Trametinib prevents mesothelial-mesenchymal transition and ameliorates abdominal adhesion formation

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Trametinib prevents mesothelial-mesenchymal transition and ameliorates abdominal adhesion formation

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ABSTRACT

Background: Intra-abdominal adhesions are a major cause of morbidity after abdominal or gynecologic surgery. However, knowledge about the pathogenic mechanism(s) is limited, and there are no effective treatments. Here, we investigated a mouse model of bowel adhesion formation and the effect(s) of an Federal Drug Administration-approved drug (trametinib) in preventing adhesion formation.

Materials and methods: C57BL/6 mice were used to develop a consistent model of intra-abdominal adhesion formation by gentle cecal abrasion with mortality rates of <10%. Adhesion formation was analyzed histologically and immunochemically to characterize the expression of pro-fibrotic marker proteins seen in pathologic scaring and included alpha smooth muscle actin (αSMA) and fibronectin EDA (FNEDA) which arises from alternative splicing of the fibronectin messenger RNA resulting in different protein isoforms. Trichrome staining assessed collagen deposition. Quantitative polymerase chain reaction analysis of RNA isolated from adhesions by laser capture microscopy was carried out to assess pro-fibrotic gene expression. To block adhesion formation, trametinib was administered via a subcutaneous osmotic pump.

Results: Adhesions were seen as early as post-operative day 1 with extensive adhesions being formed and vascularized by day 5. The expression of the FNEDA isoform occurred first with subsequent expression of αSMA and collagen. The drug trametinib was chosen for in vivo studies because it effectively blocked the mesothelial to mesenchymal transition of rat mesothelium. Trametinib, at the highest dose used (3 mg/kg/d), prevented adhesion formation while at lower doses, adhesions were usually limited, as evidenced by the presence of FNEDA isoform but not αSMA.
Introduction

Although peritoneal adhesions may be caused by infection, inflammation, or ischemia, surgical procedures are the primary cause since greater than 90% of patients will develop adhesions after abdominal surgery. In the United States between 1998 and 2002, over 18% of hospital admissions were secondary to abdominal adhesions alone resulting in an estimated cost of 1.18 billion dollars. Such adhesions are responsible for pelvic pain, bowel obstruction, and infertility. Although modern advances in surgical technique, including laparoscopy, have led to a decrease in their incidence, intestinal adhesions still pose a very significant medical as well as economic problem. Unfortunately, adequate therapeutic solutions have proven elusive. While there are some studies using therapeutics to attempt to prevent adhesion formation, these have largely not gained wide acceptance. Similarly, the use of barriers has also produced mixed results. Once formed, adhesions are removed by adhesiolysis surgery. More recent studies have shown that adhesiolysis procedures in the United States account for 967,332 d of care at a cost of 2.3 billion dollars.

Adhesion formation has a complex pathogenesis and can be broken down into several stages which are as follows.

1. An inflammatory response, whose blockage has largely proven to be unsuccessful. It is characterized by an influx of multiple cell types and production of a variety of cytokines and factors.

2. The coagulation cascade and clot formation whose characterization is critical to understand adhesion pathogenesis. Multiple studies provide a rational basis for enhancing clot lysis as a therapeutic strategy. However, in practice, this has proven difficult. These events occur early postoperatively, and the failure of fibrinolysis allows cellular infiltration of the initial fibrinous matrix. Although, in many cases, the formation of a clot is essential to limit injury, resolution of the clot, in a timely manner, is necessary to prevent adhesion formation. Thus, the balance between fibrin clot formation and its lysis is critical.

3. The final stage in the adhesion process is formation of a connective tissue scar. By and large, this stage causes the most severe complications and has many common features with fibrotic reactions found elsewhere in the body, including systemic ones (e.g., scleroderma) and those affecting individual organs including lung, heart, liver, and kidney.

