

5-2-2021

Choline-Sigma-1R as an Additional Mechanism for Potentiation of Orexin by Cocaine

Jeffrey Barr

Pingwei Zhao

G Cristina Brailoiu

Eugen Brailoiu

Follow this and additional works at: <https://jdc.jefferson.edu/petfp>

 Part of the [Medical Pharmacology Commons](#)

[Let us know how access to this document benefits you](#)

This Article is brought to you for free and open access by the Jefferson Digital Commons. The Jefferson Digital Commons is a service of Thomas Jefferson University's [Center for Teaching and Learning \(CTL\)](#). The Commons is a showcase for Jefferson books and journals, peer-reviewed scholarly publications, unique historical collections from the University archives, and teaching tools. The Jefferson Digital Commons allows researchers and interested readers anywhere in the world to learn about and keep up to date with Jefferson scholarship. This article has been accepted for inclusion in Department of Pharmacology and Experimental Therapeutics Faculty Papers by an authorized administrator of the Jefferson Digital Commons. For more information, please contact: JeffersonDigitalCommons@jefferson.edu.



Article

Choline-Sigma-1R as an Additional Mechanism for Potentiation of Orexin by Cocaine

Jeffrey L. Barr ^{1,†}, Pingwei Zhao ¹, G. Cristina Brailoiu ² and Eugen Brailoiu ^{1,*}

¹ Center for Substance Abuse Research, Lewis Katz School of Medicine at Temple University, Philadelphia, PA 19140, USA; Jeffrey.Barr@SanfordHealth.org (J.L.B.); zhaopw@temple.edu (P.Z.)

² Department of Pharmaceutical Sciences, Jefferson College of Pharmacy, Thomas Jefferson University, Philadelphia, PA 19107, USA; Gabriela.Brailoiu@jefferson.edu

* Correspondence: eugen.brailoiu@temple.edu

† Current address: Cancer and Immunotherapies Group, Sanford Research, Sioux Falls, SD 57104, USA.

Abstract: Orexin A, an endogenous peptide involved in several functions including reward, acts via activation of orexin receptors OX₁ and OX₂, Gq-coupled GPCRs. We examined the effect of a selective OX₁ agonist, OXA (17-33) on cytosolic calcium concentration, [Ca²⁺]_i, in neurons of nucleus accumbens, an important area in the reward circuit. OXA (17-33) increased [Ca²⁺]_i in a dose-dependent manner; the effect was prevented by SB-334867, a selective OX₁ receptors antagonist. In Ca²⁺-free saline, the OXA (17-33)-induced increase in [Ca²⁺]_i was not affected by pretreatment with bafilomycin A1, an endo-lysosomal calcium disrupter, but was blocked by 2-APB and xestospingon C, antagonists of inositol-1,4,5-trisphosphate (IP₃) receptors. Pretreatment with VU0155056, PLD inhibitor, or BD-1047 and NE-100, Sigma-1R antagonists, reduced the [Ca²⁺]_i response elicited by OXA (17-33). Cocaine potentiated the increase in [Ca²⁺]_i by OXA (17-33); the potentiation was abolished by Sigma-1R antagonists. Our results support an additional signaling mechanism for orexin A-OX₁ via choline-Sigma-1R and a critical role for Sigma-1R in the cocaine-orexin A interaction in nucleus accumbens neurons.

Keywords: choline; orexin A; OX₁ receptor; phospholipase D; PLD; reward



Citation: Barr, J.L.; Zhao, P.; Brailoiu, G.C.; Brailoiu, E. Choline-Sigma-1R as an Additional Mechanism for Potentiation of Orexin by Cocaine. *Int. J. Mol. Sci.* **2021**, *22*, 5160. <https://doi.org/10.3390/ijms22105160>

Academic Editor: Carmen Abate

Received: 5 April 2021
Accepted: 11 May 2021
Published: 13 May 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Orexin A and B (also known as hypocretin-1 and -2) are endogenous neuropeptides synthesized in hypothalamic neurons that control appetite, sleep/wakefulness, hormone release, stress, and drug-seeking behavior [1–3]. Hypothalamic neurons expressing orexins project to several brain areas such as ventral tegmental area, nucleus accumbens, dorsal raphe nucleus, and locus coeruleus [2,4].

Orexins act via OX₁ and OX₂ receptors, Gq-coupled GPCRs that may signal also via Gs or Gi proteins [5,6]. OX₁ receptors have a preferential role in addiction, reward, and motivation, while OX₂ receptors are involved in arousal [3,7]. OX₁ receptor activation leads to an increase in cytosolic Ca²⁺ concentration, [Ca²⁺]_i, subsequent to activation of phospholipase C (PLC) and generation of inositol-1,4,5-trisphosphate (IP₃) [6,8]. In addition to the PLC coupling, activation of OX₁ receptor leads to phospholipase D (PLD) activation [9,10]. PLD-mediated hydrolysis of phosphatidylcholine produces choline and phosphatidic acid [11]. Choline activates Sigma-1R [12], a chaperone protein residing at the endoplasmic reticulum that potentiates IP₃-induced Ca²⁺ release [13].

OX₁ receptors were identified in brain nuclei from the reward circuit, including nucleus accumbens [14,15], and OX₁-selective antagonists have been evaluated as potential therapeutic agents for addiction treatment [16–18]. Previous studies indicate that orexins via OX₁ receptor activation are involved in the response to cocaine and play multiple roles in cocaine addiction-related behaviors [7,19–21]. Orexin-OX₁ signaling is required for stimulant locomotor sensitization and cocaine seeking when it is driven by highly

motivated states [3]. OX_1 receptors in the nucleus accumbens mediate chronic cocaine-induced locomotor sensitization [22]. Other studies indicate that SB-334867, a selective OX_1 receptor antagonist, prevents cocaine seeking and is a potential treatment target for cocaine relapse prevention [23].

