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Review ROS and miRNA Dysregulation in Ovarian Cancer Development, Angiogenesis and Therapeutic Resistance

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Abstract: The diverse repertoires of cellular mechanisms that progress certain cancer types are being uncovered by recent research and leading to more effective treatment options. Ovarian cancer (OC) is among the most difficult cancers to treat. OC has limited treatment options, especially for patients diagnosed with late-stage OC. The dysregulation of miRNAs in OC plays a significant role in tumorigenesis through the alteration of a multitude of molecular processes. The development of OC can also be due to the utilization of endogenously derived reactive oxygen species (ROS) by activating signaling pathways such as PI3K/AKT and MAPK. Both miRNAs and ROS are involved in regulating OC angiogenesis through mediating multiple angiogenic factors such as hypoxia-induced factor (HIF-1) and vascular endothelial growth factor (VEGF). The NAPDH oxidase subunit NOX4 plays an important role in inducing endogenous ROS production in OC. This review will discuss several important miRNAs, NOX4, and ROS, which contribute to therapeutic resistance in OC, highlighting the effective therapeutic potential of OC through these mechanisms.

Keywords: ovarian cancer; miRNA dysregulation; ROS; NOX4; HIF1- α ; VEGF; angiogenesis; HER3; therapeutic resistance

1. Ovarian Cancer

Known as the silent killer, ovarian cancer (OC) has the lowest survival rate and the worst prognosis among all gynecologic malignancies in the US; and is the eighth most common cancer in women worldwide [1,2]. In 2022, the American Cancer Society estimates about 21,000 new cases of OC will be diagnosed, and approximately 14,000 women will die from this type of cancer. The overall 5-year survival rate is only 48% due to OC's ambiguous symptoms and inadequate screening capabilities at the early stages of the disease. Due to late detection, about 60% of new cases are diagnosed when the disease has already progressed to the advanced stage [2]. OC is a heterogeneous disease with several subtypes that differ in their gene expression, tumor origin, pathway alterations, and pathogenesis. The majority of OC originates from three main cell types: epithelial cells (90%), stromal cells (7%), and germ cells (3%) [1,3,4]. In general, epithelial OC can be further divided into five histotypes: high-grade serous (HGSOC; 70%), endometrioid (ENOC; 10%), clear cell (CCOC; 10%), mucinous (MOC; 5%), and low-grade serous (LGSOC; less than 5%) OC [4]. In addition, another classification system was introduced a decade ago that divided OC into type I and II tumors. Type I tumors are low-grade neoplasms, including mucinous carcinomas, endometrioid carcinomas, malignant Brenner tumors, and clear cell carcinomas. Type I tumors are typically characterized by mutations in BRAF, KRAS, and PTEN with DNA instability. Type II tumors are high-grade serous carcinoma, carcinosarcoma, and undifferentiated carcinoma, which are frequently observed with mutations in p53, BRCA1/2, HER-2/HER-3 overexpression, and p16 inactivation [5–8]. Depending on the specific subtype and histopathology, OC treatment involves a combination of surgery and



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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). chemotherapy. For patients with advanced-stage tumors, debulking surgery is recommended; however, large tumors or residual tumors may show negative side effects leading to blockage of the perfusion area and the possibility of developing drug resistance [1,9]. Platinum-based chemotherapy is the standard line of treatment for OC, either in conjunction with or following surgery [10–12]. The combination of paclitaxel/carboplatin has been recognized as the standard postoperative chemotherapy for many years [13]. In recent years, PARP inhibitors have been incorporated into clinical treatment as a recommended maintenance drug [14]. However, due to the aggressive growth rates and the propensity of advanced tumors to evade treatment, there are critical limitations to the current lines of therapy. A better understanding of the molecular biology of OC is allowing more research efforts to establish new effective treatment options for advanced-stage tumors.

2. ROS

Reactive oxygen species (ROS) have remained a highly relevant topic over the last few decades due to their expansive effects on normal cellular function. Oxidative stress is generated through the accumulation of ROS, either through exogenous exposure or endogenous production. ROS are oxygen ions with unpaired electrons (singlet oxygen ${}^{1}O_{2}$, superoxide O_{2} ·⁻) or oxygen-containing molecules, such as hydroxyl radicals (OH·⁻), hydrogen peroxide (H₂O₂), nitric oxide (NO), and nitrogen dioxide (NO₂) [15]. Superoxide radicals are converted into H₂O₂ by the enzyme superoxide dismutase (SOD). However, superoxide can also react with nitric oxide to produce peroxynitrite (ONOO⁻), a strong oxidizer with damaging cellular effects [16]. The accumulation of H_2O_2 has detrimental effects on nuclear and mitochondrial DNA, which may lead to genetic instability to drive cancer progression with increased expression of oncogenes and decreased expression of tumor suppressors [17,18]. Several enzymes work in conjunction to convert H₂O₂ into the water, including catalase, glutathione peroxidases 1 and 4, and peroxiredoxins 3 and 5 [19–22]. Furthermore, H_2O_2 can also participate in the Fenton reaction, in which free iron Fe(II) reacts with H_2O_2 , generating highly reactive hydroxyl radicals (·OH)(shown below). The production of hydroxyl radicals (·OH) by the Haber–Weiss reaction (shown below) further perpetuates the damaging effects of the accumulation of ROS.

Fenton Reaction:

$$Fe(II) + H_2O_2 \leftrightarrow Fe(III) + \cdot OH + OH^-$$

Haber-Weiss Reaction:

$$O_2 \cdot \overline{} + H_2 O_2 \longleftrightarrow OH + OH^- + O_2$$

Original studies implicated the mitochondria as primary endogenous sources of superoxide through the process of cellular respiration, a process dependent on the availability of O_2 [23–25]. Based on this view, the production of ROS was thought to be a harmful by-product of intracellular metabolism. Then a family of transmembrane enzymes known as NADPH oxidase (NOX) proteins was identified, whose primary function was the production of endogenous ROS. NOX2, the first NOX protein discovered, was the primary producer of endogenous ROS in leukocytes to generate an oxidative burst, an essential process for the neutralization of pathogens [26–29]. The characterization of a disease called chronic granulomatous disease (CGD) caused by a mutation in the phagocytic NOX gene provided insight into the emerging role of endogenous ROS production on cellular functionality [30,31]. Subsequent work demonstrated a pivotal role of NOX proteins in mammalian cell transformation through the production of ROS in OC cells was attributed to H₂O₂ increased levels induced by NOX4 [34], identifying an endogenous mechanism for the overproduction of ROS and alteration of intracellular signaling in OC tumor development.

Under normal cellular conditions, low levels of endogenous ROS activate several signaling pathways involved in cell proliferation. However, the accumulation of ROS causes

extensive damage to DNA, RNA, proteins, and lipids, thus causing a significant hindrance to normal cellular functions and contributing to the development of multiple human pathologies [35–38]. The damage can induce cell death pathways or trigger the mutation of DNA, as commonly found in cancer [39,40]. In addition to the endogenous production of ROS and oxidative stress, external or environmental exposure to ROS can have detrimental effects on mammals [41]. For instance, many chemotherapeutic agents induce oxidative stress as a means of inducing cellular damage and cell death pathways [42]. However, as demonstrated by more recent findings, ROS play an important role in the progression and advancement of human diseases. The counterweight for endogenous ROS is the genetically programmed redox system. This includes groups of genes coding for antioxidant proteins such as superoxide dismutase (SOD), catalase, and the glutathione system, which neutralize the ROS produced in cells [43-45]. The failure to neutralize endogenous ROS leads to a build-up of harmful oxygen species and, consequently, oxidative stress. In normal cells, oxidative stress leads to deleterious cellular effects, such as protein, lipid, and DNA damage, organelle dysfunction, and cell cycle arrest [46]. Higher levels of oxidative stress cause the activation of cell death pathways such as apoptosis and necrosis [46], which may be mitigated in cancer cells by an increase in antioxidant production. The upregulation of nuclear factor erythroid 2-related factor 2 (NRF2), a master transcriptional regulator of antioxidant genes, contributes to the neutralization of endogenous ROS in OC cells [47–49], making NRF2 a viable target for chemotherapeutic treatment in certain cases of OC. In addition, the genetic mutation of cellular pathways that induce cell death mechanisms in response to increased oxidative stress allows cancer cells to evade the activation of cell death pathways [50], thus providing cancer cells the ability to continue continuous proliferation in the presence of adverse cellular conditions, such as oxidative stress.

3. ROS in the Development of Ovarian Cancer

There is an established link between an increase in ROS production and cancer development in humans [51]. As secondary cellular signaling molecules, ROS are involved in the activation of several signaling pathways involved in cell proliferation and growth. Consequently, these pathways are constitutively activated in cancer cells with increased ROS levels that contribute to tumorigenesis [51]. For example, endogenously derived ROS activate the ERK1/2 MAPK signaling pathway and the AKT signaling pathway in OC, both of which promote cell proliferation [52,53]. The increased ROS generation also contributes to a genetic mutation in cancer cells, further contributing to cell transformation [54,55]. As opposed to the traditional view of ROS generation in cancer as a harmful secondary by-product, the increasing knowledge of cancer cell metabolism and signal transduction is exposing ROS as a positive contributing factor in tumorigenesis and cancer development. The increased metabolic activity of cancer cells was originally thought to be responsible for the accumulation of ROS as a byproduct of increased glycolytic metabolism and mitochondrial respiration [56]. However, the discovery of the role of NOX proteins in endogenous ROS production revealed a more important role for ROS production in non-phagocytic cells, particularly in cancer [57–59]. The endogenous production of ROS by NOX1 was found to be responsible for increased viability and proliferation in colon cancer [60,61]. Similarly, the role of NOX2-mediated ROS production was discovered to be critical for cell viability and proliferation in breast, colorectal, myelomonocytic leukemia, gastric, and prostate cancers [62–67]. NOX4 overexpression contributed to an oncogenic proliferation in renal cell carcinoma, melanoma, glioblastoma, ovarian, prostate, and lung cancers [34,68–72]. In OC cell lines, there is a significant increase in ROS production, which contributes to tumorigenesis [34]. The increase in ROS is a result of NADPH oxidase activity and mitochondrial metabolism, as this increase is diminished by NADPH oxidase and mitochondrial complex I inhibitors [34]. Moreover, the increased levels of ROS result from the upregulation of the NADPH oxidase subunit NOX4, which serves as the main contributor to ROS production in OC cells to promote tumor growth and angiogenesis [34]. Furthermore, the activation of NOX4 is positively correlated with TGF-β1 and NF-κB activity, which is suppressed by their inhibitors [34]. This system demonstrates that endogenous NOX4-derived ROS are a driving force in OC development. Moreover, NOX4 is a potential target for the therapeutic resistance of OC which is dependent on ROS production for an increase in oncogenic signaling.

