
21. Scilabra SD, Yamamoto K, Pigoni M, Sakamoto K, Muller SA, Papadopoulou A, Lichtenthaler SF, Troeberg L, Nagase H, Kadomatsu K. Dissecting the interaction between tissue inhibitor of metalloproteinases-3 (TIMP-3) and low density lipoprotein receptor-related protein-1 (LRP-1): Development of a “TRAP” to increase levels of TIMP-3 in the tissue. *Matrix Biol.* 2016 In Press.
22. Belotti D, Capelli C, Resovi A, Introna M, Taraboletti G. Thrombospondin-1 promotes mesenchymal stromal cell functions via TGF β and in cooperation with PDGF. *Matrix Biol.* 2016 In Press.
23. Buraschi S, Neill T, Owens RT, Iniguez LA, Purkins G, Vadigepalli R, Evans B, Schaefer L, Peiper SC, Wang Z, Iozzo RV. Decorin protein core affects the global gene expression profile of the tumor microenvironment in a triple-negative orthotopic breast carcinoma xenograft model. *PLoS ONE.* 2012; 7:e45559. [PubMed: 23029096]
24. Choi AMK, Ryter SW, Levine B. Autophagy in human health and disease. *New Engl J Med.* 2013; 368:651–662. [PubMed: 23406030]
25. Levine B, Kroemer G. Autophagy in the pathogenesis of disease. *Cell.* 2008; 132:27–42. [PubMed: 18191218]
26. Lala N, Gannareddy VG, Cloutier-Bosworth A, Lala PK. Mechanisms in decorin regulation of vascular endothelial growth factor-induced human trophoblast migration and acquisition of endothelial phenotype. *Biol Reprod.* 2012; 87:59, 1–14. [PubMed: 22699486]
27. Kalas W, Swiderek E, Switalska M, Wietrzyk J, Rak J, Strzadala L. Thrombospondin-1 receptor mediates autophagy of RAS-expressing cancer cells and triggers tumour growth inhibition. *AntiCancer Res.* 2013; 33:1429–1438. [PubMed: 23564783]
28. Soto-Pantoja DR, Miller TW, Pendrak ML, DeGraff WG, Sullivan C, Ridnour LA, Abu-Asab M, Wink DA, Toskos M, Roberts DD. CD47 deficiency confers cell and tissue radioprotection by activation of autophagy. *Autophagy.* 2012; 8:1628–1642. [PubMed: 22874555]
29. Santiago-García J, Kodama T, Pitas RE. The class A scavenger receptor binds to proteoglycans and mediates adhesion of macrophages to the extracellular matrix. *J Biol Chem.* 2003; 278:6942–6946. [PubMed: 12488451]
30. Myren M, Kirby DJ, Noonan ML, Maeda A, Owens RT, Ricard-Blum S, Kram V, Kilts TM, Young MF. Biglycan potentially regulates angiogenesis during fracture repair by altering expression and function of endostatin. *Matrix Biol.* 2016; 52–54:141–150.
31. Berendsen AD, Pinnow EL, Maeda A, Brown AC, McCartney-Francis N, Kram V, Owens RT, Robey PG, Holmbeck K, de Castro LF, Kilts TM, Young MF. Biglycan modulates angiogenesis and bone formation during fracture healing. *Matrix Biol.* 2014; 35:223–231. [PubMed: 24373744]
32. Xing X, Gu X, Ma T, Ye H. Biglycan up-regulated vascular endothelial growth factor (VEGF) expression and promoted angiogenesis in colon cancer. *Tumour Biol.* 2015; 36:1773–1780. [PubMed: 25371074]
33. Cohen IR, Murdoch AD, Naso MF, Marchetti D, Berd D, Iozzo RV. Abnormal expression of perlecan proteoglycan in metastatic melanomas. *Cancer Res.* 1994; 54:5771–5774. [PubMed: 7954396]
34. Mathiak M, Yenisey C, Grant DS, Sharma B, Iozzo RV. A role for perlecan in the suppression of growth and invasion in fibrosarcoma cells. *Cancer Res.* 1997; 57:2130–2136. [PubMed: 9187109]
35. Mongiat M, Sweeney S, San Antonio JD, Fu J, Iozzo RV. Endorepellin, a novel inhibitor of angiogenesis derived from the C terminus of perlecan. *J Biol Chem.* 2003; 278:4238–4249. [PubMed: 12435733]
36. Iozzo RV, San Antonio JD. Heparan sulfate proteoglycans: heavy hitters in the angiogenesis arena. *J Clin Invest.* 2001; 108:349–355. [PubMed: 11489925]
37. Lord MS, Chuang CY, Melrose J, Davies MJ, Iozzo RV, Whitelock JM. The role of vascular-derived perlecan in modulating cell adhesion, proliferation and growth factor signaling. *Matrix Biol.* 2014; 35:112–122. [PubMed: 24509440]

38. Iozzo RV, Zoeller JJ, Nyström A. Basement membrane proteoglycans: Modulators *par excellence* of cancer growth and angiogenesis. *Mol Cells*. 2009; 27:503–513. [PubMed: 19466598]
39. Zoeller JJ, Whitelock J, Iozzo RV. Perlecan regulates developmental angiogenesis by modulating the VEGF-VEGFR2 axis. *Matrix Biol*. 2009; 28:284–291. [PubMed: 19422911]
40. Iozzo RV, Sanderson RD. Proteoglycans in cancer biology, tumour microenvironment and angiogenesis. *J Cell Mol Med*. 2011; 15:1013–1031. [PubMed: 21155971]
41. Grindel BJ, Martinez JR, Pennington CL, Muldoon M, Stave J, Chung LW, Farach-Carson MC. Matrilysin/matrix metalloproteinase-7(MMP7) cleavage of perlecan/HSPG2 creates a molecular switch to alter prostate cancer cell behavior. *Matrix Biol*. 2014; 36:64–76. [PubMed: 24833109]
42. Qiang B, Lim SY, Lekas M, Kuliszewski MA, Wolff R, Osherov AB, Rudenko D, Leong-Poi H, Noyan H, Husain M, Tran K, Tryggvason K, Hedin U, Tran-Lundmark K, Strauss BH. Perlecan heparan sulfate proteoglycan is a critical determinant of angiogenesis in response to mouse hind-limb ischemia. *Can J Cardiol*. 2014; 30:1444–1451. [PubMed: 25249499]
43. Poluzzi C, Iozzo RV, Schaefer L. Endostatin and endorepellin: A common route of action for similar angiostatic cancer avengers. *Adv Drug Deliv Rev*. 2016; 97:156–173. [PubMed: 26518982]
44. McCarthy KJ. The basement membrane proteoglycans perlecan and agrin: Something old, something new. *Curr Top Membr*. 2015; 76:255–303. [PubMed: 26610917]
45. Theocharis AD, Gialeli C, Bouris P, Giannopoulou E, Skandalis SS, Aletas AJ, Iozzo RV, Karamanos NK. Cell-matrix interactions: focus on proteoglycan-proteinase interplay and pharmacological targeting in cancer. *FEBS J*. 2014; 281:5023–5042. [PubMed: 25333340]
46. Woodall BP, Nyström A, Iozzo RA, Eble JA, Niland S, Krieg T, Eckes B, Pozzi A, Iozzo RV. Integrin $\alpha 2 \beta 1$ is the required receptor for endorepellin angiostatic activity. *J Biol Chem*. 2008; 283:2335–2343. [PubMed: 18024432]
47. Douglass S, Goyal A, Iozzo RV. The role of perlecan and endorepellin in the control of tumor angiogenesis and endothelial cell autophagy. *Connect Tissue Res*. 2015; 19:1–11.