Cocaine acts primarily by blocking the dopamine transporter, thus increasing dopamine transmission in the nucleus accumbens [24], an important area in the reward circuit [25]. In addition, cocaine binds to and activates Sigma-1R [26]. Since nucleus accumbens neurons express Sigma-1R [27–29] and OX_1 receptors [14,15] and behavioral studies support the cocaine–orexin interaction at this level [3,22], in this work, we examined the underlying mechanisms and role of Sigma-1R in the cocaine–orexin interaction in nucleus accumbens neurons.

2. Results

2.1. OXA (17-33) Increases Cytosolic Ca^{2+} , $[Ca^{2+}]_i$, in Nucleus Accumbens Neurons via OX_1 Receptor Activation

OXA (17-33) (0.1–100 nM), i.e., truncated orexin A, a selective OX_1 agonist [1,30], increased $[Ca^{2+}]_i$ in nucleus accumbens neurons in a dose-dependent manner (Figure 1). OXA (17-33) (10 nM) increased the fluorescence F340/380 ratio of Fura-2AM-loaded nucleus accumbens neurons; the effect was prevented by pretreatment with SB-334867 (1 μ M), a selective OX_1 antagonist [31] (Figure 1A). OXA (17-33) (10 nM) produced a transient increase in $[Ca^{2+}]_i$ that was abolished by SB-334867 (Figure 1B). Comparison of the amplitude of the increase in $[Ca^{2+}]_i$ produced by different concentrations of OXA (17-33) (0.1, 1, 10, 100 nM) is illustrated in Figure 1C ($n = 6$ neurons/each concentration). Of note, 20–30 neurons were tested for each condition, and an increase in $[Ca^{2+}]_i$ was identified in about 25% of neurons tested; the amplitude of $[Ca^{2+}]_i$ from the response of six neurons was used for analysis.

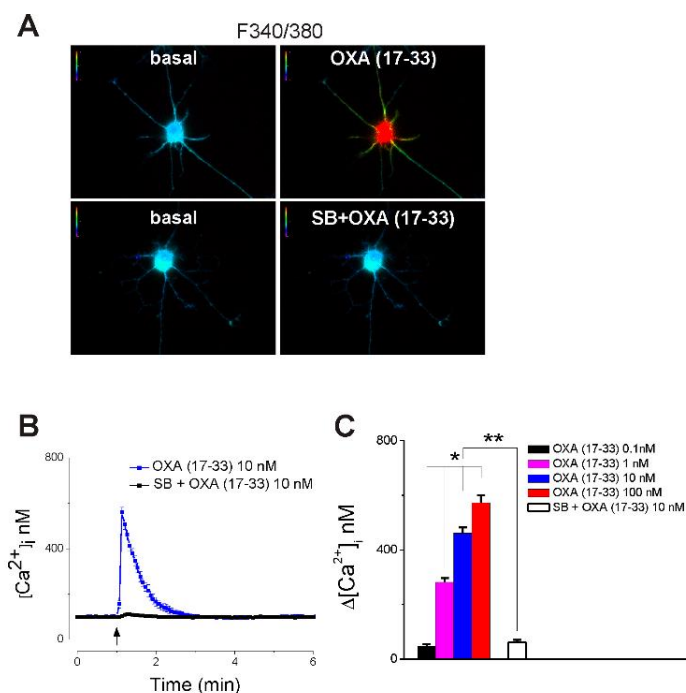


Figure 1. OXA (17-33) increases cytosolic Ca^{2+} concentration, $[Ca^{2+}]_i$, in nucleus accumbens neurons via OX_1 receptor activation. (A) Representative examples of fluorescence F340/380 ratio of Fura-2AM-loaded nucleus accumbens neurons in basal conditions (left) and after treatment with OXA (17-33) (10 nM), a selective OX_1 agonist, alone (top right) or in the presence of OX_1 antagonist, SB-334867 (1 μ M) (bottom right). (B) OXA (17-33) (10 nM) produced a transient increase in $[Ca^{2+}]_i$; the effect was abolished by SB-334867. (C) Comparison of the amplitude of $[Ca^{2+}]_i$ increase (mean + SD) produced by OXA (17-33) (0.1, 1, 10, and 100 nM); $p < 0.05$ as compared with the amplitude of $[Ca^{2+}]_i$ increase produced by each concentration (*) or by OXA (17-33) (10 nM) (**); $n = 6$ neurons/each concentration tested.

2.2. OXA (17-33) Increases $[Ca^{2+}]_i$ via IP_3 -Dependent Mechanism

In Ca^{2+} -free saline, OXA (17-33) (10 nM) elicited an increase in $[Ca^{2+}]_i$ of lower amplitude (Figure 2) than in Ca^{2+} -containing saline (Figure 1). The Ca^{2+} response to OXA (17-33) (10 nM) in Ca^{2+} -free saline was abolished by pretreatment with IP_3 receptors antagonists 2-aminoethoxydiphenyl borate (2-APB, 100 μ M, 15 min) and xestospongine C (10 μ M, 15 min) [32], indicating a PLC-dependent mechanism. Disruption of lysosomal Ca^{2+} stores with bafilomycin A1 (1 μ M, 1 h preincubation), a V-type ATPase inhibitor that prevents lysosomal acidification [33], did not affect the Ca^{2+} response to orexin (10 nM) (Figure 2). OXA (17-33) (10 nM)-induced Ca^{2+} responses (average \pm SD) in Ca^{2+} -free saline in nucleus accumbens neurons in the absence and presence of 2-APB and xestospongine C or bafilomycin A1 are illustrated in Figure 2A, and a comparison of the amplitude of the $[Ca^{2+}]_i$ increase in each condition is illustrated in Figure 2B ($n = 6$ neurons/condition).