Because the pathophysiology of fibrotic reactions has received insufficient attention, there exists an urgent need for cellular and molecular characterization of adhesion formation. The critical cell in the formation of an adhesion is the myofibroblast which produces increased amounts of fibrillar collagens as well as other matrix proteins and which expresses alpha smooth muscle actin (αSMA), a molecular marker of activated myofibroblasts. Although the origins of myofibroblasts may differ depending on the affected organ and the initiating event, in the abdominal cavity, they may arise through a process of trans-differentiation of mesothelial cells in which these cells lose their specific epithelial phenotypic markers such as expression of E-cadherin and acquire a mesenchymal or myofibroblast phenotype. This change in mesothelial phenotype has been termed mesothelial-mesenchymal transition (MMT).

Since its first identification, it has been known that transforming growth factor-β (TGF-β), a pleiotropic growth factor with a wide and diverse spectrum of biological activities, plays a key role in fibrotic diseases by mediating the formation of myofibroblasts and stimulating the production of extracellular matrix (ECM). In addition to TGF-β, interleukin-6, another pleiotropic cytokine with a diverse range of biological activities, was also found to be elevated in peritoneal fluid during and/or after abdominal surgeries thus potentially implicating it in the cascade of events which lead to adhesion formation. Significantly, the levels of these cytokines correlated with the severity of abdominal adhesion formation. The complex signaling pathways activated by TGF-β involve both canonical and noncanonical signaling pathways. In the present context, the critical downstream event elicited by noncanonical signaling is the mitogen activated protein kinase (MEK) activation of Erk1/2, which, when activated by phosphorylation, enters the nucleus and, in association with other factors, mediates the transcription of pro-fibrotic genes and cell cycle regulatory proteins.

We have previously found that U0126, a MEK1/2 inhibitor not in clinical use, blocked the rat peritoneal MMT induced by TGF-β. To extend these findings to a drug with clinical potential, in the present study we have evaluated the effect of the MEK1/2 inhibitor, trametinib, currently in clinical use in the treatment of malignant melanoma in humans, on the TGF-β-induced rat peritoneal MMT and abdominal adhesion formation in a mouse model. Trametinib effectively blocked the MMT in vivo and markedly diminished adhesion formation, likely by inhibiting the activation of Erk1/2. Taken together, these findings indicate that trametinib may be a useful drug for the inhibition of adhesion formation and warrant human clinical studies.

The goals of the present study were to use the mouse cecal abrasion model to characterize the temporal pro-fibrotic
cellular and molecular events that occur during adhesion formation and determine whether the drug trametinib, a MEK1/2 inhibitor, can block and/or alter these events.

**Materials and methods**

**Reagents and antibodies**

All reagents, unless otherwise specified, were purchased from Sigma (St. Louis, MO). Polyvinylidene difluoride membrane was purchased from Roche Diagnostics, Basel, Switzerland. Antibodies to: Phospho-MEK1/2 (Ser217/221) #9154, Phospho-Smad2 (Ser465/467) #3108, Phospho-Smad2 (Ser245/250/255) #3104, Phospho-p44/42 MAPK (Erk1/2) (Thr202/Tyr204)#4370 were purchased from Cell Signaling Technology, Danvers, MA. Antibody to the EDA isoform of rat fibronectin (FNEDA) was developed and characterized by one of us, and aSMA #ab5694 was purchased from Abcam, Cambridge, MA. Trametinib was purchased from LC Laboratories, Woburn, MA.

**Isolation and culture of rat peritoneal mesothelial cells**

The experiments in this study were approved by the Institutional Animal Care and Use Committee at Thomas Jefferson University and were performed in accordance with the National Institutes of Health guidelines for the care and