48. Mongiat M, Taylor K, Otto J, Aho S, Uitto J, Whitelock J, Iozzo RV. The protein core of the proteoglycan perlecan binds specifically to fibroblast growth factor-7. *J Biol Chem*. 2000; 275:7095–7100. [PubMed: 10702276]
49. Mongiat M, Otto J, Oldershaw R, Ferrer F, Sato JD, Iozzo RV. Fibroblast growth factor-binding protein is a novel partner for perlecan protein core. *J Biol Chem*. 2001; 276:10263–10271. [PubMed: 11148217]
50. Smith SML, West LA, Govindraj P, Zhang X, Ornitz DM, Hassell JR. Heparan and chondroitin sulfate on growth plate perlecan mediate binding and delivery of FGF-2 to FGF receptors. *Matrix Biol*. 2007; 26:175–184. [PubMed: 17169545]
51. Muthusamy A, Cooper CR, Gomes RR Jr. Soluble perlecan domain I enhances vascular endothelial growth factor-165 activity and receptor phosphorylation in human bone marrow endothelial cells. *BMC Biochemistry*. 2010; 11:43. [PubMed: 21047416]
52. Rossi M, Morita H, Sormunen R, Airene S, Kreivi M, Wang L, Fukai N, Olsen BR, Tryggvason K, Soininen R. Heparan sulfate chains of perlecan are indispensable in the lens capsule but not in the kidney. *EMBO J*. 2003; 22:236–245. [PubMed: 12514129]
53. Zhou Z, Wang J, Cao R, Morita H, Soininen R, Chan KM, Liu B, Cao Y, Tryggvason K. Impaired angiogenesis, delayed wound healing and retarded tumor growth in perlecan heparan sulfate-deficient mice. *Cancer Res*. 2004; 64:4699–4702. [PubMed: 15256433]
54. Ning L, Xu Z, Furuya N, Nonaka R, Yamada Y, Arikawa-Hirasawa E. Perlecan inhibits autophagy to maintain muscle homeostasis in mouse soleus muscle. *Matrix Biol*. 2015; 48:26–35. [PubMed: 26319110]
55. Gubbiotti MA, Neill T, Iozzo RV. A current view of perlecan in physiology and pathology: A mosaic of functions. *Matrix Biol*. 2016 In Press.
56. Bix G, Castello R, Burrows M, Zoeller JJ, Weech M, Iozzo RA, Cardi C, Thakur MT, Barker CA, Camphausen KC, Iozzo RV. Endorepellin in vivo: targeting the tumor vasculature and retarding cancer growth and metabolism. *J Natl Cancer Inst*. 2006; 98:1634–1646. [PubMed: 17105986]
57. Poluzzi C, Casulli J, Goyal A, Mercer TJ, Neill T, Iozzo RV. Endorepellin evokes autophagy in endothelial cells. *J Biol Chem*. 2014; 289:16114–16128. [PubMed: 24737315]

58. Goyal A, Gubbiotti MA, Chery DR, Han L, Iozzo RV. Endorepellin-evoked autophagy contributes to angiostasis. *J Biol Chem.* 2016 In press.