4. miRNA Dysregulation in Cancer

The progression of cancer is often associated with dysregulation of non-coding RNAs, including microRNAs (miRNAs) [73,74]. miRNAs are 18–25 nucleotide long, non-coding single-stranded RNA molecules that regulate the expression of messenger RNA (mRNA) [75]. The discovery of miRNA in 1993 by Ambros and colleagues in the nematode C. Elegans revealed the critical role of miRNAs in the post-transcriptional regulation of mRNA [76,77]. In these studies, the miRNA lin-4 was found to regulate the expression of the critical developmental transcription factor, lin-14 [76,77]. The primary transcripts of miRNA (pri-miRNA) are modified within the nucleus by the RNase III DROSHA and its cofactor DGCR8 before being exported to the cytoplasm as pre-miRNA [78,79]. Mature miRNA molecules are the result of the cleavage of pre-miRNA at the terminal loop by the RNase III endonuclease, DICER [80,81]. The regulation of miRNA processing can have expansive effects on cellular processes, as demonstrated by the gain-of-function mutation of DICER as a contributory factor in cancer development [82]. As transcriptional regulatory molecules, miRNAs typically recognize and bind the 3'-UTR of target mRNAs to repress expression or induce degradation [83]. The activation of genes by miRNAs occurs through association with the promoter region and upstream regulatory regions of target genes [84]. The search for the role of miRNA in humans yielded a plethora of data that are still accumulating, particularly the dysregulation of miRNAs in oncogenesis. The original studies identifying the role of miRNAs in human oncogenesis demonstrated the effect of miR-15a/16arepression on promoting the oncogenic protein Bcl-2 in chronic lymphocytic leukemia [85]. Most human miRNAs function as tumor suppressors by directly targeting and inhibiting oncogenes, such as RAS and MYC. For instance, the downregulation of Let-7 family of miRNAs, which target KRAS and C-MYC, is found in OC, which induces tumor growth and development [86,87]. However, some miRNAs function as oncogenes by directly targeting and inhibiting tumor suppressors such as p53 [88,89]. For example, miR-25 and miR-30d target p53 for degradation and contribute to colon cancer development; the downregulation of both miR-25 and miR-30d led to an increase in p53 protein expression and increased apoptosis in multiple cancer types [90]. Many miRNAs are dysregulated in multiple cancers, including the upregulation of miR-155 in lymphomas and colorectal cancers [91,92], indicating a commonality in the mode of miRNA dysregulation in multiple cancer/tissue types. The molecular effects of miRNA dysregulation include feedback mechanisms, such as the miR-17-92 cluster/E2F family/c-MYC loop. In this feedback mechanism, miR-17-92 is activated by c-MYC and inhibits E2F family protein translation [93,94]. The E2F family of proteins (E2F1, E2F2, E2F3) are critical cell-cycle regulated inducers of proliferation, therefore proper regulation of these proteins is necessary under normal conditions [95]. Further investigation revealed that c-MYC activation of E2F family proteins activates miR-17-92, leading to a feedback loop to tightly control the expression of E2F proteins in healthy cells [96,97]. However, in cancer cells the amplification and overexpression of miR-17-92 disrupts this feedback loop and contributes to high cell proliferation and tumorigenesis [98]. In another example, miR-221/222 upregulation in cancer cells contributes to oncogenesis through the inhibition of cell cycle regulating protein p27 [99–101]. The dysregulation of particular miRNAs can differ between subtypes of OC. For instance, the overexpression of miR-483 occurs in serous epithelial ovarian cancer (EOC), but does not occur in non-serous EOC [102]. As demonstrated in voluminous publications, miRNA dysregulation affects a variety of cellular processes that contribute to oncogenesis in a wide variety of cancers. The complex role of miRNAs in cancer development highlights the potential for therapies targeting specific miRNAs that are dysregulated in different cancers.

5. ROS and miRNA Dysregulation in Angiogenesis and Ovarian Cancer Development

The development of tumors involves a wide variety of cellular processes. In this regard, ROS contribute to critical cellular processes that occur within tumors, including angiogenesis and micro-RNA (miRNA) dysregulation. Angiogenesis is the creation of new blood vessels within existing vasculature, which is essential for processes such as embryogenesis, tissue repair, and organ regeneration [103]. Unsurprisingly, angiogenesis plays a pivotal role in cancer development through the establishment of nutrients and blood supply to newly formed tumors [104]. A significant contribution to angiogenic signaling is made by vascular endothelial growth factor (VEGF), which is highly upregulated in developing embryonic cells and tumor cells [105–107]. The limited oxygen availability in tumors often leads to hypoxic conditions, in which signaling pathways are activated to initiate tumor growth and angiogenesis [108]. The hypoxia-inducible factor 1 alpha (HIF-1 α) plays a vital role in the hypoxic response in tumor cells, partially through the upregulation of VEGF [109–111]. The dysregulation of HIF1- α occurs in a wide variety of cancers which contributes to tumorigenesis [112]. The upregulation of HIF-1 α and VEGF are positively correlated with NOX4-derived ROS production in OC cells and promotes angiogenesis and tumor growth [34]. In turn, HIF-1 α induces the expression of VEGF; and promotes the production of NOX4 through an alternative splicing mechanism [34,113]. This positive feedback system demonstrates the capacity of OC cells to utilize the overproduction of NOX4-derived ROS to activate HIF-1 α and VEGF and promote angiogenesis and tumor growth.

This regulation system is complicated more by the dysregulation of miRNAs by increased intracellular ROS, which contributes to OC tumorigenesis. For instance, miR-199a and miR-125b downregulate the expression of the oncogenic proteins HER2 and HER3 under normal cellular conditions [114]. However, increased ROS in OC cells results in the downregulation of miR-199a and miR-125b through DNA hypermethylation, thereby increasing the expression of HER2/3 and contributing to tumorigenesis [114]. More importantly, this demonstrates the role of epigenetic regulation of miRNA expression. Another example of epigenetic regulation of miRNA expression is the increased acetylation of miR-466-5p in response to increased ROS, thereby inducing the expression of miR-466-5p, which then activates pro-apoptotic genes [115,116]. The expression of some miRNAs is regulated by ROS-stress-responsive transcription factors, which contribute to molecular signaling cascades. The tumor suppressor p53 is induced in response to ROS and subsequently activates the miR-200 family of miRNAs [117]. The miR-200 family members have been implicated as tumor suppressors, and overexpression of these miRNAs inhibits tumor development in OC [118,119]. However, two members of this family of miRNAs, miR-141 and miR-200a, directly target p38 α in response to increased levels of ROS, resulting in evasion of apoptosis induction and upregulation in antioxidant production [120,121], thus demonstrating the complexity of molecular roles of a single family of miRNAs in OC progression. As contributing factors to the pathway described above, miR-21 and miR-27a induce angiogenesis in OC through the upregulation of HIF1- α and VEGF, respectively [122,123]. Additional pathways affected by miRNA dysregulation also contribute to angiogenesis in OC. For instance, miR-141 is upregulated in OC to induce the expression of VEGFR2, resulting in an increase in angiogenesis [124]. By a differing mechanism, the upregulation of miR-205 in OC results in an increase in angiogenesis through the downregulation of tumor suppressor PTEN and an increase in AKT signaling [125]. Similarly, miR-204 upregulation in OC contributes to angiogenesis through the downregulation of anti-angiogenic protein THBS1 [126,127]. The miRNAs listed in Table 1 are upregulated in OC cells and contribute positively to tumor growth, development, angiogenesis, and therapeutic resistance. However, there are substantial data demonstrating a tumor suppressor role for various miRNAs in OC, whose downregulation results in tumorigenesis, angiogenesis, and treatment resistance (Table 2) [128]. For instance, miR-145 acts as a tumor suppressor, and the downregulation of miR-145 in OC contributes to angiogenesis through the upregulation of HIF-1 α and VEGF [129]. The overall role of the miRNAs in OC development described here is shown

in Figure 1. Another important avenue of miRNA research involves the dysregulation of specific circulatory miRNAs, which has the inherent propensity to impact a multitude of tissue types. Recent work has shown that the expression levels of miR-200b, miR-200c, miR-141, and miR-1274A in OC patients' circulatory systems are negatively correlated with survival [130]. Therefore, the roles of miRNA dysregulation in OC angiogenesis and development remain to be fully understood which warrants further investigation to provide therapeutic options and/or targets in the future.

Table 1. miRNAs upregulated in OC cells that contribute to tumor growth and development, angiogenesis, and therapeutic resistance.

| miRNA | Target mRNA(s)/Function | Ref |
|----------|---|-----------|
| miR-21 | APAF1/Promotes angiogenesis and treatment resistance (paclitaxel) | [131,132] |
| miR-22 | MXI1/Promotes tumor growth | [133] |
| miR-27a | VEGF/Promotes VEGF expression to promote angiogenesis | [123] |
| miR-30a | FOXD1/Promotes cell cycle progression and growth | [134] |
| miR-92a | DKK1/Promotes Wnt signaling and tumor growth | [135] |
| miR-99a | FN1, VTN/Promotes tumor metastasis | [136] |
| miR-106a | CASP7/Promotes treatment resistance (paclitaxel) | [137] |
| miR-141 | $p38\alpha$, KEAP1/Promotes tumor growth and treatment resistance (cisplatin) | [121,138] |
| miR-181a | SMAD7/Promotes tumor growth, angiogenesis, and treatment resistance | [139] |
| miR-182 | MTSS1, PDCD4/Promotes tumor growth, metastasis, and treatment resistance (cisplatin and paclitaxel) | [140–142] |
| miR-200a | $p38\alpha$ /Promotes tumor growth | [121] |
| miR-203 | PDHB/Promotes cell proliferation | [143] |
| miR-204 | THBS1/Promotes angiogenesis and tumor growth | [126,127] |
| miR-205 | SMAD4, PTEN/Promotes metastasis and angiogenesis | [144] |
| miR-210 | PTPN1/Promotes survival and evasion of cell death mechanisms | [145] |
| miR-214 | PTEN/Promote cell survival and treatment resistance (cisplatin) | [146] |
| miR-223 | <i>PTEN</i> /Promote tumor growth and treatment resistance (cisplatin) | [147] |
| miR-376a | KLF15/Promotes cell cycle progression and growth | [148] |
| miR-443 | MAD2/Promote tumor growth and treatment resistance (paclitaxel) | [149] |
| miR-551b | STAT3/Promotes tumor growth | [150] |
| miR-552 | PTEN/Promotes tumor metastasis | [151] |
| miR-622 | <i>KU70, KU80</i> /Promotes treatment resistance (cisplatin and PARP Inhibitors) | [152] |
| miR-939 | APC2/Promotes Wnt signaling and tumor growth | [153] |
| miR-1246 | CAV1/Promotes treatment resistance (paclitaxel) | [154] |