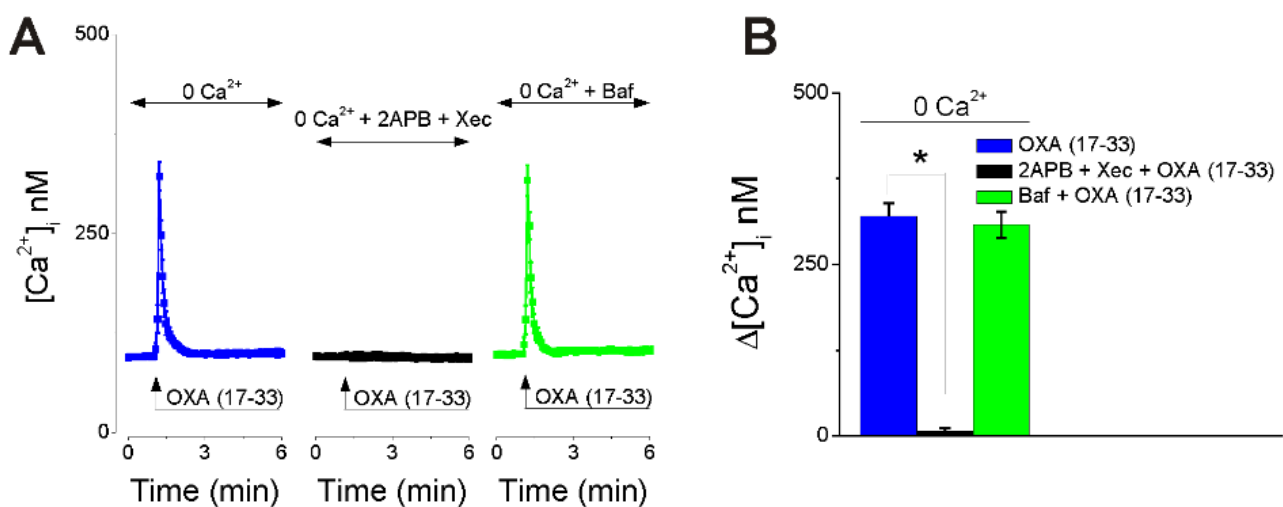


Figure 2. OXA (17-33) increases $[Ca^{2+}]_i$ via IP_3 -dependent mechanism. (A) Illustration of average Ca^{2+} transients (\pm SD) induced in Ca^{2+} -free saline by OXA (17-33) (10 nM) alone (left) and OXA (17-33) (10 nM) after pretreatment with 2-aminoethoxydiphenyl borate (2-APB, 100 μ M) and xestospongine C (XeC, 10 μ M, 15 min), IP_3 receptor antagonists (middle), or with bafilomycin A1 (Baf, 1 μ M) (right). (B) Comparison of the amplitude of the increase in $[Ca^{2+}]_i$ (average \pm SD) in each condition. Pretreatment with 2-APB and xestospongine C abolished the Ca^{2+} response induced by OXA (17-33). * $p < 0.05$; $n = 6$ neurons/condition.

2.3. OXA (17-33) Increases $[Ca^{2+}]_i$ via Choline-Sigma-1R-Dependent Mechanism

Pretreatment with VU0155056 (1 μ M, 30 min), a PLD inhibitor [34], reduced the amplitude of OXA (17-33) (10 nM)-induced increase in $[Ca^{2+}]_i$ by 33% (Figure 3). Pretreatment with BD1047 (50 μ M, 30 min) or NE-100 (5 μ M, 30 min) (Sigma-1R antagonists) [35,36] reduced the Ca^{2+} response to OXA (17-33) (10 nM) by 18.1% and 20.4%, respectively. Average Ca^{2+} responses induced by OXA (17-33) alone and in the presence of PLD inhibitor and Sigma-1R antagonists are illustrated in Figure 3A, and a comparison of the amplitude of the $[Ca^{2+}]_i$ increase in each condition is illustrated in Figure 3B ($n = 6$ neurons/condition).

2.4. Cocaine Potentiates OXA (17-33)-Induced Increase in $[Ca^{2+}]_i$ via Sigma-1R Activation

Cocaine (10 μ M), while it did not elicit a Ca^{2+} response by itself, potentiated the increase in $[Ca^{2+}]_i$ produced by OXA (17-33) (10 nM), when added at the same time as OXA (17-33) (Figure 4). Pretreatment with BD1047 (50 μ M, 30 min) or NE-100 (5 μ M, 30 min), Sigma-1R antagonists, reduced the increase in $[Ca^{2+}]_i$ produced by cocaine + OXA (17-33) (10 nM), by 30.7% and 33.1%, respectively (Figure 4). This indicates that antagonism of Sigma-1R abolished the potentiation produced by cocaine and further reduced the Ca^{2+} response to OXA (17-33) (10 nM) to the same level as in neurons treated with Sigma-1R

antagonists before OXA (17-33) alone (Figure 4 vs. Figure 3). A comparison of the amplitude of the $[Ca^{2+}]_i$ increase in each condition is illustrated in Figure 4B ($n = 6$ neurons/condition).