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Fig. 1 – TGF-β and interleukin-6 pathways leading to activation of MEK1/2 and Erk1/2 and their inhibition by trametinib. Both TGF-β and interleukin-6 have been shown to activate a “downstream” common signaling pathway through Ras, which results in the activation of Erk1/2. Erk1/2 has many potential downstream activities, but only their role(s) as transcription factors is illustrated. Although the relevant pathways have been simplified for clarity, the figure shows the most important elements in the present context. TGF-β = transforming growth factor-β; LAP = latency associated peptide, GP = glycoprotein; SARA = SMAD anchor for receptor activation; SMAD = SMA and MAD related family of signal transducers; CBP = CREB binding protein; TF = transcription factor; SHC = SHC transforming protein, an adapter protein that couples growth factor receptors to signaling pathways; GRB = growth factor receptor bound protein. (Color version of figure is available online.) This figure has been modified from two produced by ProteinLounge.
handling of laboratory animals. Rat peritoneal mesothelial cells (RPMCs) were isolated and cultured as described previously. Briefly, Sprague Dawley rats weighing 150g-250 g, purchased from Jackson Laboratory, were injected intraperitoneally with 30 mL of 0.25% trypsin/2.21 mM EDTA under isoflurane anesthesia and were kept on the metal pad warmed to 37°C for 1 h after which the abdominal fluid was collected and centrifuged at 300 x g for 10 min. The pelleted cells were resuspended and cultured in DMEM/F12 medium (ThermoFisher Scientific, Philadelphia, PA) supplemented with 10% (v/v) fetal bovine serum (ThermoFisher Scientific) at 37°C in a humidified atmosphere of 5% CO₂ in air. The RPMCs, from the fourth to seventh passages (split ratio 1:4), at 90% confluence were used for the experiments. The cells were treated either with 10 ng/mL of TGF-β1 (R&D systems, Minneapolis, MN) alone or with TGF-β1 and the MEK1/2 inhibitor, trametinib (2 or 5 nM).

Cecal abrasion model

Equal numbers of male and female C57BL/6 mice (18-25g, 8-10 wk of age, Jackson Laboratories, Bar Harbor, Maine) were used in initial experiments while only male mice were used in the drug escalation studies because they sustained greater accumulations of fibrotic tissue (data not shown). Mice were allowed to acclimate in the animal facility for at least 1 wk before surgery, given free access to standard chow and water, and a 12-h light-dark cycle in standard acrylic cages with wood chip bedding. Animals were randomly assigned into either an experimental group (laparotomy and cecal abrasion) or a control group (laparotomy only).

Briefly, mice underwent induction and maintenance anesthesia with 1%-3% isoflurane with supplemental oxygen. After adequate sedation was achieved, mice were weighed, and 0.1 mg/kg subcutaneous Buprenex (Hospira, Inc, Lake Forest, IL) was administered to ensure analgesia. Animals were treated with three different doses of the drug trametinib dissolved in dimethyl sulfoxide (DMSO) in a dose escalation study. Groups of five animals were given 0.1, 1.0, or 3.0 mg/kg animal weight of drug/d via osmotic pumps (Alzet Osmotic Pump 1002, Cupertino, CA) for 8 d before sacrifice. The volume delivered/d was 6 μL of drug. Control mice underwent induction with anesthesia and laparotomy only. In addition, five animals underwent laparotomy and placement of the osmotic pumps which were filled with “drug vehicle” (DMSO) alone, that is, no drug. After 8 d of drug or vehicle treatment, mice were euthanized by CO₂ asphyxiation followed by cervical dislocation. The entire large intestine and cecum were removed and partitioned for histology and immunofluorescence microscopy. Adhesions were examined by 2 independent practitioners.

Histology

Bowel and abdominal wall involved in the adhesion were removed en bloc and fixed in 4% buffered formalin. Bowel from nondrug-treated mice was also taken as a control. Tissues were dehydrated, embedded in paraffin, and sectioned at either 5 or 10 microns (μ). Sections were deparaffinized in a graded ethanol series and stained with Masson’s trichrome stain. Three complete bowel segments (5 for 3-mg dosed mice) for each drug dose (0.1, 1.0, and 3.0 mg/kg/d) were serially sectioned and all regions where bowel segments were either close to one another or touching were photographed using phase microscopy, and selected sections were then subsequently stained with trichrome and/or processed for immunohistochemistry. Photographs were taken with a Zeiss light microscope equipped with a Nikon digital camera.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Forward primer (5’-3’)</th>
<th>Reverse primer (5’-3’)</th>
</tr>
</thead>
<tbody>
<tr>
<td>18S</td>
<td>ACCAGACGCGAAGACATTGGCCA</td>
<td>TCGCATGTTTTATGTCGGAA</td>
</tr>
<tr>
<td>Col1α1</td>
<td>GCATGGCCAAAGAGACATGG</td>
<td>TCCAGCTCGCTACGATGGGG</td>
</tr>
<tr>
<td>Col3α1</td>
<td>AGCTTTTGGCAAAAGTGGAACTCTGG</td>
<td>CAAATGCGCTGGATCCCAATTCTCAT</td>
</tr>
<tr>
<td>Fn1</td>
<td>TCCAGGACACACAGCATGTCGTA</td>
<td>CCAACGTCGGCTGGCAACCTTCA</td>
</tr>
<tr>
<td>Fn-EDA</td>
<td>TAAAGGACTGCCATTCGTGGA</td>
<td>GTGAAAGGCAACACCACCGTGC</td>
</tr>
<tr>
<td>α-SMA</td>
<td>GAGCTCTCTTCCAGGCTCCTTTC</td>
<td>GACAGCAAGGTGTTAAGTGA</td>
</tr>
</tbody>
</table>

