

4-26-2022

Cyclophosphamide induces durable molecular and clinical responses in patients with relapsed T-LGL leukemia.

Zachary Braunstein

Eric McLaughlin

Anjali Mishra

Jonathan E Brammer

Follow this and additional works at: <https://jdc.jefferson.edu/medoncfp>

 Part of the [Oncology Commons](#)

[Let us know how access to this document benefits you](#)

This Article is brought to you for free and open access by the Jefferson Digital Commons. The Jefferson Digital Commons is a service of Thomas Jefferson University's [Center for Teaching and Learning \(CTL\)](#). The Commons is a showcase for Jefferson books and journals, peer-reviewed scholarly publications, unique historical collections from the University archives, and teaching tools. The Jefferson Digital Commons allows researchers and interested readers anywhere in the world to learn about and keep up to date with Jefferson scholarship. This article has been accepted for inclusion in Department of Medical Oncology Faculty Papers by an authorized administrator of the Jefferson Digital Commons. For more information, please contact: JeffersonDigitalCommons@jefferson.edu.

TO THE EDITOR:

Cyclophosphamide induces durable molecular and clinical responses in patients with relapsed T-LGL leukemia

Zachary Braunstein,¹ Eric McLaughlin,² Anjali Mishra,^{3,4} and Jonathan E. Brammer⁵

¹Department of Internal Medicine, The Ohio State University Wexner Medical Center, Columbus, OH; ²Center for Biostatistics, Department of Biomedical Informatics, The Ohio State University, Columbus, OH; ³Department of Medical Oncology and ⁴Department of Cancer Biology, Sydney Kimmel Cancer Center, Thomas Jefferson University, Philadelphia, PA; and ⁵Division of Hematology, Department of Internal Medicine, James Comprehensive Cancer Center, The Ohio State University, Columbus, OH

T-cell large granular lymphocytic leukemia (T-LGL) is a clonal proliferation of cytotoxic T-lymphocytes with a terminal effector memory phenotype ($CD3^+/CD8^+/CD5dim/CD57^+/CD62L/CD45RA^+/CD45RO^-$) that can result in severe cytopenias, including neutropenia, anemia, and pancytopenia/bone marrow failure in severe cases.¹⁻³ The pathogenesis of T-LGL is thought to be mediated by increased circulating interleukin-15, and subsequent upregulation of the STAT3 pathway, leading to dysregulation of apoptosis and resultant cellular proliferation and marrow damage. The management of T-LGL therefore is immune-suppressive therapy, with methotrexate (MTX), cyclophosphamide (Cy), and cyclosporine serving as the standard frontline agents. MTX is the standard frontline therapy based off the prospective ECOG5998 (E5998) study but overall response rate (ORR) was modest at 38%.⁴ An initial early report with frontline Cy demonstrated a response rate of 63% in 10/16 patients, including 6 complete response (CR), though at that time, CR was defined as hematologic CR, and response criteria differed from current E5998 criteria.⁵ For patients that fail frontline MTX, oral Cy, with a target dose of 100 mg/day is the standard approach. Data from the ECOG5998 trial suggested an increased ORR with Cy in the second-line setting, with 21% CR,⁶ and retrospective data from the French cohort study suggested improved response rate, with 63% CR,⁷ when Cy was used second line. More intriguingly, anecdotal evidence suggests that because of its alkylating effects, Cy may eradicate the T-LGL clone, which has not been observed with immune-suppressive-based strategies. However, the degree to which a complete molecular remission (CMR) can be attained, and the lengths of these remissions with Cy remains unknown.

Recently, we published a report on T-LGL that showed improved clinical responses in patients treated with Cy, especially if this was used as second line after frontline MTX.⁸ In that study, 100% of patients (7/7) that were treated with Cy as second-line therapy after frontline MTX had a response to treatment (3 CR and 4 partial response [PR]). Here, we report on additional patients ($n = 22$) treated with Cy for relapsed or refractory T-LGL, and for the first time report survival outcomes, duration of response, and presence of CMR in the relapsed setting.