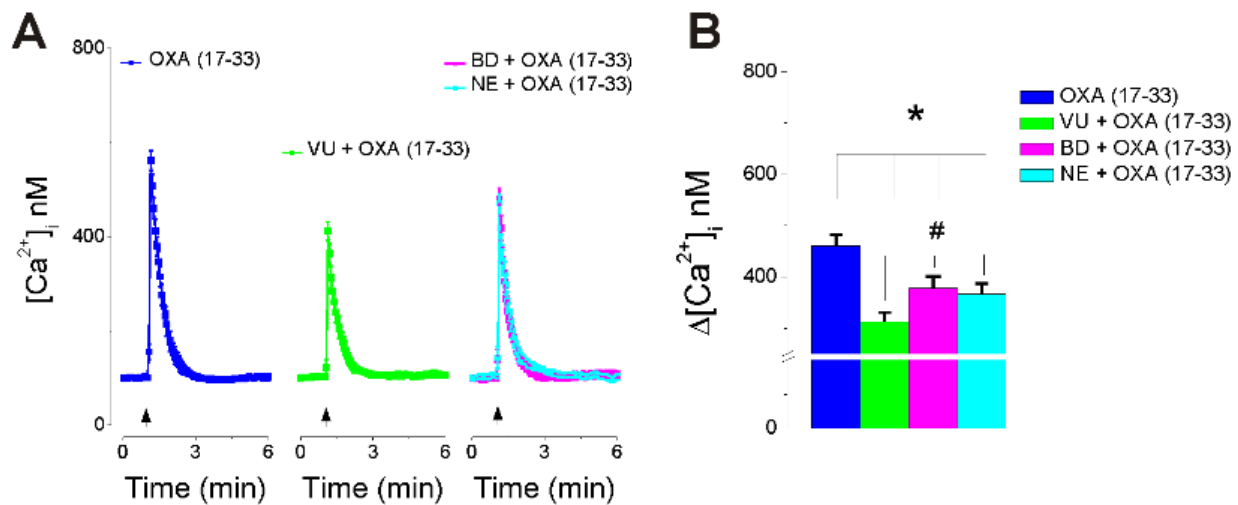


Figure 3. OXA (17-33) A increases $[Ca^{2+}]_i$ via choline-Sigma-1R-dependent mechanism. (A) Illustration of average Ca^{2+} transients (\pm SD) induced by OXA (17-33) (10 nM) alone (left) and in the presence of VU0155056 (1 μ M), PLD inhibitor (middle), and BD1047 (50 μ M) or NE-100 (5 μ M), Sigma-1R antagonists (right). (B) Comparison of the amplitude of the increase in $[Ca^{2+}]_i$ (average + SD) in each condition. Inhibition of PLD or antagonism of Sig-1R reduces the Ca^{2+} response elicited by OXA (17-33) (10 nM); $p < 0.05$ as compared to amplitude of $[Ca^{2+}]_i$ increase produced by OXA (17-33) (*) or produced in the presence of the inhibitors (#) ($n = 6$ neurons/condition).

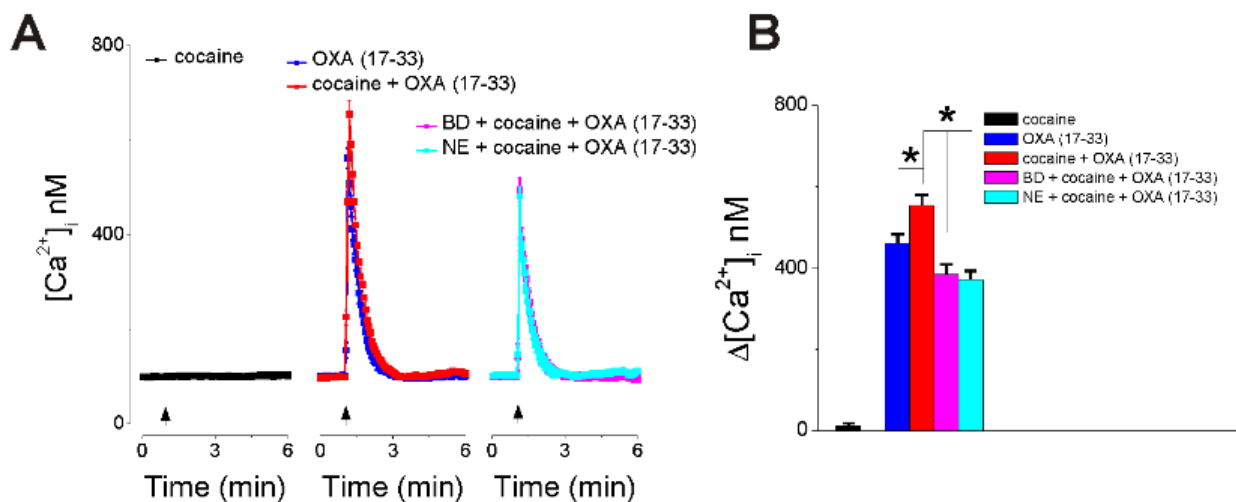


Figure 4. Cocaine potentiates OXA (17-33)-induced increase in $[Ca^{2+}]_i$ via Sigma-1R activation. (A) Illustration of average Ca^{2+} responses (\pm SD) produced by application of cocaine (10 μ M) alone (left, no response), cocaine (10 μ M) and OXA (17-33) (10 nM) (middle), and cocaine and OXA (17-33) in the presence of Sigma-1R antagonists BD1047 (50 μ M) or NE-100 (5 μ M) (right). (B) Comparison of the amplitude of the increase in $[Ca^{2+}]_i$ (average + SD) in each condition. Cocaine potentiates the Ca^{2+} response induced by OXA (17-33), while antagonism of Sigma-1R abolished the potentiation produced by cocaine on the Ca^{2+} response elicited by OXA (17-33) (10 nM). * $p < 0.05$ ($n = 6$ neurons/condition).

A diagram summarizing the proposed mechanism of potentiation of orexin by cocaine via Sigma-1R activation in nucleus accumbens neurons is illustrated in Figure 5.