59. Goldoni S, Owens RT, McQuillan DJ, Shriver Z, Sasisekharan R, Birk DE, Campbell S, Iozzo RV. Biologically active decorin is a monomer in solution. *J Biol Chem.* 2004; 279:6606–6612. [PubMed: 14660661]
60. Neill T, Schaefer L, Iozzo RV. Decoding the matrix: Instructive roles of proteoglycan receptors. *Biochemistry.* 2015; 54:4583–4598. [PubMed: 26177309]
61. Neill T, Schaefer L, Iozzo RV. Decorin, a guardian from the matrix. *Am J Pathol.* 2012; 181:380–387. [PubMed: 22735579]
62. Gubbiotti MA, Vallet SD, Ricard-Blum S, Iozzo RV. Decorin interacting network: A comprehensive analysis of decorin-binding partners and their versatile functions. *Matrix Biol.* 2016; 55:7–21. [PubMed: 27693454]
63. Moscatello DK, Santra M, Mann DM, McQuillan DJ, Wong AJ, Iozzo RV. Decorin suppresses tumor cell growth by activating the epidermal growth factor receptor. *J Clin Invest.* 1998; 101:406–412. [PubMed: 9435313]
64. Iozzo RV, Moscatello D, McQuillan DJ, Eichstetter I. Decorin is a biological ligand for the epidermal growth factor receptor. *J Biol Chem.* 1999; 274:4489–4492. [PubMed: 9988678]
65. Santra M, Reed CC, Iozzo RV. Decorin binds to a narrow region of the epidermal growth factor (EGF) receptor, partially overlapping with but distinct from the EGF-binding epitope. *J Biol Chem.* 2002; 277:35671–35681. [PubMed: 12105206]
66. Zhu J-X, Goldoni S, Bix G, Owens RA, McQuillan D, Reed CC, Iozzo RV. Decorin evokes protracted internalization and degradation of the EGF receptor via caveolar endocytosis. *J Biol Chem.* 2005; 280:32468–32479. [PubMed: 15994311]
67. Goldoni S, Seidler DG, Heath J, Fassan M, Baffa R, Thakur ML, Owens RA, McQuillan DJ, Iozzo RV. An anti-metastatic role for decorin in breast cancer. *Am J Pathol.* 2008; 173:844–855. [PubMed: 18688028]
68. Goldoni S, Iozzo RV. Tumor microenvironment: Modulation by decorin and related molecules harboring leucine-rich tandem motifs. *Int J Cancer.* 2008; 123:2473–2479. [PubMed: 18798267]
69. Baghy K, Horváth Z, Regős E, Kiss K, Schaff Z, Iozzo RV, Kovalszky I. Decorin interferes with platelet-derived growth factor receptor signaling in experimental hepatocarcinogenesis. *FEBS J.* 2013; 280:2150–2164. [PubMed: 23448253]
70. Goldoni S, Humphries A, Nyström A, Sattar S, Owens RT, McQuillan DJ, Ireton K, Iozzo RV. Decorin is a novel antagonistic ligand of the Met receptor. *J Cell Biol.* 2009; 185:743–754. [PubMed: 19433454]
71. Buraschi S, Pal N, Tyler-Rubinstein N, Owens RT, Neill T, Iozzo RV. Decorin antagonizes Met receptor activity and downregulates β -catenin and Myc levels. *J Biol Chem.* 2010; 285:42075–42085. [PubMed: 20974860]
72. Neill T, Torres A, Buraschi S, Owens RT, Hoek J, Baffa R, Iozzo RV. Decorin induces mitophagy in breast carcinoma cells via peroxisome proliferator-activated receptor γ coactivator-1 α (PGC-1 α) and mitostatin. *J Biol Chem.* 2014; 289:4952–4968. [PubMed: 24403067]
73. Yao T, Zhang CG, Gong MT, Zhang M, Wang L, Ding W. Decorin-mediated inhibition of the migration of U87MG glioma cells involves activation of autophagy and suppression of TGF- β signaling. *FEBS Open Bio.* 2016; 6:707–719.
74. Wei Y, Zou Z, Becker N, Anderson M, Sumpter R, Xiao G, Kinch L, Koduru P, Christudass CS, Veltri RW, Grishin NV, Peyton M, Minna J, Bhagat G, Levine B. EGFR-mediated beclin 1 phosphorylation in autophagy suppression, tumor progression, and tumor chemoresistance. *Cell.* 2013; 154:1269–1284. [PubMed: 24034250]
75. Feng W, Marquez RT, Lu Z, Liu J, Lu KH, Issa J-PJ, Fishman DM, Yu Y, Bast RC. Imprinted tumor suppressor genes *ARHI* and *PEG3* are the most frequently down-regulated in human ovarian cancers by loss of heterozygosity and promoter methylation. *Cancer.* 2008; 112:1489–1502. [PubMed: 18286529]
76. Kohda T, Asai A, Kuroiwa Y, Kobayashi S, Aisaka K, Nagashima G, Yoshida MC, Kondo Y, Kagiya N, Kirino T, Kaneko-Ishino T, Ishino F. Tumour suppressor activity of human imprinted gene *PEG3* in a glioma cell line. *Genes Cells.* 2001; 6:237–247. [PubMed: 11260267]