| miRNA | Target mRNA(s)/Function | Ref |
|---------------|--|---------|
| Let-7 Family | KRAS, c-MYC/Tumor suppression through KRAS and c-MYC downregulation. | [86,87] |
| miR-31 | CDKN2A/Tumor suppression through CDKN2A downregulation. | [155] |
| miR-125b | <i>VEGF, HER3, HIF1-</i> α /Tumor suppression through <i>VEGF, HER3</i> and <i>HIF1-</i> α downregulation. | [114] |
| miR-135a | CCR2/Tumor suppression through CCR2 degradation. | [156] |
| miR-145 | P70S6K1/Tumor suppression through P70S6K1 downregulation. | [129] |
| miR-181 | RTKN2/Tumor suppression through RTKN2 downregulation. | [157] |
| miR-199a | HER3/Tumor suppression through HER3 downregulation. | [114] |
| miR-200b/200c | DNMT3A/3B/Tumor suppression through DNMT3A/3B downregulation. | [158] |
| miR-206 | <i>c-MET</i> /Tumor suppression through <i>c-MET</i> downregulation. | [159] |
| miR-298 | EZH2/Tumor suppression through EZH2 downregulation. | [160] |
| miR-424 | CCNE1/Tumor suppression through CCNE1 downregulation. | [161] |
| miR-490 | CDK1/Tumor suppression through CDK1 downregulation. | [162] |
| miR-508 | MAPK1/Tumor suppression through MAPK1 downregulation. | [163] |

Table 2. miRNAs downregulated in OC cells that function as tumor suppressors.

miRNA Dysregulation in Ovarian Cancer Development

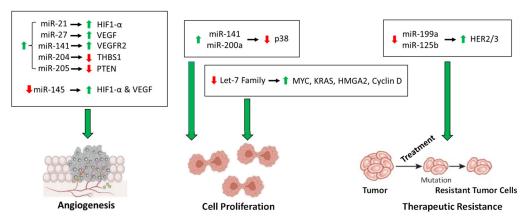


Figure 1. The roles of miRNA dysfunction in OC development. As described in the text, the miRNAs shown above are dysregulated in OC that contribute to angiogenesis, cell proliferation, and therapeutic resistance. The dysregulation of each miRNA is denoted by an up arrow (upregulation) or a down arrow(downregulation). The regulation of proteins affected by the dysregulation miRNAs is denoted in the same manner.

6. Potential Mechanism of ROS in Therapeutic Resistance in Ovarian Cancer

The two major obstacles facing efficient treatment of OC are late detection/diagnosis and acquired therapeutic resistance. The standard treatment for OC includes preliminary debulking surgery followed by platinum-based (carboplatin and cisplatin) and/or taxane family-based (paclitaxel and docetaxel) chemotherapy [164,165]. The mode of action of platinum-based therapies is oxidative stress-induced cellular damage and initiation of cell death pathways, such as apoptosis, which is triggered by this class of chemotherapeutics [166]. The taxane family-based drugs are used to inhibit cell division through microtubule stabilization [167]. However, due to toxic side effects associated with high-dose treatment and acquired resistance to carboplatin and cisplatin treatment, this traditional route of therapy has critical limitations. The mechanisms of drug resistance to these treatment options include an increase in DNA damage repair and an increase in antioxidant production to detoxify cancer cells [168]. Regarding this, other chemotherapeutic agents are used to treat resistant tumors, including gemcitabine, doxorubicin, and bevacizumab [169–171]. There are additional combinations of chemotherapy used to treat OC, such as targeted treatment of anti-apoptotic proteins that are overexpressed in OC cells. For instance, the anti-apoptotic protein Bcl-2 is overexpressed in OC, and treatment of tumor cells with a combination of cisplatin or carboplatin and Bcl-2 inhibitors show an increased level of cancer cell death induction [172–175]. The class of VEGF regulators known as specific proteins (Sp) are also targeted by small molecule inhibitors to induce cell death [176]. Another important mechanism for cancer cells to evade treatment is the upregulation of the glycoproteins that form the molecular pumps to export chemotherapeutic agents out of the cancer cells, driving the process of multi-drug resistance [177,178]. The complexity of miRNA dysregulation in OC also contributes to treatment resistance. For instance, OC cells evade apoptosis in response to paclitaxel treatment through upregulation of miR-21 and miR-106a, that target and downregulate the pro-apoptotic proteins APAF1 and CASP7, respectively [131,132]. Similarly, miR-182 upregulation in OC results in evasion of apoptosis in response to cisplatin/paclitaxel treatment through the downregulation of pro-apoptotic protein PDCD4 [142]. Regarding therapeutic efficacy, the ROS-induced miRNAs mentioned previously, miR-200a and miR-141, although shown as oncogenic, can increase the sensitivity of OC to paclitaxel treatment through the downregulation of p38 [121,179]. Similarly, overexpression of miR-522 can increase the sensitivity of OC to paclitaxel treatment [180]. A better understanding of the molecular mechanisms driving treatment resistance in OC is of vital importance for the design of therapies that will effectively treat aggressive, resistant tumors.

The increase in intracellular ROS levels in OC has been shown to contribute to therapeutic resistance. For instance, an increase in ROS in OC results in the overexpression of dCTP pyrophosphatase I (DCTPP1), which has a role in DNA damage repair and plays a major contribution to cisplatin resistance [181]. By a differing mechanism, the upregulation of calcium/calmodulin-dependent protein kinase II gamma (CAMK2G) in response to increasing levels of ROS reprograms the cellular redox system through the phosphorylation of inositol triphosphate3-kinase B (ITPKB), resulting in adaptive redox homeostasis and increased resistance to cisplatin treatment [182]. Similarly, the upregulation of PGC1- α by increasing intracellular ROS contributes to chemotherapy resistance through the upregulation of drug resistance-related proteins, MDR1 and ABCG2, leading to increased antioxidant production and drug efflux [183]. The increase in ROS in OC downregulates miR-199a and miR-125b, resulting in the increased expression of HER2 and HER3 and therapeutic resistance [114]. Thus, another mode of treatment for OC is vaccines targeting human HER2 and HER3 [184,185]. In work highlighted here, the increase in NOX4-derived ROS contributes to therapeutic resistance in OC through the upregulation of HER3. The upregulation of HER3 is a clinical marker for OC, which is positively correlated with poor prognosis [186]. NOX4 directly activates HER3 and contributes to the increased resistance of OC cells to chemotherapy and radiation treatments [113]. The deletion of NOX4 results in a reduction in the therapeutic resistance of OC cells [113]. Similarly, inhibition of NOX4 acts synergistically with HER3 inhibition to decrease tumor growth in OC [113]. The knockdown of NOX4 using siRNA also results in enhanced sensitivity to radiation treatment in OC cells, proving this pathway relevant in multi-modal therapeutic resistance [113]. The NOX4-driven system of endogenous ROS production demonstrates a new mechanism in OC cells to promote tumor development, angiogenesis, and an increase in therapeutic resistance through the upregulation of HER3, reflecting a candidate for targeted therapy of treatment-resistant OC (Figure 2). These findings shed light on the importance of endogenous NOX4-derived ROS production in cell signaling and the progression of OC and the propensity of tumors to evade current lines of treatment.

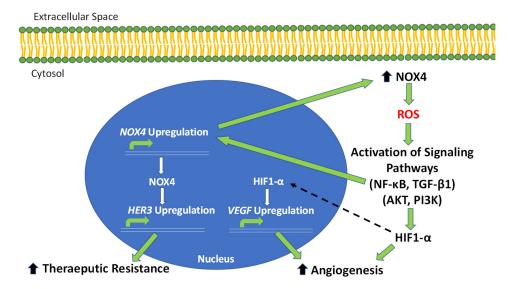


Figure 2. NOX4-driven pathways of OC tumor progression, angiogenesis and therapeutic resistance. The overexpression of NOX4 in OC results in an increase in intracellular ROS production. Increased ROS leads to an increase in HIF1-A through activating PI3K and AKT signaling. HIF1- α then activates the critical angiogenic factor, VEGF. Increased ROS also activates NF- κ B and TGF- β 1 signaling, which lead to the direct upregulation of NOX4. The increase in NOX4 contributes, in a positive-feedback manner, to increased ROS production. NOX4 also activates the expression of HER3, contributing to therapeutic resistance in OC tumors.

7. Future Directions

The key to effective treatment of OC is the understanding of the molecular mechanisms that drive tumor development and resistance to current treatments. In the system described above, the increased levels of endogenous ROS produced by NOX4 is utilized by OC cells to stimulate tumorigenesis, angiogenesis, and treatment resistance (Figure 2). This adaptation in cellular signaling allows OC tumors to proliferate and develop resistance to chemotherapeutics through ROS production and upregulation of HER3, thus identifying this NOX4-driven pathway as a potential target for the treatment of chemoresistant tumors. In support of this, clinical trial studies show HER3 upregulation is associated with poor prognosis in OC, which serves as a clinical marker of tumor development, and HER3 expression is induced in response to current chemotherapeutics agents [186,187]. Therefore, this pathway provides an explanation for the ineffectiveness of traditional therapies for advanced OC and the development of therapeutic resistance. The implications of the findings reviewed here include the potential for NOX4 overexpression and increased levels of ROS to be utilized as a diagnostic biomarker in OC. Furthermore, there is clinical relevance for identifying new treatable targets in OC affected by this NOX4-driven system, particularly in resistant tumors.