Col1α1 = collagen type I alpha chain; Col3α1 = collagen type III alpha chain; Fn-EDA = fibronectin splicing isoform EDA; Fn1 = plasma fibronectin; α-SmA = α-smooth muscle actin.
Immunofluorescence microscopy

Bowel tissue was placed in Tissue-Tek O.C.T. Compound (Sakura Finetek, Torrance, CA) and immediately frozen in liquid nitrogen. Frozen sections were cut at either 5 or 10 μm, allowed to adhere to albumin-coated slides and then washed with phosphate buffered saline (PBS), followed by double staining with goat anti-αSMA polyclonal antibody (1:100), and anti-FnEDA antibody (1:75) after prior fixation in cold (4°C) methanol overnight at 4°C. After washing 3X with PBS, species-matched Alexa-Fluor secondary antibodies (Invitrogen, Carlsbad, CA) were added and incubated for 1 h at room temperature followed by three washes with PBS. Slides were mounted with 4′,6-diamino-2-phenylindole (DAPI) Fluoromount-G (Southern Biotech, Birmingham, AL), and fluorescence images were taken either with a Zeiss epifluorescence microscope (5μm sections) or a Nikon confocal microscope (10μm sections). Controls included: (1) omitting the primary antibody and replacing it either with saline or indifferent IgG from a control animal and (2) omission of the secondary antibody. In all instances, controls were either negative or showed very slight nonspecific staining with the secondary antibody alone (Fig. 7B).

RNA isolation and real-time PCR

Tissue from selected adhesions was isolated by laser capture microscopy affixed to membranes which were placed in lysis buffer using a protocol provided by the instrument manufacturer (Arcturus, ThermoFisher Scientific). Subsequently, RNA was isolated using the RNaseq Mini Kit (Qiagen, Valencia, CA). Isolated RNA was reverse transcribed using All-in-One cDNA Synthesis SuperMix (Bimake, Houston, TX) to generate first-strand complementary DNA. Transcript levels of pro-fibrotic genes associated with fibrosis and myofibroblast differentiation were determined using SYBR Green real-time PCR as previously described.36,37 Primers were designed using Primer Quest (Integrated DNA Technologies, Skokie, Illinois) and were validated for specificity. The sequences of the primers employed are shown in Table 1. The differences in the number of mRNA copies in each PCR were corrected for 18S RNA endogenous control transcript levels; expression levels in day 5 mice were set at 100%, and all other values were expressed as normalized multiples of these values.

Western blotting analysis

RPMCs were lysed in ice-cold modified RIPA buffer with protease inhibitor cocktail (50 mM/L Tris-HCl, 1% NP-40, 0.25% sodium deoxycholate, 150 mM/L NaCl, 1 mM/L EDTA, 1 mM/L phenylmethyl sulfonyl fluoride, 1 mM/L sodium orthovanadate, 1 mM/L sodium fluoride, pH 7.4). Equivalent
amounts of homogenate (50 μg/well), determined by Coomassie blue assay, were separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis, transferred to either nitrocellulose or polyvinylidene difluoride membranes, and detected by either SuperSignal West Femto or SuperSignal West Pico chemiluminescence substrate (ThermoFisher Scientific).