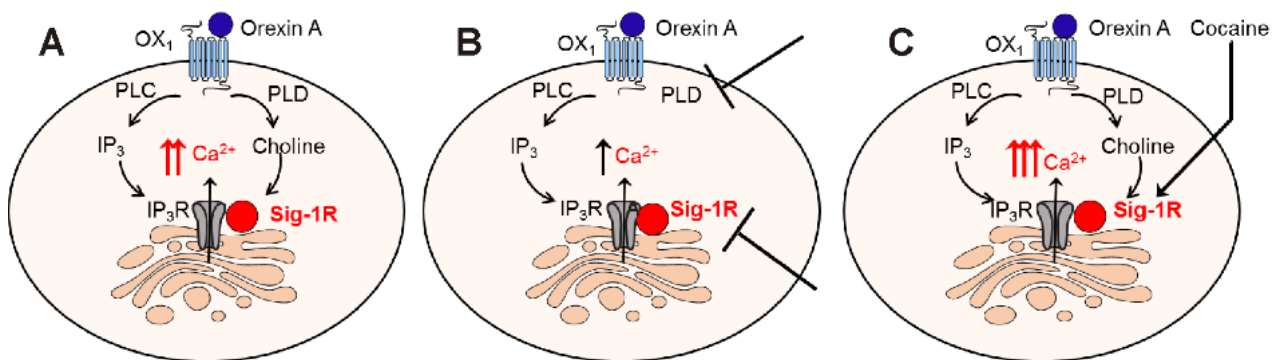


Figure 5. Diagram illustrating the proposed model of potentiation of orexin by cocaine via Sigma-1R in nucleus accumbens neurons. (A) Orexin A acting on OX₁ receptor activates PLC and PLD. PLC increases IP₃ level and promotes the Ca²⁺ release from endoplasmic reticulum via IP₃ receptors (IP₃R). PLD produces choline (from hydrolysis of phosphatidylcholine) that acts on Sigma-1R to potentiate Ca²⁺ increase via IP₃ R (higher increase in [Ca²⁺]_i). (B) Inhibition of PLD or antagonism of Sigma-1R limits the orexin A-OX₁ receptor signaling to PLC-mediated IP₃-dependent increase in Ca²⁺ (smaller increase in [Ca²⁺]_i). (C) Cocaine, via Sigma-1R activation, potentiates the PLC- and PLD-mediated increase in [Ca²⁺]_i produced by orexin A acting on OX₁ (highest increase in [Ca²⁺]_i). The diagram was created using the Motifolio Illustration Toolkit Neuroscience (<https://www.motifolio.com> accessed on 7 March 2021).

3. Discussion

Orexin A, via activation of OX₁ receptor, can activate both phospholipase C (PLC) and phospholipase D (PLD) in various cell models [9,10] including neurons [37]. PLC activation leads to hydrolysis of phosphoinositides and formation of inositol-1,4,5- trisphosphate (IP₃), the Ca²⁺-releasing second messenger that releases Ca²⁺ from endoplasmic reticulum (ER) through IP₃ receptors [38]. PLD activation promotes the hydrolysis of phosphatidylcholine to choline and phosphatidic acid [11]. Whereas phosphatidic acid was considered the main effector downstream to PLD activation, we recently identified choline as a second messenger that activates Sigma-1R [12].

Sigma-1 receptor is a chaperone protein expressed in the endoplasmic reticulum (ER), mainly at the mitochondria-associated ER membrane domains (MAMs) [13]. Sigma-1Rs interact with many different signaling proteins. At the ER, Sigma-1Rs potentiate the Ca²⁺ release via IP₃ receptors [13]; they also interact with STIM1, the Ca²⁺ sensor for store-operated Ca²⁺ entry [39]. Sigma-1R ligands include antidepressants, antipsychotics, and drugs of abuse [40]. Cocaine, in addition to its canonical target that elevates synaptic dopamine levels, binds to and activates Sigma-1Rs [41,42]. Neurons in the nucleus accumbens, a key area involved in the reward circuit [25], express Sigma-1R [27–29] and OX₁ receptors [14,15]. Behavioral studies supported the cocaine–orexin interaction in nucleus accumbens [3,22], but the underlying mechanisms remained unclear; this prompted us to investigate the mechanisms of cocaine–orexin interaction at this level.

Orexin A has been reported to increase cytosolic Ca²⁺ concentration, [Ca²⁺]_i, in various cells expressing orexin receptors [1], including neurons [43]. We first tested the effect of truncated orexin A peptide, OXA (17-33), a selective OX₁ agonist [30], on [Ca²⁺]_i in cultured nucleus accumbens neurons. OXA (17-33) increased [Ca²⁺]_i in a dose-dependent manner; the effect was abolished by SB-334867 (1 μM), an OX₁ antagonist [10,31] indicating that it was mediated by OX₁ receptors.

We next demonstrated that the OXA (17-33)-induced increase in [Ca²⁺]_i was mediated by IP₃-dependent Ca²⁺ release from ER, as previously reported [6]; the effect was abolished by IP₃ receptor antagonists, but not affected by disruption of lysosomal Ca²⁺ stores.

In other series of experiments, pretreatment with PLD inhibitor reduced the Ca²⁺ response elicited by OXA (17-33), supporting the involvement of PLD activation in addition to PLC/IP₃-dependent mechanisms in nucleus accumbens neurons. This is in agreement with previous studies reporting PLD-dependent mechanisms downstream to OX₁ activation [9,10,37].

In addition, antagonism of Sigma-1R reduced the Ca^{2+} response produced by OXA (17-33), indicating for the first time the role of Sigma-1R in the response to OX_1 activation in the nucleus accumbens. The reduction in the response to OXA (17-33) produced by PLD inhibition and Sigma-1R antagonism indicates that choline produced by PLD hydrolysis of phosphatidylcholine, acting on Sigma-1R, as recently reported [12], potentiates the IP_3 -mediated increase in $[Ca^{2+}]_i$.

Our results also indicate that cocaine, while it did not elicit a response by itself, potentiated the increase in $[Ca^{2+}]_i$ induced by OXA(17-33). This is similar to the potentiation of orexin A-induced increase in $[Ca^{2+}]_i$ by cocaine reported in VTA neurons [43]. In VTA neurons, the effect of orexin and the potentiation by cocaine were abolished by suvorexant (MK-4305), a dual orexin receptor OX_1/OX_2 antagonist [43,44]. Moreover, in nucleus accumbens neurons, the potentiation of orexin response by cocaine was abolished by Sigma-1R antagonists. Cocaine is a Sigma-1R agonist [26], and we previously reported that, in nucleus accumbens neurons, cocaine via Sigma-1R potentiates the IP_3 -mediated increase in $[Ca^{2+}]_i$ [29]. Here, we identify an additional signaling mechanism for orexin A– OX_1 via choline–Sigma-1R and a critical role for Sigma-1R in the cocaine–orexin A interaction in nucleus accumbens neurons.

4. Materials and Methods

4.1. Chemicals

OXA (17-33), i.e., truncated orexin A, a selective OX_1 agonist [30], SB-334867, a selective nonpeptide OX_1 antagonist [31], and BD-1047 and NE-100 (Sigma-1 antagonists) were obtained from Tocris (Bio-Techne Corporation, Minneapolis, MN, USA). VU0155056, a PLD inhibitor [34], was purchased from Avanti Polar Lipids (Alabaster, AL, USA). Cocaine was supplied by the National Institute on Drug Abuse's Drug Supply Program. All other chemicals were from Sigma Aldrich (St. Louis, MO, USA), unless otherwise mentioned.