As a significant mediator of miRNA dysregulation, ROS can have widespread effects on cellular processes. The roles of miRNA dysregulation in OC complicate the understanding of signaling pathways altered by tumors, with some acting as oncogenes and others acting as tumor suppressors. Similarly, the dysregulation of miRNAs in a cell-type-specific manner provides an opportunity to target specific miRNAs in different types of cancers. This could be accomplished by targeting the suppression of oncogenic miRNAs, which are typically upregulated in tumors, whereas the expression levels of tumor suppressor-like miRNAs are typically downregulated or lost in tumors [188]. The suppression of oncogenic miRNAs can be achieved with the use of anti-miRNA molecules targeting specific miRNA for inhibition or degradation [189]. For instance, anti-miR-21 treatment in breast cancer and glioblastoma induces apoptosis through the inhibition of PI3K signaling [190,191]. Alternatively, the upregulation of tumor suppressor miRNAs can be achieved with the use of miRNA mimics, which are delivered as mature miRNA molecules [192]. The use of miRNA mimics in combination with other forms of therapies improves treatment efficacy and the elimination of tumor cells. For example, the delivery of miR-204-5p in combination with oxaliplatin in colon cancer reduced tumor growth and induced apoptosis [193]. In further support of this, the treatment of relapsed, multidrug-resistant OC tumors with anti-Let-7 improved the efficacy of paclitaxel-induced cell death [194]. Since miRNAs are upstream regulators of a variety of cellular processes, the manipulation of their expression could cause adverse effects on surrounding tissues [195]. However, current research focusing on miRNA dysregulation is deciphering the mechanisms by which miRNAs affect different types of cancer. The increasing understanding of miRNA dysregulations in OC will allow for more direct targeting of the molecular pathways that are altered at each stage of tumor development. In addition, the up or downregulation of certain miRNAs in OC can also act as diagnostic biomarkers, as they have been demonstrated to have potential in many different cancer types [196]. Altogether, the altered molecular mechanisms driving OC development and treatment resistance are in part regulated by increased levels of endogenous ROS production and miRNA dysregulations. There are potentially new opportunities for more effective treatment of advanced OC by targeting the overlap in signaling pathways between these two mechanisms. However, limitations in our complete understanding of the roles of increased ROS and miRNA dysregulations in OC development necessitate more research efforts in these areas of study.

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References

- 1. Stewart, C.; Ralyea, C.; Lockwood, S. Ovarian Cancer: An Integrated Review. *Semin. Oncol. Nurs.* **2019**, *35*, 151–156. [CrossRef] [PubMed]
- 2. Siegel, R.L.; Miller, K.D.; Fuchs, H.E.; Jemal, A. Cancer Statistics, 2021. CA Cancer J. Clin. 2021, 71, 7–33. [CrossRef] [PubMed]
- Chen, V.W.; Ruiz, B.; Killeen, J.L.; Coté, T.R.; Wu, X.C.; Correa, C.N.; Howe, H.L. Pathology and classification of ovarian tumors. *Cancer* 2003, 97, 2631–2642. [CrossRef] [PubMed]
- 4. Reid, B.M.; Permuth, J.B.; Sellers, T.A. Epidemiology of Ovarian Cancer: A Review. Cancer Biol. Med. 2017, 14, 9–32.
- 5. Banerjee, S.; Kaye, S.B. New Strategies in the Treatment of Ovarian Cancer: Current Clinical Perspectives and Future Potential. *Clin. Cancer Res.* **2013**, *19*, 961–968. [CrossRef]
- 6. Jayson, G.C.; Kohn, E.C.; Kitchener, H.C.; Ledermann, J.A. Ovarian Cancer. Lancet 2014, 384, 1376–1388. [CrossRef]
- Shih, I.M.; Kurman, R.J. Ovarian Tumorigenesis: A Proposed Model Based on Morphological and Molecular Genetic Analysis. Am. J. Pathol. 2004, 164, 1511–1518. [CrossRef]
- 8. Rocco, J.W.; Sidransky, D. P16(Mts-1/Cdkn2/Ink4a) in Cancer Progression. Exp. Cell Res. 2001, 264, 42–55. [CrossRef]
- Berek, J.S.; Kehoe, S.T.; Kumar, L.; Friedlander, M. Cancer of the Ovary, Fallopian Tube, and Peritoneum. Int. J. Gynecol. Obstet. 2018, 143 (Suppl. 2), 59–78. [CrossRef]
- Bookman, M.A.; McGuire, W.P., 3rd; Kilpatrick, D.; Keenan, E.; Hogan, W.M.; Johnson, S.W.; O'Dwyer, P.; Rowinsky, E.; Gallion, H.H.; Ozols, R.F. Carboplatin and Paclitaxel in Ovarian Carcinoma: A Phase I Study of the Gynecologic Oncology Group. J. Clin. Oncol. 1996, 14, 1895–1902. [CrossRef]

- Bookman, M.A.; Brady, M.F.; McGuire, W.P.; Harper, P.G.; Alberts, D.S.; Friedlander, M.; Colombo, N.; Fowler, J.M.; Argenta, P.A.; De Geest, K.; et al. Evaluation of New Platinum-Based Treatment Regimens in Advanced-Stage Ovarian Cancer: A Phase III Trial of the Gynecologic Cancer InterGroup. J. Clin. Oncol. 2009, 27, 1419–1425. [CrossRef] [PubMed]
- Ozols, R.F.; Bundy, B.N.; Greer, B.E.; Fowler, J.M.; Clarke-Pearson, D.; Burger, R.A.; Mannel, R.S.; DeGeest, K.; Hartenbach, E.M.; Baergen, R.; et al. Phase III Trial of Carboplatin and Paclitaxel Compared With Cisplatin and Paclitaxel in Patients With Optimally Resected Stage III Ovarian Cancer: A Gynecologic Oncology Group Study. J. Clin. Oncol. 2003, 21, 3194–3200. [CrossRef] [PubMed]
- 13. McGuire, W.P.; Hoskins, W.J.; Brady, M.F.; Kucera, P.R.; Partridge, E.E.; Look, K.Y.; Clarke-Pearson, D.L.; Davidson, M. Cyclophosphamide and cisplatin versus paclitaxel and cisplatin: A phase III randomized trial in patients with suboptimal stage III/IV ovarian cancer (from the Gynecologic Oncology Group). *Semin. Oncol.* **1996**, 23 (Suppl. 2), 40–47. [PubMed]
- Armstrong, D.K.; Alvarez, R.D.; Bakkum-Gamez, J.N.; Barroilhet, L.; Behbakht, K.; Berchuck, A.; Chen, L.-M.; Cristea, M.; DeRosa, M.; Eisenhauer, E.L.; et al. Ovarian Cancer, Version 2.2020, NCCN Clinical Practice Guidelines in Oncology. *J. Natl. Compr. Cancer Netw.* 2021, 19, 191–226. [CrossRef] [PubMed]
- D'Autréaux, B.; Toledano, M.B. ROS as signalling molecules: Mechanisms that generate specificity in ROS homeostasis. *Nat. Rev. Mol. Cell Biol.* 2007, *8*, 813–824. [CrossRef]
- 16. Beckman, J.S.; Beckman, T.W.; Chen, J.; Marshall, P.A.; Freeman, B.A. Apparent hydroxyl radical production by peroxynitrite: Implications for endothelial injury from nitric oxide and superoxide. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 1620–1624. [CrossRef]
- 17. Negrini, S.; Gorgoulis, V.G.; Halazonetis, T.D. Genomic Instability—An Evolving Hallmark of Cancer. *Nat. Rev. Mol. Cell Biol.* **2010**, *11*, 220–228. [CrossRef]
- 18. Van Houten, B.; Woshner, V.; Santos, J.H. Role of mitochondrial DNA in toxic responses to oxidative stress. *DNA Repair* **2006**, *5*, 145–152. [CrossRef]
- Hurd, T.R.; Costa, N.J.; Dahm, C.; Beer, S.M.; Brown, S.E.; Filipovska, A.; Murphy, M. Glutathionylation of Mitochondrial Proteins. *Antioxidants Redox Signal.* 2005, 7, 999–1010. [CrossRef]
- 20. Rhee, S.G.; Kang, S.W.; Chang, T.-S.; Jeong, W.; Kim, K. Peroxiredoxin, a Novel Family of Peroxidases. *IUBMB Life* 2001, 52, 35–41. [CrossRef]
- Rhee, S.G.; Yang, K.S.; Kang, S.W.; Woo, H.A.; Chang, T.S. Controlled Elimination of Intracellular H₂O₂: Regulation of Peroxiredoxin, Catalase, and Glutathione Peroxidase Via Post-Translational Modification. *Antioxid. Redox Signal.* 2005, 7, 619–626. [CrossRef] [PubMed]
- 22. Imai, H.; Nakagawa, Y. Biological Significance of Phospholipid Hydroperoxide Glutathione Peroxidase (Phgpx, Gpx4) in Mammalian Cells. *Free. Radic. Biol. Med.* 2003, 34, 145–169. [CrossRef]
- Loschen, G.; Flohe, L.; Chance, B. Respiratory Chain Linked H₂O₂ Production in Pigeon Heart Mitochondria. *FEBS Lett.* 1971, 18, 261–264. [CrossRef]
- 24. Skulachev, V.P. Role of uncoupled and non-coupled oxidations in maintenance of safely low levels of oxygen and its one-electron reductants. *Q. Rev. Biophys.* **1996**, *29*, 169–202. [CrossRef] [PubMed]
- 25. Murphy, M.P. How mitochondria produce reactive oxygen species. Biochem. J. 2009, 417, 1–13. [CrossRef]
- 26. Rossi, F.; Zatti, M. Biochemical aspects of phagocytosis in poly-morphonuclear leucocytes. NADH and NADPH oxidation by the granules of resting and phagocytizing cells. *Experientia* **1964**, *20*, 21–23. [CrossRef]
- 27. Quie, P.G.; White, J.G.; Holmes, B.; Good, R.A. In Vitro Bactericidal Capacity of Human Polymorphonuclear Leukocytes: Diminished Activity in Chronic Granulomatous Disease of Childhood *. *J. Clin. Investig.* **1967**, *46*, 668–679. [CrossRef]
- Baehner, R.L.; Nathan, D.G. Leukocyte Oxidase: Defective Activity in Chronic Granulomatous Disease. Science 1967, 155, 835–836.
 [CrossRef]
- Holmes, B.; Page, A.R.; Good, R.A. Studies of the Metabolic Activity of Leukocytes from Patients with a Genetic Abnormality of Phagocytic Function*. J. Clin. Investig. 1967, 46, 1422–1432. [CrossRef]
- Berendes, H.; Bridges, R.A.; Good, R.A. A fatal granulomatosus of childhood: The clinical study of a new syndrome. *Minn. Med.* 1957, 40, 309–312.
- Royer-Pokora, B.; Kunkel, L.M.; Monaco, A.; Goff, S.C.; Newburger, P.; Baehner, R.L.; Cole, F.S.; Curnutte, J.T.; Orkin, S.H. Cloning the gene for an inherited human disorder—Chronic granulomatous disease—On the basis of its chromosomal location. *Nature* 1986, 322, 32–38. [CrossRef] [PubMed]
- 32. Suh, Y.-A.; Arnold, R.S.; Lassegue, B.; Shi, J.; Xu, X.X.; Sorescu, D.; Chung, A.B.; Griendling, K.K.; Lambeth, J.D. Cell transformation by the superoxide-generating oxidase Mox1. *Nature* **1999**, *401*, 79–82. [CrossRef] [PubMed]
- 33. Lambeth, J.; Cheng, G.; Arnold, R.S.; Edens, W.A. Novel homologs of gp91phox. *Trends Biochem. Sci.* 2000, 25, 459–461. [CrossRef]
- Xia, C.; Meng, Q.; Liu, L.-Z.; Rojanasakul, Y.; Wang, X.-R.; Jiang, B.-H. Reactive Oxygen Species Regulate Angiogenesis and Tumor Growth through Vascular Endothelial Growth Factor. *Cancer Res.* 2007, 67, 10823–10830. [CrossRef]
- 35. Valko, M.; Rhodes, C.J.; Moncol, J.; Izakovic, M.; Mazur, M. Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chem. Biol. Interact.* **2006**, *160*, 1–40. [CrossRef]
- Paravicini, T.M.; Touyz, R.M. Nadph Oxidases, Reactive Oxygen Species, and Hypertension: Clinical Implications and Therapeutic Possibilities. *Diabetes Care* 2008, 31 (Suppl. 2), S170–S180. [CrossRef]
- Martínez, M.C.; Andriantsitohaina, R. Reactive Nitrogen Species: Molecular Mechanisms and Potential Significance in Health and Disease. *Antioxidants Redox Signal.* 2009, 11, 669–702. [CrossRef]