77. Yamamoto K, Ohga N, Hida Y, Maishi N, Kawamoto T, Kitayama K, Akiyama K, Osawa t, Kondoh M, Matsuda K, Onodera Y, Fujie M, Kaga K, Hirano S, Shinohara N, Shinodoh M, Hida K. Biglycan is a specific marker and an autocrine angiogenic factor of tumour endothelial cells. *Brit J Cancer*. 2012; 106:1214–1223. [PubMed: 22374465]
78. Weber CK, Sommer G, Michl P, Fensterer H, Weimer M, Gansauge F, Leder G, Adler G, Gress TA. Biglycan is overexpressed in pancreatic cancer and induces G1-arrest in pancreatic cancer cell lines. *Gastroenterology*. 2001; 121:657–667. [PubMed: 11522750]
79. Gu X, Ma Y, Xiao J, Zheng H, Song C, Gong Y, Xing X. Up-regulated biglycan expression correlates with the malignancy in human colorectal cancers. *Clin Exp Med*. 2012; 12:195–199. [PubMed: 21879307]
80. Hu L, Duan YT, Li JF, Su LP, Yan M, Zhu ZG, Liu BY, Yang QM. Biglycan enhances gastric cancer invasion by activating FAK signaling pathway. *Oncotarget*. 2014; 5:1885–1896. [PubMed: 24681892]
81. Berendsen AD, Fisher LW, Kilts TM, Owens RT, Robey PG, Gutkind JS, Young MF. Modulation of canonical Wnt signaling by the extracellular matrix component biglycan. *Proc Natl Acad Sci USA*. 2011; 108:17022–17027. [PubMed: 21969569]
82. Gao C, Cao W, Bao L, Zuo W, Xie G, Cai T, Fu W, Zhang J, Wu W, Zhang X, Chen YG. Autophagy negatively regulates Wnt signalling by promoting Dishevelled degradation. *Nat Cell Biol*. 2010; 12:781–790. [PubMed: 20639871]
83. Schaefer L, Babelova A, Kiss E, Hausser H-J, Baliova M, Krzyzankova M, Marsche G, Young MF, Mihalik D, Götte M, Malle E, Schaefer RM, Gröne H-J. The matrix component biglycan is proinflammatory and signals through toll-like receptors 4 and 2 in macrophages. *J Clin Invest*. 2005; 115:2223–2233. [PubMed: 16025156]
84. Moreth K, Frey H, Hubo M, Zeng-Brouwers J, Nastase MV, Hsieh LT, Hacen R, Pfeilschifter J, Iozzo RV, Schaefer L. Biglycan-triggered TLR-2- and TLR-4-signaling exacerbates the pathophysiology of ischemic acute kidney injury. *Matrix Biol*. 2014; 35:143–151. [PubMed: 24480070]
85. Babelova A, Moreth K, Tsalastra-Greul W, Zeng-Brouwers J, Eickelberg O, Young MF, Bruckner P, Pfeilschifter J, Schaefer RM, Gröne H-J, Schaefer L. Biglycan, a danger signal that activates the NLRP3 inflammasome via Toll-like and P2X receptors. *J Biol Chem*. 2009; 284:24035–24048. [PubMed: 19605353]
86. Moreth K, Brodbeck R, Babelova A, Gretz N, Spieker T, Zeng-Brouwers J, Pfeilschifter J, Young MF, Schaefer RM, Schaefer L. The proteoglycan biglycan regulates expression of the B cell chemoattractant CXCL13 and aggravates murine lupus nephritis. *J Clin Invest*. 2010; 120:4251–4272. [PubMed: 21084753]
87. Zeng-Brouwers J, Beckmann J, Nastase MV, Iozzo RV, Schaefer L. De novo expression of circulating biglycan evokes an innate inflammatory tissue response via MyD88/TRIF pathways. *Matrix Biol*. 2014; 35:132–142. [PubMed: 24361484]
88. Frey T, Schroeder N, Manon-Jensen T, Iozzo RV, Schaefer L. Biological interplay between proteoglycans and their innate immune receptors in inflammation. *FEBS J*. 2013; 280:2165–2179. [PubMed: 23350913]
89. Hsieh LT, Frey H, Nastase MV, Tredup C, Hoffmann A, Poluzzi C, Zeng-Brouwers J, Manon-Jensen T, Schroder K, Brandes RP, Iozzo RV, Schaefer L. Bimodal role of NADPH oxidases in the regulation of biglycan-triggered IL-1 β synthesis. *Matrix Biol*. 2016; 49:61–81. [PubMed: 26689330]
90. Schaefer L. Complexity of danger: the diverse nature of damage-associated molecular patterns. *J Biol Chem*. 2014; 289:35237–35245. [PubMed: 25391648]
91. Muir AM, Massoudi D, Nguyen N, Keene DR, Lee SJ, Birk DE, Davidson JM, Marinkovich MP, Greenspan DS. BMP1-like proteinases are essential to the structure and wound healing of skin. *Matrix Biol*. 2016 In Press.
92. Eckhard U, Huesgen PF, Schilling O, Bellac CL, Butler GS, Cox JH, Dufour A, Goebeler V, Kappelhoff R, Keller UA, Klein T, Lange PF, Marino G, Morrison CJ, Prudova A, Rodriguez D, Starr AE, Wang Y, Overall CM. Active site specificity profiling of the matrix metalloproteinase family: Proteomic identification of 4300 cleavage sites by nine MMPs explored with structural and synthetic peptide cleavage analyses. *Matrix Biol*. 2016; 49:37–60. [PubMed: 26407638]