4.2. Neuronal Cell Culture

Nucleus accumbens neurons were dissociated from neonatal Sprague Dawley rats (Ace Animal Inc., Boyertown, PA, USA) of both sexes as previously described [29,45]. Newborn rats were decapitated, and the brains quickly removed surgically and immersed in ice-cold Hanks balanced salt solution (HBSS). The nucleus accumbens was identified, removed, minced, and subjected to enzymatic (papain, 37 °C) and mechanical dissociation. Cells were cultured in Neurobasal A medium (Life Technologies, ThermoFisher Scientific, Carlsbad, CA, USA) containing 10% fetal bovine serum, 1% GlutaMax, and 1% penicillin–streptomycin–amphotericin B solution at 37 °C in a humidified atmosphere with 5% CO_2 . The mitotic inhibitor cytosine β -arabino-furanoside (1 μ M) was added to the culture to inhibit glial cell proliferation. For calcium imaging, neurons were cultured on round 25 mm diameter glass coverslips coated with poly-L-lysine, in six-well plates.

4.3. Measurement of Cytosolic Ca^{2+} Concentration

Cytosolic Ca^{2+} concentration, $[Ca^{2+}]_i$, was measured by calcium imaging methods in nucleus accumbens neurons loaded with Fura-2AM, as previously described [29,45]. Cells were incubated with 5 μ M Fura-2AM (Invitrogen) in HBSS at room temperature for 45 min, in the dark, and then incubated for another 45 min in HBSS to allow for complete de-esterification of the dye. Coverslips (25 mm diameter) were subsequently mounted in an open bath chamber (Warner Instruments, Hamden, CT, USA) on the stage of an inverted microscope Nikon Eclipse TiE (Nikon Inc., Melville, NY, USA), equipped with a Perfect Focus System and a Photometrics CoolSnap HQ2 CCD camera (Photometrics, Tucson, AZ, USA). During the experiments, the Perfect Focus System was activated. Fura-2AM fluorescence (emission = 510 nm), following alternate excitation at 340 and 380 nm, was acquired at a frequency of 0.25 Hz. Images were acquired and analyzed using NIS-Elements AR software (Nikon Inc.). After appropriate calibration with ionomycin and $CaCl_2$ and

with Ca²⁺ free and EGTA, respectively, the ratio of the fluorescence signals (340/380 nm) was converted to Ca²⁺ concentrations [46].

4.4. Data Analysis

Data were expressed as the mean \pm standard deviation (SD). Datasets were compared for statistically significant differences using one-way ANOVA followed by post hoc Bonferroni test. A p -value <0.05 was considered statistically significant.

Author Contributions: Conceptualization, E.B.; methodology, J.L.B., P.Z., G.C.B., and E.B.; validation, J.L.B., G.C.B., and E.B.; formal analysis, J.L.B., G.C.B., and E.B.; investigation, J.L.B., P.Z., G.C.B., and E.B.; writing—original draft preparation, G.C.B. and E.B.; writing—review and editing, J.L.B., P.Z., G.C.B., and E.B.; visualization, J.L.B., P.Z., G.C.B., and E.B.; supervision, E.B. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the National Institutes of Health grant number P30DA013429 and research funds from the Jefferson College of Pharmacy.

Institutional Review Board Statement: Animal protocols were approved by the Institutional Animal Care and Use Committee (protocol 01460 approved 20 January 2019).

Informed Consent Statement: Not applicable.

Data Availability Statement: The data generated and analyzed during this study are available in the manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Sakurai, T.; Amemiya, A.; Ishii, M.; Matsuzaki, I.; Chemelli, R.M.; Tanaka, H.; Williams, S.C.; Richardson, J.A.; Kozlowski, G.P.; Wilson, S.; et al. Orexins and orexin receptors: A family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. *Cell* **1998**, *92*, 573–585. [[CrossRef](#)]
2. de Lecea, L.; Kilduff, T.S.; Peyron, C.; Gao, X.; Foye, P.E.; Danielson, P.E.; Fukuhara, C.; Battenberg, E.L.; Gautvik, V.T.; Bartlett, F.S.; et al. The hypocretins: Hypothalamus-specific peptides with neuroexcitatory activity. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 322–327. [[CrossRef](#)] [[PubMed](#)]
3. Mahler, S.V.; Smith, R.J.; Moorman, D.E.; Sartor, G.C.; Aston-Jones, G. Multiple roles for orexin/hypocretin in addiction. *Prog. Brain Res.* **2012**, *198*, 79–121.
4. Haghparast, A.; Fatahi, Z.; Arezoomandan, R.; Karimi, S.; Taslimi, Z.; Zarrabian, S. Functional roles of orexin/hypocretin receptors in reward circuit. *Prog. Brain Res.* **2017**, *235*, 139–154.
5. Foord, S.M.; Bonner, T.I.; Neubig, R.R.; Rosser, E.M.; Pin, J.-P.; Davenport, A.P.; Spedding, M.; Harmar, A.J. International Union of Pharmacology. XLVI. G protein-coupled receptor list. *Pharmacol. Rev.* **2005**, *57*, 279–288. [[CrossRef](#)] [[PubMed](#)]
6. Kukkonen, J.P. G-protein-dependency of orexin/hypocretin receptor signalling in recombinant Chinese hamster ovary cells. *Biochem. Biophys. Res. Commun.* **2016**, *476*, 379–385. [[CrossRef](#)]
7. Hopf, F.W. Recent perspectives on orexin/hypocretin promotion of addiction-related behaviors. *Neuropharmacology* **2020**, *168*, 108013. [[CrossRef](#)] [[PubMed](#)]
8. Kukkonen, J.P. Lipid signaling cascades of orexin/hypocretin receptors. *Biochimie* **2014**, *96*, 158–165. [[CrossRef](#)]
9. Johansson, L.; Ekholm, M.E.; Kukkonen, J.P. Multiple phospholipase activation by OX(1) orexin/hypocretin receptors. *Cell. Mol. Life Sci.* **2008**, *65*, 1948–1956. [[CrossRef](#)]
10. Jantti, M.H.; Putula, J.; Somerharju, P.; Frohman, M.A.; Kukkonen, J.P. OX1 orexin/hypocretin receptor activation of phospholipase D. *Br. J. Pharmacol.* **2012**, *165*, 1109–1123. [[CrossRef](#)] [[PubMed](#)]
11. Exton, J.H. Phospholipase D. *Ann. N. Y. Acad. Sci.* **2000**, *905*, 61–68. [[CrossRef](#)]
12. Brailoiu, E.; Chakraborty, S.; Brailoiu, G.C.; Zhao, P.; Barr, J.L.; Ilies, M.A.; Unterwald, E.M.; Abood, M.E.; Taylor, C.W. Choline is an intracellular messenger linking extracellular stimuli to IP3-evoked Ca(2+) signals through Sigma-1 receptors. *Cell Rep.* **2019**, *26*, 330–337. [[CrossRef](#)] [[PubMed](#)]
13. Hayashi, T.; Su, T.P. Sigma-1 receptor chaperones at the ER-mitochondrion interface regulate Ca(2+) signaling and cell survival. *Cell* **2007**, *131*, 596–610. [[CrossRef](#)] [[PubMed](#)]
14. D’Almeida, V.; Hipolide, D.C.; Raymond, R.; Barlow, K.B.L.; Parkes, J.H.; Pedrazzoli, M.; Tufik, S.; Nobrega, J.N. Opposite effects of sleep rebound on orexin OX1 and OX2 receptor expression in rat brain. *Brain Res. Mol. Brain Res.* **2005**, *136*, 148–157. [[CrossRef](#)]
15. Lei, K.; Kwok, C.; Darevsky, D.; Wegner, S.A.; Yu, J.; Nakayama, L.; Pedrozo, V.; Anderson, L.; Ghotra, S.; Fouad, M.; et al. Nucleus accumbens shell Orexin-1 receptors are critical mediators of binge intake in excessive-drinking individuals. *Front. Neurosci.* **2019**, *13*, 88. [[CrossRef](#)]