We retrospectively evaluated all patients treated for T-LGL with oral Cy at the Ohio State University (OSU) James Comprehensive Cancer Center from 2000 through 2019. The diagnosis of T-LGL was made based on 2016 World Health Organization criteria. T-LGL criteria included a $CD3^+ CD8^+$ population on flow cytometry ≥ 500 cells/ mm^3 and a positive monoclonal T-cell receptor (TCR). This was considered positive if detected by TCR polymerase chain reaction (PCR) or by restriction of TCR-Vbeta noted on flow cytometry. For patients diagnosed with a clonal TCR by flow cytometry, a panel of 30 TCR-Vbeta rearrangements was used and considered positive if 1 or more clone was detected in $\geq 10\%$ of events. Disease response was defined by the E5998 study criteria and was confirmed retrospectively by the investigators (Z.B. and J.E.B.). Although there is variability in follow-up given the retrospective nature of this study, in general, as standard clinical care, patients were monitored for response on Cy for a minimum of 4 months at the full 100-mg dosage. Standard laboratory values (complete blood count with differential, hepatic function panel, basic chemistry panel) were drawn every 4 to 6 weeks either at OSU or with a local physician. Flow cytometry was

Submitted 28 September 2021; accepted 16 December 2021; prepublished online on *Blood Advances* First Edition 7 January 2022; final version published online 25 April 2022. DOI 10.1182/bloodadvances.2021006263.

Requests for data sharing may be submitted to Jonathan E. Brammer (jonathan.brammer@osumc.edu).

Presented in abstract form at the 63rd American Society of Hematology Annual Meeting in December 2021.

The online version of this article contains a data supplement.

© 2022 by The American Society of Hematology. Licensed under Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0), permitting only noncommercial, nonderivative use with attribution. All other rights reserved.

Table 1. Baseline characteristics of patients attaining CR with cyclophosphamide

Age at Dx	Sex	Rheumatologic disease	Prior Tx	Cy line	Cy indication	Max Cy dose	Time on Cy, mo	Time before CR after starting Cy, mo	Did patient have CMR (time to CMR, M)?
66	F	RA	CsA (×2), MTX	4th	Neutropenia	50 mg BID	11	12	No
66	M	None	MTX	2nd	Anemia	50 mg BID	10	5.5	No
70	F	RA	MTX	2nd	Anemia	50 mg daily	12	5	Yes (7 mo)
67	F	None	MTX (×2)	3rd	Anemia and Neutropenia	100 mg daily	13	4	Yes (12 mo)
66	M	None	MTX, BNZ-1	3rd	Neutropenia	100 mg daily	15	5	Yes (11 mo)
65	M	RA	MTX	2nd	Neutropenia	50 mg BID	11	3	No

BID, twice daily; CsA, cyclosporine; Dx, diagnosis; MTX, methotrexate; RA, rheumatoid arthritis; Tx, treatment.

assessed every 2 to 3 months to evaluate for residual disease. Patients who attained an improvement in their affected blood counts (ie, neutropenia or anemia) were continued on Cy, even if full E5998 response criteria were not attained because E5998 criteria were assessed retrospectively only. In this way, patients who attained some response at 4 months could be evaluated months later to assess for a deepening of their clinical response. CMR was defined as: CR by E5998 criteria and clearance of the TCR PCR-based gene rearrangement studies or TCR Vβ flow cytometry assessment for individual clones. The time-to-response (TTR) was measured as time from start of Cy until PR or CR, with patients who failed to respond being censored at the end of their Cy treatment. Leukemia-free survival (LFS) in patients responding to Cy was measured as time from start of Cy until disease progression, with patients without progression being censored at last follow-up. TTR and LFS were compared across variables using Kaplan-Meier curves with median survival and 95% confidence intervals. The OSU's institutional review board approved this study, which was conducted in accordance with the Declaration of Helsinki.