- Cross, C.E.; Halliwell, B.; Borish, E.T.; Pryor, W.A.; Ames, B.N.; Saul, R.L.; Mccord, J.M.; Harman, D. Oxygen Radicals and Human Disease. Ann. Intern. Med. 1987, 107, 526–545. [CrossRef]
- Johnson, T.M.; Yu, Z.X.; Ferrans, V.J.; Lowenstein, R.A.; Finkel, T. Reactive oxygen species are downstream mediators of p53-dependent apoptosis. Proc. Natl. Acad. Sci. USA 1996, 93, 11848–11852. [CrossRef]
- 40. Dizdaroglu, M. Oxidatively Induced DNA Damage and Its Repair in Cancer. *Mutat. Res. Rev. Mutat. Res.* 2015, 763, 212–245. [CrossRef]
- Peluso, M.; Russo, V.; Mello, T.; Galli, A. Oxidative Stress and DNA Damage in Chronic Disease and Environmental Studies. *Int. J. Mol. Sci.* 2020, 21, 6936. [CrossRef] [PubMed]
- Yang, H.; Villani, R.M.; Wang, H.; Simpson, M.J.; Roberts, M.S.; Tang, M.; Liang, X. The role of cellular reactive oxygen species in cancer chemotherapy. J. Exp. Clin. Cancer Res. 2018, 37, 266. [CrossRef] [PubMed]
- Copin, J.-C.; Gasche, Y.; Chan, P.H. Overexpression of copper/zinc superoxide dismutase does not prevent neonatal lethality in mutant mice that lack manganese superoxide dismutase. *Free Radic. Biol. Med.* 2000, 28, 1571–1576. [CrossRef]
- Maiorino, F.M.; Brigelius-Flohé, R.; Aumann, K.; Roveri, A.; Schomburg, D.; Flohé, L. Diversity of Glutathione Peroxidases. Methods Enzymol. 1995, 252, 38–53.
- 45. Arnér, E.S.J.; Holmgren, A. Physiological functions of thioredoxin and thioredoxin reductase. *Eur. J. BioChem.* **2000**, 267, 6102–6109. [CrossRef]
- 46. Ray, P.D.; Huang, B.-W.; Tsuji, Y. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell. Signal.* **2012**, *24*, 981–990. [CrossRef]
- Khalil, H.S.; Goltsov, A.; Langdon, S.P.; Harrison, D.J.; Bown, J.; Deeni, Y. Quantitative analysis of NRF2 pathway reveals key elements of the regulatory circuits underlying antioxidant response and proliferation of ovarian cancer cells. *J. Biotechnol.* 2015, 202, 12–30. [CrossRef]
- 48. Liew, P.-L.; Hsu, C.-S.; Liu, W.-M.; Lee, Y.-C.; Lee, Y.-C.; Chen, C.-L. Prognostic and predictive values of Nrf2, Keap1, p16 and E-cadherin expression in ovarian epithelial carcinoma. *Int. J. Clin. Exp. Pathol.* **2015**, *8*, 5642–5649.
- 49. Czogalla, B.; Kahaly, M.; Mayr, D.; Schmoeckel, E.; Niesler, B.; Kolben, T.; Burges, A.; Mahner, S.; Jeschke, U.; Trillsch, F. Interaction of Eralpha and Nrf2 Impacts Survival in Ovarian Cancer Patients. *Int. J. Mol. Sci.* **2018**, 20, 112. [CrossRef]
- 50. Thompson, C.B. Apoptosis in the Pathogenesis and Treatment of Disease. Science 1995, 267, 1456–1462. [CrossRef]
- 51. Moloney, J.N.; Cotter, T.G. ROS signalling in the biology of cancer. *Semin. Cell Dev. Biol.* **2018**, *80*, 50–64. [CrossRef] [PubMed]
- 52. Chan, D.W.; Liu, V.W.; Tsao, G.S.; Yao, K.-M.; Furukawa, T.; Chan, K.K.; Ngan, H.Y. Loss of MKP3 mediated by oxidative stress enhances tumorigenicity and chemoresistance of ovarian cancer cells. *Carcinogenesis* **2008**, *29*, 1742–1750. [CrossRef] [PubMed]
- 53. Liu, L.-Z.; Hu, X.-W.; Xia, C.; He, J.; Zhou, Q.; Shi, X.; Fang, J.; Jiang, B.-H. Reactive oxygen species regulate epidermal growth factor-induced vascular endothelial growth factor and hypoxia-inducible factor-1α expression through activation of AKT and P70S6K1 in human ovarian cancer cells. *Free Radic. Biol. Med.* **2006**, *41*, 1521–1533. [CrossRef] [PubMed]
- 54. Valko, M.; Leibfritz, D.; Moncol, J.; Cronin, M.T.D.; Mazur, M.; Telser, J. Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. Cell Biol.* **2007**, *39*, 44–84. [CrossRef] [PubMed]
- 55. Tudek, B.; Winczura, A.; Janik, J.; Siomek, A.; Foksinski, M.; Oliński, R. Involvement of oxidatively damaged DNA and repair in cancer development and aging. *Am. J. Transl. Res.* **2010**, *2*, 254–284.
- 56. Fridovich, I. The biology of oxygen radicals. Science 1978, 201, 875-880. [CrossRef]
- 57. Meier, B.; Cross, A.R.; Hancock, J.T.; Kaup, F.J.; Jones, O.T.G. Identification of a superoxide-generating NADPH oxidase system in human fibroblasts. *Biochem. J.* 1991, 275, 241–245. [CrossRef]
- 58. Szatrowski, T.P.; Nathan, C.F. Production of large amounts of hydrogen peroxide by human tumor cells. *Cancer Res.* **1991**, *51*, 794–798.
- 59. Griendling, K.K.; Minieri, C.A.; Ollerenshaw, J.D.; Alexander, R.W. Angiotensin II stimulates NADH and NADPH oxidase activity in cultured vascular smooth muscle cells. *Circ. Res.* **1994**, *74*, 1141–1148. [CrossRef]
- De Carvalho, D.D.; Sadok, A.; Bourgarel-Rey, V.; Gattacceca, F.; Penel, C.; Lehmann, M.; Kovacic, H. Nox1 downstream of 12-lipoxygenase controls cell proliferation but not cell spreading of colon cancer cells. *Int. J. Cancer* 2008, 122, 1757–1764. [CrossRef]
- Kajla, S.; Mondol, A.S.; Nagasawa, A.; Zhang, Y.; Kato, M.; Matsuno, K.; Matsuno, K.; Yabe-Nishimura, C.; Kamata, T. A Crucial Role for Nox 1 in Redox-Dependent Regulation of Wnt-Beta-Catenin Signaling. *FASEB J.* 2012, 26, 2049–2059. [CrossRef] [PubMed]
- Mukawera, E.; Chartier, S.; Williams, V.; Pagano, P.J.; Lapointe, R.; Grandvaux, N. Redox-Modulating Agents Target Nox2-Dependent Ikkepsilon Oncogenic Kinase Expression and Proliferation in Human Breast Cancer Cell Lines. *Redox Biol.* 2015, 6, 9–18. [CrossRef] [PubMed]
- 63. Banskota, S.; Regmi, S.C.; Kim, J.-A. NOX1 to NOX2 switch deactivates AMPK and induces invasive phenotype in colon cancer cells through overexpression of MMP-7. *Mol. Cancer* 2015, *14*, 123. [CrossRef] [PubMed]
- 64. Wiktorin, H.G.; Nilsson, T.; Aydin, E.; Hellstrand, K.; Palmqvist, L.; Martner, A. Role of NOX2 for leukaemic expansion in a murine model of BCR-ABL1 + leukaemia. *Br. J. Haematol.* **2018**, *182*, 290–294. [CrossRef] [PubMed]
- Hole, P.S.; Zabkiewicz, J.; Munje, C.; Newton, Z.; Pearn, L.; White, P.; Marquez, N.; Hills, R.; Burnett, A.K.; Tonks, A.; et al. Overproduction of NOX-derived ROS in AML promotes proliferation and is associated with defective oxidative stress signaling. *Blood* 2013, 122, 3322–3330. [CrossRef] [PubMed]