**Results**

### Effect of trametinib on mesothelial/mesenchymal transition

Isolated mesothelial cells were incubated under control conditions without either TGF-β or trametinib, with TGF-β or trametinib alone, or with both TGF-β and trametinib for 5 d (Fig. 2A). We have previously found that this length of time was required for maximal mesothelial/mesenchymal transitional effect of TGF-β on these rat cells. As before, TGF-β produced a dramatic transitional effect (upper right panel), markedly altering the appearance of the cells from their normal morphology (upper left panel), whereas trametinib alone had no effect and no apparent toxic effects with the cells maintaining a cobblestone appearance (lower left panel). Remarkably, trametinib blocked the effect of TGF-β, and the cells retained their epithelial morphology (lower right panel).

Based on our early observations (data not shown), TGF-β–treated cells gained αSMA and COL1A1 expression, which was prevented by treatment with 5 nM trametinib. These insights led us to explore the potential mechanisms of action of trametinib using Western blotting analyses (Fig. 2B). These experiments demonstrated that TGF-β produced a substantial increase in the phosphorylation/activation of Erk1/2 and phosphorylation of the Smad2 linker region as well as increases in expression of αSMA and FNEDA. These increases in the expression of p-Erk1/2, FNEDA, αSMA, and p-SMAD2 (linker) were blocked by as low a concentration of trametinib as 2 nM (Fig. 2B).
Characterization of adhesion formation in the mouse model

To characterize adhesion formation in the mouse model, mice sacrificed at post-surgery time points after cecal abrasion were examined by both trichrome staining and immunohistochemistry using antibodies raised against pro-fibrotic proteins. The histology of the large intestine of the mouse is similar to that of humans and consists of a lining of columnar epithelium containing many goblet cells. The epithelium surrounding the lumen is underlain by a somewhat poorly characterized myofibroblast layer embedded in the lamina propria and surrounded by a submucosa and two layers of muscularis (an inner circular and an outer longitudinal layer) covered with a thin layer of mesothelium.

The first appearance of adhesions was seen as early as day 1 with the appearance of loose granulation tissue between adjacent intestinal loops and/or the cecum (Fig. 3). The development of the adhesion proceeds during day 2 with increasing cellularity and vascularity within the developing adhesion (Fig. 4). In addition, there is the appearance of FnEDA on day 2, which first is deposited in the small space between the closely apposed bowel loops (Fig. 5A). Figure 5B is an immunofluorescent image from the same region as shown in Figure 5A but at a greater magnification; it clearly shows that FnEDA is initially deposited as the adhesion is in its formative stage in the region between the adjacent intestinal loops. Between day 2 and day 5 after surgery, the adhesion develops rapidly with greatercellularity as well as vascularization of the newly generated adhesion as shown in Figure 6. The adhesion is a well-formed entity with many blood vessels (Fig. 6). Highly cellular and well-formed adhesion with both arterial and venous structures. Note the central blue staining region (arrow) within the adhesion indicating the presence of collagen. (Color version of figure is available online.)

The presence of collagen was confirmed by trichrome staining of the adhesion on day 5 post-surgery. (Fig. 6). In addition, there is the appearance of FNEDA within the developing adhesion. There is also localization of the antibody to the smooth muscle cells associated with the tunica media of blood vessels within the same area which serves as a “positive” control for the antibody (arrows). ×100. (B) Control photograph of region between two intestinal loops showing adhesion area (bracket) incubated with two secondary antibodies alone (no primaries) and DAPI nuclear stain showing no specific localization. Bracket indicates region of adhesion. ×100. αSMA = alpha smooth muscle actin. Arrows point to artery on left, small arterioles in center and large vein on right. (Color version of figure is available online.)

To determine if differences in gene expression were associated with the histologic and immunohistochemical changes observed, laser capture dissection of the adhesive tissue was carried using a Nikon A1R confocal microscope. Images of tissues stained with antibody to type I collagen were collected from three different mouse samples for each time point. Multiple adhesions from each of three 5 day and three 8-day post-surgery adhesions were analyzed using Image J image analysis of confocal photographs. For photographs used for the quantification of collagen, the camera settings were unchanged for all photographs taken with the digital camera. As shown in Table 2, there is a highly statistically significant increase (P = 1.6 x 10^-10) in the level of type I collagen found in adhesions at 8 d post-surgery versus those at 5 d.