Twenty-two patients with relapsed/refractory T-LGLL, with an average duration of 8 months (range, 1-15) of Cy treatment, and a median follow-up time of 24.4 months, were included in this analysis. Thirteen patients (59%) were treated with Cy as second line and 9 (41%) as third line or greater. Standard dosing included an initial dose of 50 mg daily for 2 weeks with increase to 100 mg if tolerated for 4 months. Cy was dosed at 150 mg (1 patient), 100 mg (17 patients), and 50 mg (3 patients), whereas 1 patient received 1260 mg for multiple myeloma. Assessment for clonal hematopoiesis (CHIP) or associated mutations was not routinely evaluated. As part of our standard panel, mutations were assessed in 11/22 (50%) patients because this panel became available for T-LGLL only in 2018. Four patients had no mutations, and 7 patients were found to have the following mutations (2 patients had 2 separate mutations): STAT3 (×5), CARD11, NOTCH2, SF3B1, and DNMT3A. Two patients were found to have 2 separate mutations, 1 with CARD11/NOTCH2 and another with STAT3 (D661Y)/SF3B1. Of the STAT3 mutations, 3 patients had a D661Y mutation, 1 had a D661V mutation, and 1 had an N647I mutation. Overall, 11 patients were tested for STAT3 mutations because this was only routinely done after 2018. Of those 11 patients, 5 (45%) were found to have a STAT3 mutation (supplemental Table 1). Of these, 4 patients with a STAT3 mutation had no response to Cy and 1 patient had a PR to Cy (this patient also had an SF3B1 mutation) (supplemental Table 1). At the low doses of Cy used for T-LGLL, Cy is generally very well tolerated, with only minor toxicities, including nausea, cytopenias, and fatigue. In our cohort of patients, the most common side effect observed

was grade 1 fatigue (n = 2). We also observed grade 1 dizziness (n = 1) and 1 patient with grade 1 leukopenia.

For relapsed/refractory T-LGLL, the ORR was 68% (15/22) with 6 CR (27%) and 9 PR (41%). At the 4-month mark, the ORR was 27% (2 CR, 4 PR). The median time to response was 5.6 months for CR and 5.8 months for PR. Three patients that had a PR at the 4-month mark later converted to a CR. The median time to maximum response TTRMax (CR or PR) was 6 months (95% confidence interval, 4-7), whereas median time to PR and CR were 9 and 7 months, respectively. Baseline characteristics of those achieving CR can be seen in Table 1. Interestingly, of the 6 patients that achieved a CR, 50% had a CMR with durable eradication of their T-cell clone. Among responders, the median follow-up time was 27 months and the median LFS was 24 months. Of note, among those with a CR, median follow-up is slightly longer at 28.5 months. When broken down by type of response, the median LFS for those with PR was 27 months, whereas the median LFS for those with CR was not reached as no patients progressed. Additionally, for patients that achieved a CMR, the LFS has not yet been reached.

Here we report that patients receiving Cy for relapsed T-LGLL can attain durable remissions, including CMR, with a prolonged duration of response with long-term follow-up. Previous reports have focused on response rates of Cy but have not evaluated the duration of response in these patients, and no studies have reported CMR in patients with relapsed/refractory T-LGLL. Of particular interest is that no patients who attained a CR have relapsed, demonstrating durable remissions are attainable with Cy in relapsed T-LGLL. We also demonstrate that a CMR is attainable in refractory patients with Cy, which has previously only been shown in patients that were treated with Cy frontline.⁹ Importantly, we demonstrate that there was significant consolidation of response after the initial 4-month treatment period, with an ORR of 27% at 4 months, but an ORR of 64% in long-term follow-up. These novel results suggest that patients should remain on Cy beyond initial response because Cy can deepen the clinical response over time. To assess for response, we recommend checking for the TCR (by PCR of flow cytometry) every 3 months while on treatment to assess for response and CMR. The standard approach at OSU is to treat patients with Cy for a maximum of 12 months because of the increased risk of mutagenesis, myelodysplasia, or marrow injury resulting from prolonged Cy.¹⁰⁻¹³ This also seems adequate time to attain a maximum response, as the median TTRMax (CR or PR) in our study was 6 months, and most patients responded by 8 months. However, 4 patients continued treatment beyond 12 months solely at the discretion of their local

physician, outside of OSU recommendations, with 2 patients on Cy for 13 total months, 1 for 14 months, and 1 for 15 months. Our data clearly demonstrate that maximal response is typically obtained by 8 months of treatment, and that these responses are durable, with no long-term relapses in patients with CR or CMR.

We acknowledge certain limitations of this study. No CR patients had pancytopenia, which can result in selection bias in a small population of patients with CR. Further, long-term follow-up is needed to assess the duration of remission, and impact of CMR on patients with relapsed T-LGLL treated with Cy. Clonal hematopoiesis (CHIP) mutations were also not routinely evaluated, and in these patients, caution should be used when initiating treatment with Cy. Further, the clinical and prognostic significance of STAT3 mutations and the response to Cy will need to be evaluated in future prospective studies. Although our results strongly suggest that Cy can produce durable remissions, and CMR, these findings will need to be further validated in a larger cohort in future prospective randomized studies, including the use of CMR as a novel endpoint in T-LGLL. Nevertheless, our results demonstrate that Cy induces durable responses in the setting of relapsed T-LGLL, including CMRs. Pending future studies, we suggest that Cy should be maintained for up to 8 to 12 months to consolidate response, with the goal of attaining a CR or CMR. Further, CMR, as a component of response criteria, should be incorporated in the design of future clinical trials.