- 66. Harrison, I.P.; Vinh, A.; Johnson, I.; Luong, R.; Drummond, G.R.; Sobey, C.G.; Tiganis, T.; Williams, E.; O'Leary, J.; Brooks, D.; et al. NOX2 oxidase expressed in endosomes promotes cell proliferation and prostate tumour development. *Oncotarget* 2018, 9, 35378–35393. [CrossRef]
- 67. Kim, S.-M.; Hur, D.Y.; Hong, S.-W.; Kim, J.H. EBV-encoded EBNA1 regulates cell viability by modulating miR34a-NOX2-ROS signaling in gastric cancer cells. *Biochem. Biophys. Res. Commun.* 2017, 494, 550–555. [CrossRef]
- Block, K.; Gorin, Y.; Hoover, P.; Williams, P.; Chelmicki, T.; Clark, R.A.; Yoneda, T.; Abboud, H.E. NAD(P)H Oxidases Regulate HIF-2α Protein Expression. J. Biol. Chem. 2007, 282, 8019–8026. [CrossRef]
- 69. Brar, S.S.; Kennedy, T.P.; Sturrock, A.B.; Huecksteadt, T.P.; Quinn, M.T.; Whorton, A.R.; Hoidal, J.R. An NAD(P)H oxidase regulates growth and transcription in melanoma cells. *Am. J. Physiol. Physiol.* 2002, 282, C1212–C1224. [CrossRef]
- 70. Diaz, B.; Shani, G.; Pass, I.; Anderson, D.; Quintavalle, M.; Courtneidge, S.A. Tks5-Dependent, Nox-Mediated Generation of Reactive Oxygen Species Is Necessary for Invadopodia Formation. *Sci. Signal.* **2009**, *2*, ra53. [CrossRef]
- Ogrunc, M.; Di Micco, R.; Liontos, M.; Bombardelli, L.; Mione, M.; Fumagalli, M.; Gorgoulis, V.G.; Daddadifagagna, F. Oncogeneinduced reactive oxygen species fuel hyperproliferation and DNA damage response activation. *Cell Death Differ.* 2014, 21, 998–1012. [CrossRef] [PubMed]
- Zhang, C.; Lan, T.; Hou, J.; Li, J.; Fang, R.; Yang, Z.; Zhang, M.; Liu, J.; Liu, B. NOX4 promotes non-small cell lung cancer cell proliferation and metastasis through positive feedback regulation of PI3K/Akt signaling. *Oncotarget* 2014, *5*, 4392–4405. [CrossRef]
- 73. Simone, N.L.; Soule, B.P.; Ly, D.; Saleh, A.D.; Savage, J.E.; DeGraff, W.; Cook, J.; Harris, C.C.; Gius, D.; Mitchell, J.B. Ionizing Radiation-Induced Oxidative Stress Alters miRNA Expression. *PLoS ONE* **2009**, *4*, e6377. [CrossRef] [PubMed]
- 74. Lu, J.; Getz, G.; Miska, E.A.; Alvarez-Saavedra, E.; Lamb, J.; Peck, D.; Sweet-Cordero, A.; Ebert, B.L.; Mak, R.H.; Ferrando, A.A.; et al. MicroRNA expression profiles classify human cancers. *Nature* **2005**, *435*, 834–838. [CrossRef]
- 75. Bartel, D.P. MicroRNAs: Target Recognition and Regulatory Functions. Cell 2009, 136, 215–233. [CrossRef] [PubMed]
- 76. Lee, R.C.; Feinbaum, R.L.; Ambros, V. The C. elegans heterochronic gene lin-4 encodes small RNAs with antisense complementarity to lin-14. *Cell* **1993**, *75*, 843–854. [CrossRef]
- 77. Wightman, B.; Ha, I.; Ruvkun, G. Posttranscriptional regulation of the heterochronic gene lin-14 by lin-4 mediates temporal pattern formation in C. elegans. *Cell* **1993**, *75*, 855–862. [CrossRef]
- 78. Lee, Y.; Ahn, C.; Han, J.; Choi, H.; Kim, J.; Yim, J.; Lee, J.; Provost, P.; Rådmark, O.; Kim, S.; et al. The nuclear RNase III Drosha initiates microRNA processing. *Nature* **2003**, *425*, 415–419. [CrossRef]
- 79. Han, J.; Lee, Y.; Yeom, K.-H.; Kim, Y.-K.; Jin, H.; Kim, V.N. The Drosha-DGCR8 complex in primary microRNA processing. *Genes Dev.* **2004**, *18*, 3016–3027. [CrossRef]
- Denli, A.M.; Tops, B.; Plasterk, R.H.A.; Ketting, R.F.; Hannon, G.J. Processing of primary microRNAs by the Microprocessor complex. *Nature* 2004, 432, 231–235. [CrossRef]
- Zhang, H.; Kolb, F.A.; Jaskiewicz, L.; Westhof, E.; Filipowicz, W. Single Processing Center Models for Human Dicer and Bacterial RNase III. Cell 2004, 118, 57–68. [CrossRef] [PubMed]
- 82. Heravi-Moussavi, A.; Anglesio, M.S.; Cheng, S.-W.G.; Senz, J.; Yang, W.; Prentice, L.; Fejes, A.P.; Chow, C.; Tone, A.; Kalloger, S.E.; et al. Recurrent Somatic *DICER1* Mutations in Nonepithelial Ovarian Cancers. *N. Engl. J. Med.* **2012**, *366*, 234–242. [CrossRef]
- 83. Bartel, D.P. Micrornas: Genomics, Biogenesis, Mechanism, and Function. Cell 2004, 116, 281–297. [CrossRef]
- Ramchandran, R.; Chaluvally-Raghavan, P. Mirna-Mediated Rna Activation in Mammalian Cells. Adv. Exp. Med. Biol. 2017, 983, 81–89. [PubMed]
- Calin, G.A.; Dumitru, C.D.; Shimizu, M.; Bichi, R.; Zupo, S.; Noch, E.; Aldler, H.; Rattan, S.; Keating, M.; Rai, K.; et al. Frequent deletions and down-regulation of micro- RNA genes *miR15* and *miR16* at 13q14 in chronic lymphocytic leukemia. *Proc. Natl. Acad. Sci. USA* 2002, 99, 15524–15529. [CrossRef]
- Bussing, I.; Slack, F.J.; Grosshans, H. Let-7 Micrornas in Development, Stem Cells and Cancer. Trends Mol. Med. 2008, 14, 400–409. [CrossRef]
- Wang, X.; Cao, L.; Wang, Y.; Wang, X.; Liu, N.; You, Y. Regulation of let-7 and its target oncogenes (Review). Oncol. Lett. 2012, 3, 955–960. [CrossRef]
- 88. Calin, G.A.; Croce, C.M. MicroRNA Signatures in Human Cancers. Nat. Rev. Cancer 2006, 6, 857–866. [CrossRef]
- 89. Garofalo, M.; Leva, G.D.; Croce, C.M. Micrornas as Anti-Cancer Therapy. Curr. Pharm. Des. 2014, 20, 5328–5335. [CrossRef]
- Kumar, M.; Lu, Z.; Takwi, A.A.L.; Chen, W.; Callander, N.S.; Ramos, K.S.; Young, K.H.; Li, Y. Negative regulation of the tumor suppressor p53 gene by microRNAs. *Oncogene* 2011, 30, 843–853. [CrossRef]
- Costinean, S.; Zanesi, N.; Pekarsky, Y.; Tili, E.; Volinia, S.; Heerema, N.; Croce, C.M. Pre-B Cell Proliferation and Lymphoblastic Leukemia/High-Grade Lymphoma in E(Mu)-Mir155 Transgenic Mice. Proc. Natl. Acad. Sci. USA 2006, 103, 7024–7029. [CrossRef]
- Valeri, N.; Gasparini, P.; Fabbri, M.; Braconi, C.; Veronese, A.; Lovat, F.; Adair, B.; Vannini, I.; Fanini, F.; Bottoni, A.; et al. Modulation of mismatch repair and genomic stability by miR-155. *Proc. Natl. Acad. Sci. USA* 2010, 107, 6982–6987. [CrossRef] [PubMed]
- O'Donnell, K.A.; Wentzel, E.A.; Zeller, K.I.; Dang, C.; Mendell, J.T. c-Myc-regulated microRNAs modulate E2F1 expression. *Nature* 2005, 435, 839–843. [CrossRef] [PubMed]
- 94. Sylvestre, Y.; De Guire, V.; Querido, E.; Mukhopadhyay, U.K.; Bourdeau, V.; Major, F.; Ferbeyre, G.; Chartrand, P. An E2F/miR-20a Autoregulatory Feedback Loop. *J. Biol. Chem.* 2007, 282, 2135–2143. [CrossRef] [PubMed]