93. Hsieh LT, Nastase MV, Zeng-Brouwers J, Iozzo RV, Schaefer L. Soluble biglycan as a biomarker of inflammatory renal diseases. *Int J Biochem Cell Biol.* 2014; 54C:223–235.
94. Nastase MV, Iozzo RV, Schaefer L. Key roles for the small leucine-rich proteoglycans in renal and pulmonary pathophysiology. *Biochim Biophys Acta.* 2014; 1840:2460–2470. [PubMed: 24508120]
95. Frey H, Moreth K, Hsieh LT, Zeng-Brouwers J, Rathkolb B, Fuchs H, Gailus-Durner V, Iozzo RV, de Angelis MH, Schaefer L. A novel biological function of soluble biglycan: Induction of erythropoietin production and polycythemia. *Glycoconj J.* 2016
96. Imtiyaz HZ, Williams EP, Hickey MM, Patel SA, Durham AC, Yuan LJ, Hammond R, Gimotty PA, Keith B, Simon MC. Hypoxia-inducible factor 2 α regulates macrophage function in mouse models of acute and tumor inflammation. *J Clin Invest.* 2010; 120:2699–2714. [PubMed: 20644254]
97. Dunn GP, Old LJ, Schreiber RD. The immunobiology of cancer immunosurveillance and immunoediting. *Immunity.* 2004; 21:137–148. [PubMed: 15308095]
98. Pesic M, Greten FR. Inflammation and cancer: tissue regeneration gone awry. *Curr Opin Cell Biol.* 2016; 43:55–61. [PubMed: 27521599]
99. Mittal M, Siddiqui MR, Tran K, Reddy SP, Malik AB. Reactive oxygen species in inflammation and tissue injury. *Antioxid Redox Signal.* 2014; 20:1126–1167. [PubMed: 23991888]
100. Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell.* 2010; 140:883–899. [PubMed: 20303878]
101. Aprile G, Avellini C, Reni M, Mazzer M, Foltran L, Rossi D, Cereda S, Iaiza E, Fasola G, Piga A. Biglycan expression and clinical outcome in patients with pancreatic adenocarcinoma. *Tumour Biol.* 2013; 34:131–137. [PubMed: 23007878]
102. Mikula M, Rubel T, Karczmariski J, Goryca K, Dadlez M, Ostrowski J. Integrating proteomic and transcriptomic high-throughput surveys for search of new biomarkers of colon tumors. *Funct Integr Genomics.* 2011; 11:215–224. [PubMed: 21061036]
103. Pan S, Cheng L, White JT, Lu W, Utleg AG, Yan X, Urban ND, Drescher CW, Hood L, Lin B. Quantitative proteomics analysis integrated with microarray data reveals that extracellular matrix proteins, catenins, and p53 binding protein 1 are important for chemotherapy response in ovarian cancers. *OMICS.* 2009; 13:345–354. [PubMed: 19422301]
104. Nishino R, Honda M, Yamashita T, Takatori H, Minato H, Zen Y, Sasaki M, Takamura H, Horimoto K, Ohta T, Nakanuma Y, Kaneko S. Identification of novel candidate tumour marker genes for intrahepatic cholangiocarcinoma. *J Hepatol.* 2008; 49:207–216. [PubMed: 18490072]
105. Zhu YH, Yang F, Zhang SS, Zeng TT, Xie X, Guan XY. High expression of biglycan is associated with poor prognosis in patients with esophageal squamous cell carcinoma. *Int J Clin Exp Pathol.* 2013; 6:2497–2505. [PubMed: 24228112]
106. Jaeger J, Koczan D, Thiesen HJ, Ibrahim SM, Gross G, Spang R, Kunz M. Gene expression signatures for tumor progression, tumor subtype, and tumor thickness in laser-microdissected melanoma tissues. *Clin Cancer Res.* 2007; 13:806–815. [PubMed: 17289871]
107. Theocharis AD, Skandalis SS, Neill T, Mulhaupt HA, Hubo M, Frey H, Gopal S, Gomes A, Afratis N, Lim HC, Couchman JR, Filmus J, Ralph DS, Schaefer L, Iozzo RV, Karamanos NK. Insights into the key roles of proteoglycans in breast cancer biology and translational medicine. *Biochim Biophys Acta.* 2015; 1855:276–300. [PubMed: 25829250]
108. Suhovskih AV, Aidagulova SV, Kashuba VI, Grigorieva EV. Proteoglycans as potential microenvironmental biomarkers for colon cancer. *Cell Tissue Res.* 2015; 361:833–844. [PubMed: 25715761]
109. Farnebo L, Shahangian A, Lee Y, Shin JH, Scheeren FA, Sunwoo JB. Targeting Toll-like receptor 2 inhibits growth of head and neck squamous cell carcinoma. *Oncotarget.* 2015; 6:9897–9907. [PubMed: 25846753]