Acknowledgment: Funding support for this article was provided by the National Institutes of Health/National Center for Advancing Translational Sciences (KL2T R002734 [J.E.B.]).

Authorship: Z.B. and J.E.B. designed the study; E.M. performed the statistical analysis; A.M. assisted with analysis; Z.B. and J.E.B. were responsible for data collection, data analysis, data interpretation, manuscript preparation, writing and completion and final approval of manuscript; and all authors approved the final version of the manuscript and the submission.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

ORCID profiles: A.M., 0000-0001-6269-5424.

Correspondence: Jonathan E. Brammer, James Comprehensive Cancer Center, The Ohio State University, 1800 Cannon Dr, Lincoln Tower, 1120D, Columbus, OH 43210; e-mail: Jonathan.brammer@osumc.edu.

References

1. Loughran TP Jr, Kadin ME, Starkebaum G, et al. Leukemia of large granular lymphocytes: association with clonal chromosomal

abnormalities and autoimmune neutropenia, thrombocytopenia, and hemolytic anemia. *Ann Intern Med.* 1985;102(2): 169-175.

2. Swerdlow SH, Campo E, Pileri SA, et al. The 2016 revision of the World Health Organization classification of lymphoid neoplasms. *Blood.* 2016;127(20):2375-2390.
3. Semenzato G, Zambello R, Starkebaum G, Oshimi K, Loughran TP Jr. The lymphoproliferative disease of granular lymphocytes: updated criteria for diagnosis. *Blood.* 1997;89(1): 256-260.
4. Loughran TP Jr, Kidd PG, Starkebaum G. Treatment of large granular lymphocyte leukemia with oral low-dose methotrexate. *Blood.* 1994;84(7):2164-2170.
5. Dhodapkar MV, Li CY, Lust JA, Tefferi A, Phylliky RL. Clinical spectrum of clonal proliferations of T-large granular lymphocytes: a T-cell clonopathy of undetermined significance? *Blood.* 1994;84(5): 1620-1627.
6. Loughran TP Jr, Zickl L, Olson TL, et al. Immunosuppressive therapy of LGL leukemia: prospective multicenter phase II study by the Eastern Cooperative Oncology Group (E5998). *Leukemia.* 2015; 29(4):886-894.
7. Bareau B, Rey J, Hamidou M, et al. Analysis of a French cohort of patients with large granular lymphocyte leukemia: a report on 229 cases. *Haematologica.* 2010;95(9):1534-1541.
8. Braunstein Z, Mishra A, Staub A, Freud AG, Porcu P, Brammer JE. Clinical outcomes in T-cell large granular lymphocytic leukaemia: prognostic factors and treatment response. *Br J Haematol.* 2021; 192(3):484-493.
9. Moignet A, Hasanali Z, Zambello R, et al. Cyclophosphamide as a first-line therapy in LGL leukemia. *Leukemia.* 2014;28(5):1134-1136.
10. Faurschou M, Mellekjaer L, Voss A, Keller KK, Hansen IT, Baslund B. Prolonged risk of specific malignancies following cyclophosphamide therapy among patients with granulomatosis with polyangiitis. *Rheumatology (Oxford).* 2015;54(8): 1345-1350.
11. Sanderson BJ, Johnson KJ, Henner WD. Induction of mutant lymphocytes in cyclophosphamide- and chlorambucil-treated patients. *Mutagenesis.* 2001;16(3):197-202.
12. Xu Y, Wang H, Zhou S, et al. Risk of second malignant neoplasms after cyclophosphamide-based chemotherapy with or without radiotherapy for non-Hodgkin lymphoma. *Leuk Lymphoma.* 2013;54(7): 1396-1404.
13. T-Cell Lymphomas. In: Network NCC ed. NCCN Clinical Practice Guidelines. Vol. 1. 2021. Plymouth Meeting, PA: National Comprehensive Cancer Network; 2020. Available at: <https://jnccn.org/view/journals/jnccn/18/11/article-p1460.xml>.