16. Perrey, D.A.; Zhang, Y. Therapeutics development for addiction: Orexin-1 receptor antagonists. *Brain Res.* **2020**, *1731*, 145922. [[CrossRef](#)]
17. Han, Y.; Yuan, K.; Zheng, Y.; Lu, L. Orexin receptor antagonists as emerging treatments for psychiatric disorders. *Neurosci. Bull.* **2020**, *36*, 432–448. [[CrossRef](#)]
18. Khoo, S.Y.; Brown, R.M. Orexin/hypocretin based pharmacotherapies for the treatment of addiction: DORA or SORA? *CNS Drugs* **2014**, *28*, 713–730. [[CrossRef](#)]
19. Bentzley, B.S.; Aston-Jones, G. Orexin-1 receptor signaling increases motivation for cocaine-associated cues. *Eur. J. Neurosci.* **2015**, *41*, 1149–1156. [[CrossRef](#)]
20. Zhou, L.; Ghee, S.M.; Chan, C.; Lin, L.; Cameron, M.D.; Kenny, P.J.; See, R.E. Orexin-1 receptor mediation of cocaine seeking in male and female rats. *J. Pharmacol. Exp. Ther.* **2012**, *340*, 801–809. [[CrossRef](#)]
21. Prince, C.D.; Rau, A.R.; Yorgason, J.T.; Espana, R.A. Hypocretin/Orexin regulation of dopamine signaling and cocaine self-administration is mediated predominantly by hypocretin receptor 1. *ACS Chem. Neurosci.* **2015**, *6*, 138–146. [[CrossRef](#)] [[PubMed](#)]
22. Yang, M.; Ma, H.; Jia, M.; Li, Y.; Miao, D.; Cui, C.; Wu, L. The role of the nucleus accumbens OXR1 in cocaine-induced locomotor sensitization. *Behav. Brain Res.* **2020**, *379*, 112365. [[CrossRef](#)] [[PubMed](#)]
23. Martin-Fardon, R.; Weiss, F. Blockade of hypocretin receptor-1 preferentially prevents cocaine seeking: Comparison with natural reward seeking. *Neuroreport* **2014**, *25*, 485–488. [[CrossRef](#)] [[PubMed](#)]
24. Giros, B.; Jaber, M.; Jones, S.R.; Wightman, R.M.; Geron, M.G. Hyperlocomotion and indifference to cocaine and amphetamine in mice lacking the dopamine transporter. *Nature* **1996**, *379*, 606–612. [[CrossRef](#)]
25. Koob, G.F.; Volkow, N.D. Neurocircuitry of addiction. *Neuropsychopharmacology* **2010**, *35*, 217–238. [[CrossRef](#)]
26. Sharkey, J.; Glen, K.A.; Wolfe, S.; Kuhar, M.J. Cocaine binding at sigma receptors. *Eur. J. Pharmacol.* **1988**, *149*, 171–174. [[CrossRef](#)]
27. Gundlach, A.L.; Largent, B.L.; Snyder, S.H. Autoradiographic localization of sigma receptor binding sites in guinea pig and rat central nervous system with (+)3H-3-(3-hydroxyphenyl)-N-(1-propyl)piperidine. *J. Neurosci.* **1986**, *6*, 1757–1770. [[CrossRef](#)]
28. Alonso, G.; Phan, V.; Guillemain, I.; Saunier, M.; Legrand, A.; Anoa, M.; Maurice, T. Immunocytochemical localization of the sigma(1) receptor in the adult rat central nervous system. *Neuroscience* **2000**, *97*, 155–170. [[CrossRef](#)]
29. Barr, J.L.; Deliu, E.; Brailoiu, C.G.; Zhao, P.; Yan, G.; Abood, M.E.; Unterwald, E.M.; Brailoiu, E. Mechanisms of activation of nucleus accumbens neurons by cocaine via sigma-1 receptor-inositol 1,4,5-trisphosphate-transient receptor potential canonical channel pathways. *Cell Calcium* **2015**, *58*, 196–207. [[CrossRef](#)]
30. Feng, X.M.; Mi, W.L.; Xia, F.; Mao-Ying, W.L.; Jiang, J.W.; Xiao, S.; Wang, Z.F.; Wang, Y.Q.; Wu, G.C. Involvement of spinal orexin A in the electroacupuncture analgesia in a rat model of post-laparotomy pain. *BMC Complement. Altern. Med.* **2012**, *12*, 225. [[CrossRef](#)]
31. Smart, D.; Sabido-David, C.; Brough, S.J.; Jewitt, F.; Johns, A.; Porter, R.A.; Jerman, J.C. SB-334867-A: The first selective orexin-1 receptor antagonist. *Br. J. Pharmacol.* **2001**, *132*, 1179–1182. [[CrossRef](#)]
32. Maruyama, T.; Kanaji, T.; Nakade, S.; Kanno, T.; Mikoshiba, K. 2APB, 2-aminoethoxydiphenyl borate, a membrane-penetrable modulator of Ins(1,4,5)P₃-induced Ca²⁺ release. *J. Biochem.* **1997**, *122*, 498–505. [[CrossRef](#)] [[PubMed](#)]
33. Bowman, E.J.; Siebers, A.; Altendorf, K. Bafilomycins: A class of inhibitors of membrane ATPases from microorganisms, animal cells, and plant cells. *Proc. Natl. Acad. Sci. USA* **1988**, *85*, 7972–7976. [[CrossRef](#)]
34. Scott, S.A.; Selvy, P.E.; Buck, J.R.; Cho, H.P.; Criswell, T.L.; Thomas, A.L.; Armstrong, M.D.; Arteaga, C.L.; Lindsley, C.W.; Brown, H.A. Design of isoform-selective phospholipase D inhibitors that modulate cancer cell invasiveness. *Nat. Chem. Biol.* **2009**, *5*, 108–117. [[CrossRef](#)]
35. Matsumoto, R.R.; Bowen, W.D.; Tom, M.A.; Vo, V.N.; Truong, D.D.; de Costa, B.R. Characterization of two novel sigma receptor ligands: Antidystonic effects in rats suggest sigma receptor antagonism. *Eur. J. Pharmacol.* **1995**, *280*, 301–310. [[CrossRef](#)]
36. Chaki, S.; Okuyama, S.; Ogawa, S.; Tanaka, M.; Muramatsu, M.; Nakazato, A.; Tomisawa, K. Solubilization and characterization of binding sites for [³H]NE-100, a novel and potent sigma 1 ligand, from guinea pig brain. *Life Sci.* **1996**, *59*, 1331–1340. [[CrossRef](#)]
37. Bjornstrom, K.; Turina, D.; Strid, T.; Sundqvist, T.; Eintrei, C. Orexin A inhibits propofol-induced neurite retraction by a phospholipase D/protein kinase Cepsilon-dependent mechanism in neurons. *PLoS ONE* **2014**, *9*, e97129. [[CrossRef](#)]
38. Berridge, M.J. The Inositol Trisphosphate/Calcium signaling pathway in health and disease. *Physiol. Rev.* **2016**, *96*, 1261–1296. [[CrossRef](#)]
39. Srivats, S.; Balasuriya, D.; Pasche, M.; Vistal, G.; Edwardson, J.M.; Taylor, C.W.; Murell-Lagnado, R.D. Sigma1 receptors inhibit store-operated Ca²⁺ entry by attenuating coupling of STIM1 to Orai1. *J. Cell Biol.* **2016**, *213*, 65–79. [[CrossRef](#)]
40. Maurice, T.; Su, T.P. The pharmacology of sigma-1 receptors. *Pharmacol. Ther.* **2009**, *124*, 195–206. [[CrossRef](#)]
41. Katz, J.L.; Hong, W.C.; Hiranita, T.; Su, T.P. A role for sigma receptors in stimulant self-administration and addiction. *Pharmacother.* **2011**, *4*, 880–914. [[CrossRef](#)]
42. Hayashi, T.; Tsai, S.Y.; Mori, T.; Fujimoto, M.; Su, T.P. Targeting ligand-operated chaperone sigma-1 receptors in the treatment of neuropsychiatric disorders. *Expert Opin. Ther. Targets* **2011**, *15*, 557–577. [[CrossRef](#)] [[PubMed](#)]
43. Gentile, T.A.; Simmons, S.J.; Watson, M.N.; Connelly, K.L.; Brailoiu, E.; Zhang, Y.; Muschamp, J.W. Effects of suvorexant, a dual Orexin/Hypocretin receptor antagonist, on impulsive behavior associated with cocaine. *Neuropsychopharmacology* **2018**, *43*, 1001–1009. [[CrossRef](#)]