- 95. Trimarchi, J.; Lees, J.A. Sibling rivalry in the E2F family. Nat. Rev. Mol. Cell Biol. 2002, 3, 11–20. [CrossRef]
- 96. Coller, H.A.; Forman, J.J.; Legesse-Miller, A. Myc'ed Messages: Myc Induces Transcription of E2f1 While Inhibiting Its Translation Via a Microrna Polycistron. *PLoS Genet.* **2007**, *3*, e146. [CrossRef] [PubMed]
- Woods, K.; Thomson, J.M.; Hammond, S.M. Direct Regulation of an Oncogenic Micro-RNA Cluster by E2F Transcription Factors. J. Biol. Chem. 2007, 282, 2130–2134. [CrossRef]
- 98. He, L.; Thomson, J.M.; Hemann, M.T.; Hernando-Monge, E.; Mu, D.; Goodson, S.; Powers, S.; Cordon-Cardo, C.; Lowe, S.W.; Hannon, G.J.; et al. A microRNA polycistron as a potential human oncogene. *Nature* **2005**, *435*, 828–833. [CrossRef]
- Galardi, S.; Mercatelli, N.; Giorda, E.; Massalini, S.; Frajese, G.V.; Ciafrè, S.A.; Farace, M.G. miR-221 and miR-222 Expression Affects the Proliferation Potential of Human Prostate Carcinoma Cell Lines by Targeting p27Kip1. *J. Biol. Chem.* 2007, 282, 23716–23724. [CrossRef]
- Le Sage, C.; Nagel, R.; Egan, D.A.; Schrier, M.; Mesman, E.; Mangiola, A.; Anile, C.; Maira, G.; Mercatelli, N.; Ciafrè, S.A.; et al. Regulation of the p27Kip1 tumor suppressor by miR-221 and miR-222 promotes cancer cell proliferation. *EMBO J.* 2007, 26, 3699–3708. [CrossRef]
- 101. Visone, R.; Russo, L.; Pallante, P.; De Martino, I.; Ferraro, A.; Leone, V.; Borbone, E.; Petrocca, F.; Alder, H.; Croce, C.M.; et al. MicroRNAs (miR)-221 and miR-222, both overexpressed in human thyroid papillary carcinomas, regulate p27Kip1 protein levels and cell cycle. *Endocr.-Relat. Cancer* 2007, 14, 791–798. [CrossRef] [PubMed]
- Rattanapan, Y.; Korkiatsakul, V.; Kongruang, A.; Siriboonpiputtana, T.; Rerkamnuaychoke, B.; Chareonsirisuthigul, T. MicroRNA Expression Profiling of Epithelial Ovarian Cancer Identifies New Markers of Tumor Subtype. *MicroRNA* 2020, *9*, 289–294. [CrossRef] [PubMed]
- 103. Hoeben, A.; Landuyt, B.; Highley, M.S.; Wildiers, H.; Van Oosterom, A.T.; De Bruijn, E.A. Vascular Endothelial Growth Factor and Angiogenesis. *Pharmacol. Rev.* 2004, *56*, 549–580. [CrossRef]
- Chung, A.S.; Ferrara, N. Developmental and Pathological Angiogenesis. Annu. Rev. Cell Dev. Biol. 2011, 27, 563–584. [CrossRef]
 [PubMed]
- Leung, D.W.; Cachianes, G.; Kuang, W.-J.; Goeddel, D.V.; Ferrara, N. Vascular Endothelial Growth Factor Is a Secreted Angiogenic Mitogen. Science 1989, 246, 1306–1309. [CrossRef] [PubMed]
- 106. Breier, G.; Albrecht, U.; Sterrer, S.; Risau, W. Expression of vascular endothelial growth factor during embryonic angiogenesis and endothelial cell differentiation. *Development* **1992**, *114*, 521–532. [CrossRef]
- Berse, B.; Brown, L.F.; Van De Water, L.; Dvorak, H.F.; Senger, D.R. Vascular Permeability Factor (Vascular Endothelial Growth Factor) Gene Is Expressed Differentially in Normal Tissues, Macrophages, and Tumors. *Mol. Biol. Cell* 1992, 3, 211–220. [CrossRef]
- 108. Muz, B.; de la Puente, P.; Azab, F.; Azab, A.K. The Role of Hypoxia in Cancer Progression, Angiogenesis, Metastasis, and Resistance to Therapy. *Hypoxia* 2015, *3*, 83–92. [CrossRef]
- 109. Semenza, G.L.; Wang, G.L. A nuclear factor induced by hypoxia via de novo protein synthesis binds to the human erythropoietin gene enhancer at a site required for transcriptional activation. *Mol. Cell. Biol.* **1992**, *12*, 5447–5454.
- 110. Wang, G.L.; Semenza, G.L. Characterization of hypoxia-inducible factor 1 and regulation of DNA binding activity by hypoxia. *J. Biol. Chem.* **1993**, *268*, 21513–21518. [CrossRef]
- 111. Forsythe, J.A.; Jiang, B.H.; Iyer, N.V.; Agani, F.; Leung, S.W.; Koos, R.D.; Semenza, G.L. Activation of vascular endothelial growth factor gene transcription by hypoxia-inducible factor 1. *Mol. Cell. Biol.* **1996**, *16*, 4604–4613. [CrossRef] [PubMed]
- 112. Semenza, G.L. Defining the role of hypoxia-inducible factor 1 in cancer biology and therapeutics. *Oncogene* **2010**, *29*, 625–634. [CrossRef]
- 113. Liu, W.-J.; Huang, Y.-X.; Wang, W.; Zhang, Y.; Liu, B.-J.; Qiu, J.-G.; Jiang, B.-H.; Liu, L.-Z. NOX4 Signaling Mediates Cancer Development and Therapeutic Resistance through HER3 in Ovarian Cancer Cells. *Cells* **2021**, *10*, 1647. [CrossRef] [PubMed]
- 114. He, J.; Xu, Q.; Jing, Y.; Agani, F.; Qian, X.; Carpenter, R.; Li, Q.; Wang, X.-R.; Peiper, S.S.; Lu, Z.; et al. Reactive oxygen species regulate ERBB2 and ERBB3 expression via miR-199a/125b and DNA methylation. *EMBO Rep.* 2012, 13, 1116–1122. [CrossRef] [PubMed]
- Druz, A.; Chu, C.; Majors, B.; Santuary, R.; Betenbaugh, M.; Shiloach, J. A Novel Microrna Mmu-Mir-466h Affects Apoptosis Regulation in Mammalian Cells. *Biotechnol. Bioeng.* 2011, 108, 1651–1661. [CrossRef]
- 116. Druz, A.; Betenbaugh, M.; Shiloach, J. Glucose Depletion Activates Mmu-Mir-466h-5p Expression through Oxidative Stress and Inhibition of Histone Deacetylation. *Nucleic Acids Res.* **2012**, *40*, 7291–7302. [CrossRef]
- 117. Liu, B.; Chen, Y.; Clair, D.K.S. ROS and p53: A versatile partnership. Free Radic. Biol. Med. 2008, 44, 1529–1535. [CrossRef]
- 118. Feng, X.; Wang, Z.; Fillmore, R.; Xi, Y. MiR-200, a new star miRNA in human cancer. Cancer Lett. 2014, 344, 166–173. [CrossRef]
- 119. Burk, U.; Schubert, J.; Wellner, U.; Schmalhofer, O.; Vincan, E.; Spaderna, S.; Brabletz, T. A reciprocal repression between ZEB1 and members of the miR-200 family promotes EMT and invasion in cancer cells. *EMBO Rep.* **2008**, *9*, 582–589. [CrossRef]
- Gutierrez-Uzquiza, A.; Arechederra, M.; Bragado, P.; Aguirre-Ghiso, J.A.; Porras, A. P38alpha Mediates Cell Survival in Response to Oxidative Stress Via Induction of Antioxidant Genes: Effect on the P70s6k Pathway. J. Biol. Chem. 2012, 287, 2632–2642. [CrossRef]
- 121. Mateescu, B.; Batista, L.; Cardon, M.; Gruosso, T.; De Feraudy, Y.; Mariani, O.; Nicolas, A.; Meyniel, J.-P.; Cottu, P.; Sastre-Garau, X.; et al. miR-141 and miR-200a act on ovarian tumorigenesis by controlling oxidative stress response. *Nat. Med.* 2011, 17, 1627–1635. [CrossRef]