To quantify the progressive connective tissue changes in adhesions temporally, measurements of pixel density were carried using a Nikon A1R confocal microscope. Images of tissues stained with antibody to type I collagen were collected from three different mouse samples for each time point. Multiple adhesions from each of three 5 day and three 8-day post-surgery adhesions were analyzed using Image J image analysis of confocal photographs. For photographs used for the quantification of collagen, the camera settings were unchanged for all photographs taken with the digital camera. As shown in Table 2, there is a highly statistically significant increase (P = 1.6 x 10^-10) in the level of type I collagen found in adhesions at 8 d post-surgery versus those at 5 d.

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carried out. Adhesions were identified and boundaries of the adhesions defined by the staining of the muscularis externa with phalloidin (Fig. 9). Tissue between adjacent bowel loops was captured and subjected to RNA isolation and quantitative polymerase chain reaction (qPCR) quantification as described in the Methods. These data, which are shown in Figure 9 and which show an approximately four fold increase in type I collagen gene expression, corroborate measurements of type I collagen pixel density (Table 2). Thus, immunohistochemistry and qPCR both show increased expression of type I collagen between day 5 and day 8. In addition, qPCR measurements also showed increased expression for type III collagen (~seven fold increase), plasma Fn (~5- fold increase), αSMA (~five fold increase), and FnEDA (~10-fold increase). Interestingly, of the pro-fibrotic genes measured, the largest overall increase was that of FnEDA (Fig. 10).

Trametinib drug treatment

To evaluate the effect(s) of the drug trametinib on adhesion formation, a series of experiments were carried out in which mice were treated with three different concentrations of trametinib over an 8-day period. Animals appeared to tolerate the drug with no adverse events noted (changes in behavior, eating, weight loss). Changes in average weight within each dosage group over the 8 d period of drug delivery were unremarkable: 1.0 mg dose - 2.2 % average decrease in weight, 3.0 mg dose: 0.5% average increase in weight. As well, there appears to be no effect on wound closure and healing (Fig. 11). The surgical site shown is representative of the animals receiving the highest drug dose (3 mg/kg/d). In most of the animals, the wound was essentially healed by 8 d post-surgery. Since the drug is minimally soluble in water, DMSO was used as a “vehicle” to solubilize it. To investigate the possibility that the DMSO itself may induce side effects, an additional series of five animals were subjected to the same surgeries as the mice receiving drug, that is, laparotomies and sub-dermal osmotic pump placement; however, for these additional animals, the pumps contained DMSO but no drug. Analysis of these animals demonstrated that the DMSO neither had any effect on survivability nor wound healing. Thus, although these animals developed abdominal adhesions, they showed no discernible consequences of the treatment with DMSO (data not shown).

In Figure 12A, the adhesion between adjacent bowel loops of a day 8 post-surgery mouse treated with trametinib (0.1 mg/kg/d) show cells largely localizing antibody to FnEDA in the center of the adhesion. There does not appear to be any cells which localize antibody to αSMA. In Figure 12B, the adhesion, also treated with 0.1 mg/kg/d of trametinib, appears to be a more advanced adhesion than that shown in Figure 12A since there are more myofibroblast cells localizing antibody to αSMA. Nevertheless, few cells (arrows) localize antibody to αSMA suggesting the presence of a modest
However, our in vitro myofibroblasts because it is beyond the scope of this study; developing adhesion. We did not address the origin of the adhesions. We have used a mouse model of cecal abrasion to initiate the formation of abdominal adhesions. Adhesion formation begins almost immediately after surgery and adhesion sites show evidence of the presence of myofibroblasts within 5 d, as demonstrated by cells expressing α-SMA and FnEDA within the developing adhesion. We did not address the origin of the myofibroblasts because it is beyond the scope of this study; however, our in vitro experiments with RPMS suggest the possible contribution of mesothelial cells to the pathophysiology of adhesion formation.

Myofibroblasts, associated with fibrotic reactions, may arise from different sources depending on the affected organ and the initiating event including: (1) activation of tissue resident fibroblasts in response to specific signals from infiltrating inflammatory cells; (2) recruitment of bone marrow precursor cells known as fibrocytes which express bone marrow cellular surface markers such as CD34 but are capable of ECM production; and (3) trans-differentiation of epithelial, mesothelial, and endothelial cells into activated myofibroblasts. Although this process was originally described in epithelial cells and designated epithelial to mesenchymal transition, it is now known that very similar processes occur in the case of mesothelial (MMT) and endothelial (EndoMT) cells.