110. Dajon M, Iribarren K, Cremer I. Toll-like receptor stimulation in cancer: A pro- and anti-tumor double-edged sword. *Immunobiology.* 2016
111. Hu L, Zang MD, Wang HX, Li JF, Su LP, Yan M, Li C, Yang QM, Liu BY, Zhu ZG. Biglycan stimulates VEGF expression in endothelial cells by activating the TLR signaling pathway. *Mol Oncol.* 2016

112. Ngo VN, Young RM, Schmitz R, Jhavar S, Xiao W, Lim KH, Kohlhammer H, Xu W, Yang Y, Zhao H, Shaffer AL, Romesser P, Wright G, Powell J, Rosenwald A, Muller-Hermelink HK, Ott G, Gascoyne RD, Connors JM, Rimsza LM, Campo E, Jaffe ES, Delabie J, Smeland EB, Fisher RI, Braziel RM, Tubbs RR, Cook JR, Weisenburger DD, Chan WC, Staudt LM. Oncogenically active MYD88 mutations in human lymphoma. *Nature*. 2011; 470:115–119. [PubMed: 21179087]
113. Schaefer L, Iozzo RV. Small leucine-rich proteoglycans, at the crossroad of cancer growth and inflammation. *Curr Opin Genet Dev*. 2012; 22:56–57. [PubMed: 22326829]
114. Moreth K, Iozzo RV, Schaefer L. Small leucine-rich proteoglycans orchestrate receptor crosstalk during inflammation. *Cell Cycle*. 2012; 11:2084–2091. [PubMed: 22580469]
115. Dinarello CA. Interleukin-1beta and the autoinflammatory diseases. *N Engl J Med*. 2009; 360:2467–2470. [PubMed: 19494224]
116. Chow MT, Sceneay J, Paget C, Wong CS, Duret H, Tschopp J, Moller A, Smyth MJ. NLRP3 suppresses NK cell-mediated responses to carcinogen-induced tumors and metastases. *Cancer Res*. 2012; 72:5721–5732. [PubMed: 22986739]
117. Chiera F, Meccia E, Degan P, Aquilina G, Pietraforte D, Minetti M, Lambeth D, Bignami M. Overexpression of human NOX1 complex induces genome instability in mammalian cells. *Free Radic Biol Med*. 2008; 44:332–342. [PubMed: 17963706]
118. Wu Y, Antony S, Meitzler JL, Doroshow JH. Molecular mechanisms underlying chronic inflammation-associated cancers. *Cancer Lett*. 2014; 345:164–173. [PubMed: 23988267]
119. Talks KL, Turley H, Gatter KC, Maxwell PH, Pugh CW, Ratcliffe PJ, Harris AL. The expression and distribution of the hypoxia-inducible factors HIF-1alpha and HIF-2alpha in normal human tissues, cancers, and tumor-associated macrophages. *Am J Pathol*. 2000; 157:411–421. [PubMed: 10934146]
120. Bangoura G, Liu ZS, Qian Q, Jiang CQ, Yang GF, Jing S. Prognostic significance of HIF-2alpha/EPAS1 expression in hepatocellular carcinoma. *World J Gastroenterol*. 2007; 13:3176–3182. [PubMed: 17589895]
121. Onita T, Ji PG, Xuan JW, Sakai H, Kanetake H, Maxwell PH, Fong GH, Gabril MY, Moussa M, Chin JL. Hypoxia-induced, perinecrotic expression of endothelial Per-ARNT-Sim domain protein-1/hypoxia-inducible factor-2alpha correlates with tumor progression, vascularization, and focal macrophage infiltration in bladder cancer. *Clin Cancer Res*. 2002; 8:471–480. [PubMed: 11839666]
122. Skuli N, Liu L, Runge A, Wang T, Yuan L, Patel S, Iruela-Arispe L, Simon MC, Keith B. Endothelial deletion of hypoxia-inducible factor-2alpha (HIF-2alpha) alters vascular function and tumor angiogenesis. *Blood*. 2009; 114:469–477. [PubMed: 19439736]
123. Koh MY, Lemos R Jr, Liu X, Powis G. The hypoxia-associated factor switches cells from HIF-1alpha- to HIF-2alpha-dependent signaling promoting stem cell characteristics, aggressive tumor growth and invasion. *Cancer Res*. 2011; 71:4015–4027. [PubMed: 21512133]
124. Gordan JD, Bertout JA, Hu CJ, Diehl JA, Simon MC. HIF-2alpha promotes hypoxic cell proliferation by enhancing c-myc transcriptional activity. *Cancer Cell*. 2007; 11:335–347. [PubMed: 17418410]
125. Yamamoto K, Ohga N, Hida Y, Maishi N, Kawamoto T, Kitayama K, Akiyama K, Osawa t, Kondoh M, Matsuda K, Onodera Y, Fujie M, Kaga K, Hirano S, Shinohara N, Shindoh M, Hida K. Biglycan is a specific marker and an autocrine angiogenic factor of tumour endothelial cells. *Br J Cancer*. 2012; 106:1214–1223. [PubMed: 22374465]
126. Maishi N, Ohba Y, Akiyama K, Ohga N, Hamada J, Nagao-Kitamoto H, Alam MT, Yamamoto K, Kawamoto T, Inoue N, Taketomi A, Shindoh M, Hida Y, Hida K. Tumour endothelial cells in high metastatic tumours promote metastasis via epigenetic dysregulation of biglycan. *Sci Rep*. 2016; 6:28039. [PubMed: 27295191]