44. Cox, C.D.; Breslin, M.J.; Whitman, D.B.; Schreier, J.D.; McGaughey, G.B.; Bogusky, M.J.; Roecker, A.J.; Mercer, S.P.; Bednar, R.A.; Lemaire, W.; et al. Discovery of the dual orexin receptor antagonist [(7R)-4-(5-chloro-1,3-benzoxazol-2-yl)-7-methyl-1,4-diazepan-1-yl][5-methyl-2-(2H-1,2,3-triazol-2-yl)phenyl]methanone (MK-4305) for the treatment of insomnia. *J. Med. Chem.* **2010**, *53*, 5320–5332. [[CrossRef](#)]
45. Brailoiu, G.C.; Deliu, E.; Barr, J.L.; Console-Bram, L.M.; Ciuciu, A.M.; Abood, M.E.; Unterwald, E.M.; Brailoiu, E. HIV Tat excites D1 receptor-like expressing neurons from rat nucleus accumbens. *Drug Alcohol Depend.* **2017**, *178*, 7–14. [[CrossRef](#)] [[PubMed](#)]
46. Gryniewicz, G.; Poenie, M.; Tsien, R.Y. A new generation of Ca²⁺ indicators with greatly improved fluorescence properties. *J. Biol. Chem.* **1985**, *260*, 3440–3450. [[CrossRef](#)]