In these transitions, the epithelial, mesothelial, or endothelial cells lose their cell type—specific markers and phenotypic traits, such as expression of E-cadherin and acquire a mesenchymal or myofibroblast phenotype, ultimately initiating the expression of α-SMA and vimentin and ECM proteins including FnEDA. The expression of these proteins is consistent with a transition of the mesothelial cells to myofibroblasts. Similarly, our immunofluorescence data in the mouse model at 2 d post-surgery show the presence of FnEDA within the adhesions suggesting that these FnEDA positive cells either have or are transitioning into a contractile myofibroblast.

Cells which are FnEDA “positive” and α-SMA “negative” are considered proto-myofibroblasts, and it is only later after the proto-myofibroblasts express α-SMA that they are termed myofibroblasts. Thus, the presence of contractile myofibroblasts is a hallmark of fibrotic tissue whose maturation is preceded by the expression of FnEDA. The fact that some cells within the adhesions, shown in Figure 11A, express only FnEDA and not α-SMA (compare Fig. 12A and B) suggests that trametinib may interfere with the conversion of the precursor proto-myofibroblast into a myofibroblast. A functional consequence of this may be the inability of the proto-myofibroblast to contract since...
both $\alpha$SMA and FnEDA are required to mediate the transfer of intracellular force to the ECM. It has been demonstrated that myofibroblast contraction activates latent TGF-$\beta_1$ releasing it from the ECM, which initiates the pro-fibrotic cascade of events. It is important to note that the occurrence of adhesions in the drug-treated animals given the highest dose of trametinib (3 mg/kg/d) must be an extremely rare event if it occurs at all. In the five animals whose bowels were serially sectioned, we observed regions where the cecum and intestinal loops were in close proximity to one another but without development of a discernible adhesion (Fig. 14). When such regions were found in the trametinib-treated animals (0.1 and 1.0 mg/kg/d dose levels), lesion development appeared to be inversely proportional to the level of drug administered. For example, fairly extensive lesions were found at the lowest dose while mice administered the 1 mg/kg/d had adhesions which were poorly formed and which appeared to have low levels of $\alpha$SMA “positive” cells. These data support the view that adhesions in mice treated with the middle dose of trametinib (1 mg/kg/d) had limited complete trans-differentiation of cells into myofibroblasts, which we and others define as a cell expressing both $\alpha$SMA and FNEDA. This diminution of myofibroblast presence is in contrast to that found either in untreated animals or in animals receiving the lowest dose (Fig. 12B). This suggests that trametinib may be active in ameliorating and/or blocking the early stages of adhesion formation, something which may be useful in preventing formation in humans.

The localization of FNEDA (Figs. 5 and 12A) occurred in the central region either between the cecum and bowel loops or associated with the surface of the abraded cecum. These central regions quickly became highly cellular and vascularized. Our in vitro studies with rat mesothelial cells show that after stimulation with TGF-$\beta$, they undergo MMT and begin expressing both $\alpha$SMA and FnEDA and that U0126, a Mek1/2 inhibitor, was very effective in blocking their MMT. Practically, mesothelial cells are reasonable candidates to trans-differentiate into myofibroblasts as others have suggested. However, much additional work will need to be carried out to validate this possibility because it is also

Fig. 12 — Immunofluorescence confocal photographs of two adhesions at day 8 post-surgery from mice treated with 0.1 mg/kg/d of trametinib (FnEDA = green, $\alpha$SMA = red, DAPI nuclear stain = blue, M = muscularis). (A) Antibody to FnEDA is prominently localized to the adhesion center while only a few cells localize antibody to $\alpha$SMA ×200. (B) Extensive localization of antibody to $\alpha$SMA to the smooth muscle cells of the muscularis externa is shown. In addition, note the localization of $\alpha$SMA to many myofibroblasts and blood vessels within the adhesions. ×200. $\alpha$SMA = alpha smooth muscle actin; FnEDA = fibronectin EDA. (Color version of figure is available online.)