127. Allen M, Louise JJ. Jekyll and Hyde: the role of the microenvironment on the progression of cancer. *J Pathol*. 2011; 223:162–176. [PubMed: 21125673]
128. Rys JP, Monteiro DA, Alliston T. Mechanobiology of TGFbeta signaling in the skeleton. *Matrix Biol*. 2016; 52–54:413–425.

129. Sipila KH, Ranga V, Rappu P, Torittu A, Pirila L, Kapyla J, Johnson MS, Larjava H, Heino J. Extracellular citrullination inhibits the function of matrix associated TGF- β . *Matrix Biol.* 2016; 55:77–89. [PubMed: 26923761]
130. Horiguchi M, Ota M, Rifkin DB. Matrix control of transforming growth factor- β function. *J Biochem.* 2012; 152:321–329. [PubMed: 22923731]
131. Robertson IB, Horiguchi M, Zilberberg L, Dabovic B, Hadjiolova K, Rifkin DB. Latent TGF- β -binding proteins. *Matrix Biol.* 2015; 47:44–53. [PubMed: 25960419]
132. Hinz B. The extracellular matrix and TGF β 1: Tale of a strained relationship. *Matrix Biol.* 2015; 47:54–65. [PubMed: 25960420]
133. Hinz B, Phan SH, Thannickal VJ, Galli A, Bochaton-Piallat ML, Gabbiani G. The myofibroblast: one function, multiple origins. *Am J Pathol.* 2007; 170:1807–1816. [PubMed: 17525249]
134. Miettinen PJ, Ebner R, Lopez AR, Derynck R. TGF-beta induced transdifferentiation of mammary epithelial cells to mesenchymal cells: involvement of type I receptors. *J Cell Biol.* 1994; 127:2021–2036. [PubMed: 7806579]
135. Baghy K, Iozzo RV, Kovalszky I. Decorin-TGF β axis in hepatic fibrosis and cirrhosis. *J Histochem Cytochem.* 2012; 60:262–268. [PubMed: 22260996]
136. Mauviel A, Santra M, Chen YQ, Uitto J, Iozzo RV. Transcriptional regulation of decorin gene expression. Induction by quiescence and repression by tumor necrosis factor- α . *J Biol Chem.* 1995; 270:11692–11700. [PubMed: 7744809]
137. Pradeep S, Huang J, Mora EM, Nick AM, Cho MS, Wu SY, Noh K, Pecot CV, Rupaimoole R, Stein MA, Brock S, Wen Y, Xiong C, Gharpure K, Hansen JM, Nagaraja AS, Previs RA, Vivas-Mejia P, Han HD, Hu W, Mangala LS, Zand B, Stagg LJ, Ladbury JE, Ozpolat B, Alpay SN, Nishimura M, Stone RL, Matsuo K, Armaiz-Pena GN, Dalton HJ, Danes C, Goodman B, Rodriguez-Aguayo C, Kruger C, Schneider A, Haghepykar S, Jaladurgam P, Hung MC, Coleman RL, Liu J, Li C, Urbauer D, Lopez-Berestein G, Jackson DB, Sood AK. Erythropoietin Stimulates Tumor Growth via EphB4. *Cancer Cell.* 2015; 28:610–622. [PubMed: 26481148]
138. Inbar D, Cohen-Armon M, Neumann D. Erythropoietin-driven signalling and cell migration mediated by polyADP-ribosylation. *Br J Cancer.* 2012; 107:1317–1326. [PubMed: 22955851]
139. Lester RD, Jo M, Campana WM, Gonias SL. Erythropoietin promotes MCF-7 breast cancer cell migration by an ERK/mitogen-activated protein kinase-dependent pathway and is primarily responsible for the increase in migration observed in hypoxia. *J Biol Chem.* 2005; 280:39273–39277. [PubMed: 16207704]
140. Lee AS, Kim DH, Lee JE, Jung YJ, Kang KP, Lee S, Park SK, Kwak JY, Lee SY, Lim ST, Sung MJ, Yoon SR, Kim W. Erythropoietin induces lymph node lymphangiogenesis and lymph node tumor metastasis. *Cancer Res.* 2011; 71:4506–4517. [PubMed: 21586615]
141. Niedworok C, Rock K, Kretschmer I, Freudenberger T, Nagy N, Szarvas T, Vom DF, Reis H, Rubben H, Fischer JW. Inhibitory role of the small leucine-rich proteoglycan biglycan in bladder cancer. *PLoS One.* 2013; 8:e80084. [PubMed: 24223213]
142. Rydstrom K, Joost P, Ehinger M, Eden P, Jerkeman M, Cavallin-Stahl E, Linderroth J. Gene expression profiling indicates that immunohistochemical expression of CD40 is a marker of an inflammatory reaction in the tumor stroma of diffuse large B-cell lymphoma. *Leuk Lymphoma.* 2012; 53:1764–1768. [PubMed: 22335531]
143. Merline R, Moreth K, Beckmann J, Nastase MV, Zeng-Brouwers J, Tralhão JG, Lemarchand P, Pfeilschifter J, Schaefer RM, Iozzo RV, Schaefer L. Signaling by the matrix proteoglycan decorin controls inflammation and cancer through PDCD4 and microRNA-21. *Sci Signal.* 2011; 4:ra75. [PubMed: 22087031]
144. Koninger J, Giese NA, Bartel M, di Mola FF, Berberat PO, di Sebastiano P, Giese T, Büchler MW, Friess H. The ECM proteoglycan decorin links desmoplasia and inflammation in chronic pancreatitis. *J Clin Pathol.* 2006; 59:21–27. [PubMed: 16394277]
145. Reed CC, Gaudie J, Iozzo RV. Suppression of tumorigenicity by adenovirus-mediated gene transfer of decorin. *Oncogene.* 2002; 21:3688–3695. [PubMed: 12032837]
146. Reed CC, Waterhouse A, Kirby S, Kay P, Owens RA, McQuillan DJ, Iozzo RV. Decorin prevents metastatic spreading of breast cancer. *Oncogene.* 2005; 24:1104–1110. [PubMed: 15690056]

147. Xu W, Neill T, Yang Y, Hu Z, Cleveland E, Wu Y, Hutten R, Xiao X, Stock SR, Shevrin D, Kaul K, Brendler C, Iozzo RV, Seth P. The systemic delivery of an oncolytic adenovirus expressing decorin inhibits bone metastasis in a mouse model of human prostate cancer. *Gene Therapy*. 2015; 22:31–40.
148. Yang Y, Xu WW, Neill T, Hu Z, Wang CH, Xiao X, Stock S, Guise T, Yun CO, Brendler CB, Iozzo RV, Seth P. Systemic Delivery of an Oncolytic Adenovirus Expressing Decorin for the Treatment of Breast Cancer Bone Metastases. *Hum Gene Ther*. 2015; 26:813–825. [PubMed: 26467629]
149. Launay G, Salza R, Multedo D, Thierry-Mieg N, Ricard-Blum S. MatrixDB, the extracellular matrix interaction database: updated content, a new navigator and expanded functionalities. *Nucleic Acids Res*. 2015; 43:D321–D327. [PubMed: 25378329]
150. Naba A, Clauser KR, Ding H, Whittaker CA, Carr SA, Hynes RO. The extracellular matrix: Tools and insights for the “omics” era. *Matrix Biol*. 2016; 49:10–24. [PubMed: 26163349]

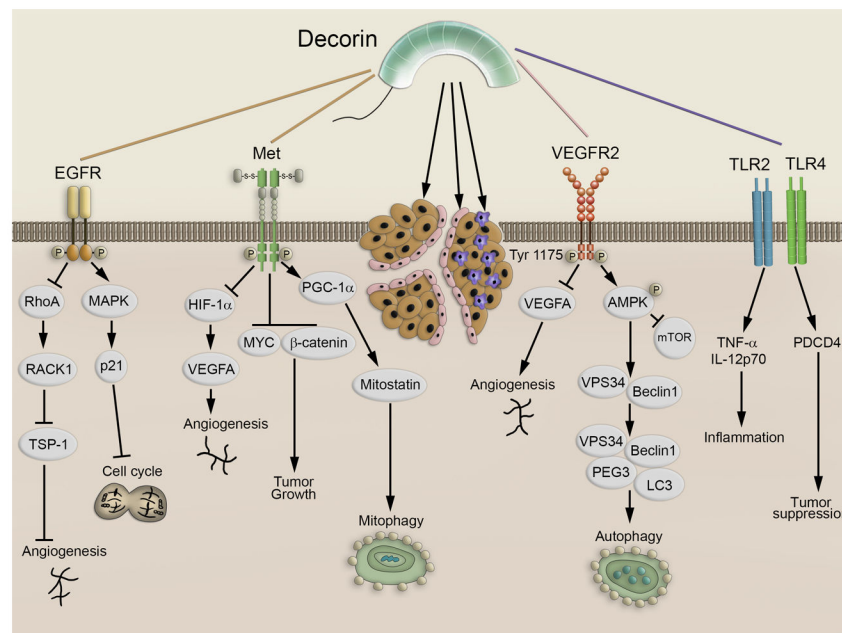
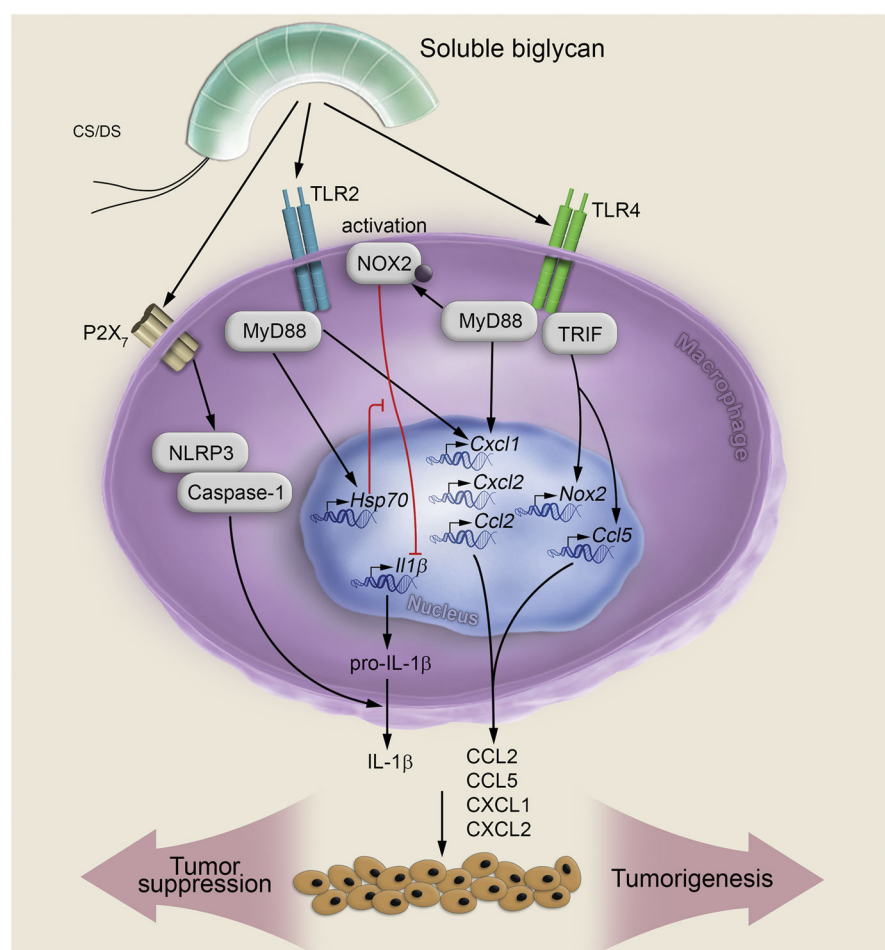


Fig. 1.

Decorin exhibits promiscuity in its ability to alter tumorigenesis via regulation of angiogenesis, autophagy, and inflammation. Decorin antagonizes RTKs, such as EGFR and Met, on the tumor cell surface to inhibit angiogenesis via suppression of pro-angiogenic factors, such as VEGFA, as well as induction of anti-angiogenic proteins, such as thrombospondin-1, while simultaneously reducing tumor growth via cell cycle arrest and inhibition of Myc and β -catenin. Signaling through Met also induces mitostatin leading to tumor cell mitophagy. Interaction between decorin and VEGFR2 in endothelial cells results in autophagic induction vis-à-vis the canonical intermediates, Beclin 1 and LC3 as well as with the novel autophagic regulator, Peg3. In inflammatory cells, decorin signals through TLR2/4 to induce pro-inflammatory mediators, which reduces tumor growth.

**Fig. 2.**

Dual role of biglycan in the control of inflammation and tumorigenesis. Soluble biglycan triggers mRNA expression of pro-inflammatory cytokines and chemokines IL1β, Cxcl1, Cxcl2, and Ccl2 in macrophages in a TLR2/4- and MyD88-dependent manner, while it selectively stimulates the expression of Ccl5 through TLR4 and TRIF. By clustering TLR2/4 and the P2X₇ soluble biglycan induces the NLRP3-inflammasome, activating caspase-1 and releasing mature IL-1β. Moreover, biglycan directly mediates the expression of Nox2 mRNA via TLR4/TRIF and the activation of NOX2 in a TLR4/MyD88-dependent manner, thereby attenuating the expression of the pro-inflammatory cytokine IL-1β. In contrast, by engaging TLR2, soluble biglycan triggers the expression of HSP70, which binds to NOX2, and consequently impairs the inhibitory function of NOX2 on biglycan-mediated IL-1β expression and maturation.

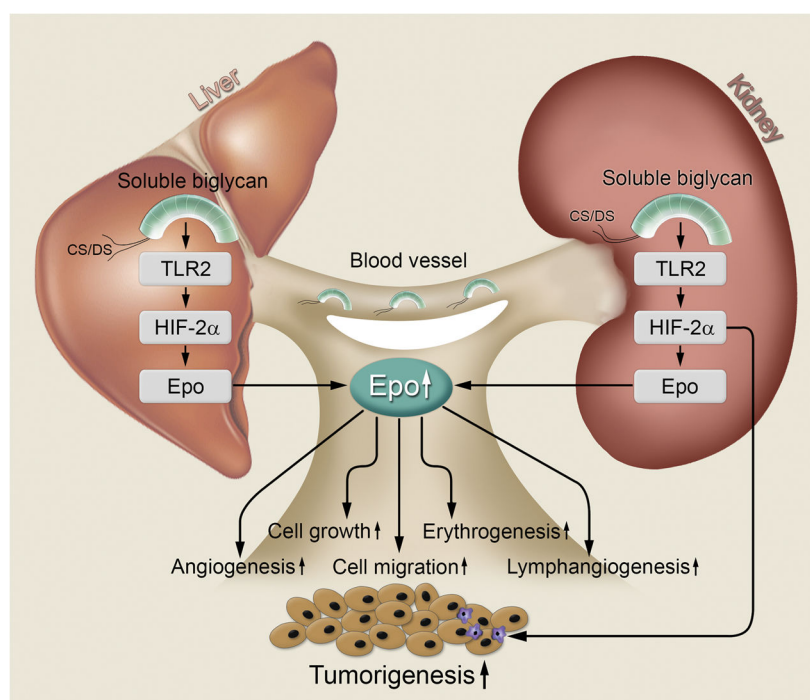


Fig. 3. Biglycan induces tumorigenesis via HIF-2 α stabilization and Epo expression. Soluble biglycan binds to the TLR2 in the kidney and/or in the liver and induces the stabilization of HIF-2 α produced in liver, kidneys and tumor-associated macrophages (purple). HIF-2 α subsequently induces the expression of erythropoietin (Epo). Epo is then released into the circulation, where it may stimulate tumor angiogenesis, cell growth and cell migration, as well as tumor lymphangiogenesis, thus inducing tumorigenesis.