Fig. 13 — Immunofluorescence confocal photograph of a section through adjacent intestinal loops showing a potential adhesion (bracket) in mice treated with 0.1 mg/kg/d of trametinib on day 8 post-surgery (FnEDA = green, $\alpha$SMA = red, DAPI nuclear stain = blue). Note that the region between the cecum and the bowel segment (a presumptive adhesion, bracket) is devoid of cells localizing antibody neither to FnEDA nor $\alpha$SMA and that only the cecum is expressing FnEDA as a result of the abrasion technique. The opposite bowel loop has intact mesothelial nuclei and likely represents uninvolved bowel. ×200. $\alpha$SMA = alpha smooth muscle actin; FnEDA = fibronectin EDA. (Color version of figure is available online.)
possible that the myofibroblasts may originate from other sources as well. Interestingly, qPCR data of RNA isolated from adhesions by laser capture microscopy at day 5 and 8 postsurgery show the expression of FnEDA, αSMA, and collagen increased significantly raising the possibility that MMT may occur in vivo.

In our current investigation, these previous in vitro studies using UO1126 were duplicated using trametinib on RPMCs with similar results to those found at both the morphologic and molecular levels. In the presence of trametinib, the characteristic cobblestone appearance of normal mesothelial cells was maintained (Fig. 2), the expression of FnEDA and αSMA was inhibited, and the phosphorylation of Erk1/2 was essentially blocked completely (Fig. 2B). Interestingly, the concentrations of trametinib used in the present experiments were considerably lower (2 or 5 nM) than that used in experiments involving cultured melanoma cells (100 nM). This suggests that a positive therapeutic response with trametinib could be attained at a much lower dose in fibrotic reactions than that required for tumor responses in vivo, minimizing any potential toxic events of the drug.

Significantly, during adhesion formation in the mouse model, there was a rapid formation of blood vessels within the adhesion. Since surgery may disrupt blood flow, a consequence is that the tissue becomes hypoxic which is accelerated by the rapid influx of highly metabolic inflammatory cells participating in the wound healing process. These cells initiate the formation of granulation tissue and promote the trans-differentiations and proliferation of the myofibroblasts. For example, fibroblasts can be induced to secrete increased amounts of TGF-β under hypoxic conditions thus contributing to the overall pro-fibrotic process during wound healing. The role of hypoxia has been further corroborated by studies in rats which demonstrated that hyperbaric oxygen can reduce the pro-fibrotic changes associated with intra-abdominal adhesions presumably by increasing PO2 tissue levels.

The present findings demonstrate that trametinib can effectively inhibit adhesion formation in a mouse model that
mimics potential human clinical situations. Since previous treatments have met with limited success, new therapeutic approaches are necessary and additional clinical testing of trametinib will be required to validate its use in humans. Most importantly with respect to its therapeutic use in humans, our observations also demonstrate that the effective dosage(s) of trametinib used had no adverse effect on the healing of the surgical wound required to access the abdominal cavity in the mouse model. At necropsy 8 d post-surgery both the abdominal musculature and the overlying skin had largely healed. This demonstrates that trametinib, at least in the mouse model, did not impede wound healing.

In summary, these data suggest that abdominal adhesion formation appears to be the result of common pathophysiologic mechanisms operative in other fibrotic reactions mediated, at least in part, by the phosphorylation of Erk1/2. Activation of these signaling pathways results in formation of an ECM containing collagen and FN in resident cells.

Conclusions

1. Cecal abrasion is a reliable model for generation of intra-abdominal adhesions.
2. Trametinib, a MEK1/2 inhibitor, ameliorated adhesion formation with little or no effect on wound healing at the laparotomy site in mice.
3. Adhesions quickly become populated by myofibroblasts as the adhesions matured which did not occur in mice treated with 3 mg/kg/d of trametinib.
4. This model may be of great use to test the effects of other therapeutic agents on abdominal adhesion formation.

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Disclosure

E.J.M. and J.R. and Thomas Jefferson University have applied for a “use” patent for the drug trametinib in blocking adhesion formation. All other authors report no proprietary or commercial interest in any product mentioned or concept disclosed in this article.

References