- 122. Xie, Z.; Cao, L.; Zhang, J. miR-21 modulates paclitaxel sensitivity and hypoxia-inducible factor-1α expression in human ovarian cancer cells. *Oncol. Lett.* **2013**, *6*, 795–800. [CrossRef] [PubMed]
- 123. Lai, Y.; Zhang, X.; Zhang, Z.; Shu, Y.; Luo, X.; Yang, Y.; Wang, X.; Yang, G.; Li, L.; Feng, Y. The Microrna-27a: Zbtb10-Specificity Protein Pathway Is Involved in Follicle Stimulating Hormone-Induced Vegf, Cox2 and Survivin Expression in Ovarian Epithelial Cancer Cells. Int. J. Oncol. 2013, 42, 776–784. [CrossRef] [PubMed]
- Masoumi-Dehghi, S.; Babashah, S.; Sadeghizadeh, M. Microrna-141-3p-Containing Small Extracellular Vesicles Derived from Epithelial Ovarian Cancer Cells Promote Endothelial Cell Angiogenesis through Activating the Jak/Stat3 and Nf-Kappab Signaling Pathways. J. Cell Commun. Signal. 2020, 14, 233–244. [CrossRef]
- 125. He, L.; Zhu, W.; Chen, Q.; Yuan, Y.; Wang, Y.; Wang, J.; Wu, X. Ovarian cancer cell-secreted exosomal miR-205 promotes metastasis by inducing angiogenesis. *Theranostics* **2019**, *9*, 8206–8220. [CrossRef] [PubMed]
- 126. Chen, X.; Mangala, L.S.; Mooberry, L.; Bayraktar, E.; Dasari, S.K.; Ma, S.; Ivan, C.; Court, K.A.; Rodriguez-Aguayo, C.; Bayraktar, R.; et al. Identifying and targeting angiogenesis-Related microRNAs in ovarian cancer. *Oncogene* 2019, 38, 6095–6108. [CrossRef]
- 127. Lawler, P.; Lawler, J. Molecular Basis for the Regulation of Angiogenesis by Thrombospondin-1 and -2. *Cold Spring Harb. Perspect. Med.* **2012**, *2*, a006627. [CrossRef]
- 128. Ghafouri-Fard, S.; Shoorei, H.; Taher, M. Mirna Profile in Ovarian Cancer. Exp. Mol. Pathol. 2020, 113, 104381. [CrossRef]
- Xu, Q.; Liu, L.-Z.; Qian, X.; Chen, Q.; Jiang, Y.; Li, D.; Lai, L.; Jiang, B.-H. MiR-145 directly targets p70S6K1 in cancer cells to inhibit tumor growth and angiogenesis. *Nucleic Acids Res.* 2012, 40, 761–774. [CrossRef]
- Halvorsen, A.R.; Kristensen, G.; Embleton, A.; Adusei, C.; Barretina-Ginesta, M.P.; Beale, P.; Helland, Å. Evaluation of Prognostic and Predictive Significance of Circulating MicroRNAs in Ovarian Cancer Patients. Dis. Markers 2017, 2017, 3098542. [CrossRef]
- 131. An, Y.; Yang, Q. MiR-21 modulates the polarization of macrophages and increases the effects of M2 macrophages on promoting the chemoresistance of ovarian cancer. *Life Sci.* 2020, 242, 117162. [CrossRef] [PubMed]
- 132. Au Yeung, C.L.; Co, N.-N.; Tsuruga, T.; Yeung, T.-L.; Kwan, S.Y.; Leung, C.S.; Li, Y.; Lu, E.S.; Kwan, K.; Wong, K.-K.; et al. Exosomal transfer of stroma-derived miR21 confers paclitaxel resistance in ovarian cancer cells through targeting APAF1. *Nat. Commun.* 2016, 7, 11150. [CrossRef] [PubMed]
- Lenkala, D.; LaCroix, B.; Gamazon, E.; Geeleher, P.; Im, H.K.; Huang, R.S. The impact of microRNA expression on cellular proliferation. *Qual. Life Res.* 2014, 133, 931–938. [CrossRef] [PubMed]
- 134. Wang, Y.; Qiu, C.; Lu, N.; Liu, Z.; Jin, C.; Sun, C.; Bu, H.; Yu, H.; Dongol, S.; Kong, B. FOXD1 is targeted by miR-30a-5p and miR-200a-5p and suppresses the proliferation of human ovarian carcinoma cells by promoting p21 expression in a p53-independent manner. *Int. J. Oncol.* 2018, *52*, 2130–2142. [CrossRef]
- Chen, M.-W.; Yang, S.-T.; Chien, M.-H.; Hua, K.-T.; Wu, C.-J.; Hsiao, S.M.; Lin, H.; Hsiao, M.; Su, J.-L.; Wei, L.-H. The STAT3miRNA-92-Wnt Signaling Pathway Regulates Spheroid Formation and Malignant Progression in Ovarian Cancer. *Cancer Res.* 2017, 77, 1955–1967. [CrossRef]
- 136. Yoshimura, A.; Sawada, K.; Nakamura, K.; Kinose, Y.; Nakatsuka, E.; Kobayashi, M.; Miyamoto, M.; Ishida, K.; Matsumoto, Y.; Kodama, M.; et al. Exosomal miR-99a-5p is elevated in sera of ovarian cancer patients and promotes cancer cell invasion by increasing fibronectin and vitronectin expression in neighboring peritoneal mesothelial cells. *BMC Cancer* 2018, 18, 1065. [CrossRef]
- 137. Huh, J.H.; Kim, T.H.; Kim, K.; Song, J.-A.; Jung, Y.J.; Jeong, J.-Y.; Lee, M.J.; Kim, Y.K.; Lee, D.H.; An, H.J. Dysregulation of miR-106a and miR-591 confers paclitaxel resistance to ovarian cancer. *Br. J. Cancer* **2013**, *109*, 452–461. [CrossRef]
- 138. Van Jaarsveld, M.T.M.; Helleman, J.; Boersma, A.W.M.; van Kuijk, P.F.; van Ijcken, W.F.; Despierre, E.; Vergote, I.; Mathijssen, R.H.J.; Berns, E.M.J.J.; Verweij, J.; et al. miR-141 regulates KEAP1 and modulates cisplatin sensitivity in ovarian cancer cells. Oncogene 2013, 32, 4284–4293. [CrossRef]
- Parikh, A.; Lee, C.; Joseph, P.; Marchini, S.; Baccarini, A.; Kolev, V.; Romualdi, C.; Fruscio, R.; Shah, H.; Wang, F.; et al. microRNA-181a has a critical role in ovarian cancer progression through the regulation of the epithelial–mesenchymal transition. *Nat. Commun.* 2014, *5*, 2977. [CrossRef]
- Liu, Z.; Liu, J.; Segura, M.F.; Shao, C.; Lee, P.; Gong, Y.; Hernando, E.; Wei, J.-J. MiR-182 overexpression in tumourigenesis of high-grade serous ovarian carcinoma. J. Pathol. 2012, 228, 204–215. [CrossRef]
- 141. McMillen, B.D.; Aponte, M.M.; Liu, Z.; Helenowski, I.B.; Scholtens, D.M.; Buttin, B.M.; Wei, J.-J. Expression analysis of MIR182 and its associated target genes in advanced ovarian carcinoma. *Mod. Pathol.* **2012**, *25*, 1644–1653. [CrossRef] [PubMed]
- 142. Wang, Y.-Q.; Guo, R.-D.; Guo, R.-M.; Sheng, W.; Yin, L.-R. MicroRNA-182 promotes cell growth, invasion, and chemoresistance by targeting programmed cell death 4 (PDCD4) in human ovarian carcinomas. *J. Cell. Biochem.* 2013, 114, 1464–1473. [CrossRef] [PubMed]
- Xiaohong, Z.; Lichun, F.; Na, X.; Kejian, Z.; Xiaolan, X.; Shaosheng, W. MiR-203 promotes the growth and migration of ovarian cancer cells by enhancing glycolytic pathway. *Tumor Biol.* 2016, *37*, 14989–14997. [CrossRef] [PubMed]
- 144. Chu, P.; Liang, A.; Jiang, A.; Zong, L. miR-205 regulates the proliferation and invasion of ovarian cancer cells via suppressing PTEN/SMAD4 expression. *Oncol. Lett.* 2018, *15*, 7571–7578. [CrossRef] [PubMed]
- 145. Li, L.; Huang, K.; You, Y.; Fu, X.; Hu, L.; Song, L.; Meng, Y. Hypoxia-induced miR-210 in epithelial ovarian cancer enhances cancer cell viability via promoting proliferation and inhibiting apoptosis. *Int. J. Oncol.* **2014**, *44*, 2111–2120. [CrossRef]

- 146. Yang, H.; Kong, W.; He, L.; Zhao, J.-J.; O'Donnell, J.D.; Wang, J.; Wenham, R.M.; Coppola, D.; Kruk, P.A.; Nicosia, S.V.; et al. MicroRNA Expression Profiling in Human Ovarian Cancer: *miR-214* Induces Cell Survival and Cisplatin Resistance by Targeting *PTEN. Cancer Res.* 2008, 68, 425–433. [CrossRef] [PubMed]
- 147. Zhu, X.; Shen, H.; Yin, X.; Yang, M.; Wei, H.; Chen, Q.; Feng, F.; Liu, Y.; Xu, W.; Li, Y. Macrophages derived exosomes deliver miR-223 to epithelial ovarian cancer cells to elicit a chemoresistant phenotype. *J. Exp. Clin. Cancer Res.* **2019**, *38*, 81. [CrossRef]
- 148. Yang, L.; Wei, Q.-M.; Zhang, X.-W.; Sheng, Q.; Yan, X.-T. MiR-376a promotion of proliferation and metastases in ovarian cancer: Potential role as a biomarker. *Life Sci.* 2017, 173, 62–67. [CrossRef]
- 149. Furlong, F.; Fitzpatrick, P.; O'Toole, S.; Phelan, S.; McGrogan, B.; Maguire, A.; O'Grady, A.; Gallagher, M.; Prencipe, M.; McGoldrick, A.; et al. Low MAD2 expression levels associate with reduced progression-free survival in patients with high-grade serous epithelial ovarian cancer. *J. Pathol.* **2012**, *226*, 746–755. [CrossRef]
- Chaluvally-Raghavan, P.; Jeong, K.J.; Pradeep, S.; Silva, A.M.; Yu, S.; Liu, W.; Moss, T.; Rodriguez-Aguayo, C.; Zhang, D.; Ram, P.; et al. Direct Upregulation of STAT3 by MicroRNA-551b-3p Deregulates Growth and Metastasis of Ovarian Cancer. *Cell Rep.* 2016, 15, 1493–1504. [CrossRef]
- 151. Zhao, W.; Han, T.; Li, B.; Ma, Q.; Yang, P.; Li, H. miR-552 promotes ovarian cancer progression by regulating PTEN pathway. *J. Ovarian Res.* **2019**, *12*, 121. [CrossRef] [PubMed]
- Choi, Y.E.; Meghani, K.; Brault, M.-E.; Leclerc, L.; He, Y.; Day, T.A.; Elias, K.M.; Drapkin, R.; Weinstock, D.M.; Dao, F.; et al. Platinum and PARP Inhibitor Resistance Due to Overexpression of MicroRNA-622 in BRCA1-Mutant Ovarian Cancer. *Cell Rep.* 2016, 14, 429–439. [CrossRef] [PubMed]
- 153. Ying, X.; Li-Ya, Q.; Feng, Z.; Yin, W.; Ji-Hong, L. MiR-939 promotes the proliferation of human ovarian cancer cells by repressing APC2 expression. *Biomed. Pharmacother.* **2015**, *71*, 64–69. [CrossRef]
- 154. Kanlikilicer, P.; Bayraktar, R.; Denizli, M.; Rashed, M.H.; Ivan, C.; Aslan, B.; Mitra, R.; Karagoz, K.; Bayraktar, E.; Zhang, X.; et al. Exosomal Mirna Confers Chemo Resistance Via Targeting Cav1/P-Gp/M2-Type Macrophage Axis in Ovarian Cancer. *EBioMedicine* 2018, 38, 100–112. [CrossRef]
- 155. Creighton, C.J.; Fountain, M.D.; Yu, Z.; Nagaraja, A.K.; Zhu, H.; Khan, M.; Olokpa, E.; Zariff, A.; Gunaratne, P.H.; Matzuk, M.M.; et al. Molecular Profiling Uncovers a p53-Associated Role for MicroRNA-31 in Inhibiting the Proliferation of Serous Ovarian Carcinomas and Other Cancers. *Cancer Res.* 2010, 70, 1906–1915. [CrossRef] [PubMed]
- 156. Gong, L.; Wang, C.; Gao, Y.; Wang, J. Decreased expression of microRNA-148a predicts poor prognosis in ovarian cancer and associates with tumor growth and metastasis. *Biomed. Pharmacother.* **2016**, *83*, 58–63. [CrossRef]
- 157. Lin, Z.; Li, D.; Cheng, W.; Wu, J.; Wang, K.; Hu, Y. Microrna-181 Functions as an Antioncogene and Mediates Nf-Kappab Pathway by Targeting Rtkn2 in Ovarian Cancers. *Reprod. Sci.* 2019, *26*, 1071–1081. [CrossRef]
- 158. Liu, J.; Zhang, X.; Huang, Y.; Zhang, Q.; Zhou, J.; Zhang, X.; Wang, X. miR-200b and miR-200c co-contribute to the cisplatin sensitivity of ovarian cancer cells by targeting DNA methyltransferases. *Oncol. Lett.* **2019**, *17*, 1453–1460. [CrossRef]
- 159. Dai, C.; Xie, Y.; Zhuang, X.; Yuan, Z. Mir-206 Inhibits Epithelial Ovarian Cancer Cells Growth and Invasion Via Blocking C-Met/Akt/Mtor Signaling Pathway. *Biomed. Pharmacother.* **2018**, *104*, 763–770. [CrossRef]
- 160. Zhou, F.; Chen, J.; Wang, H. MicroRNA-298 inhibits malignant phenotypes of epithelial ovarian cancer by regulating the expression of EZH2. *Oncol. Lett.* **2016**, *12*, 3926–3932. [CrossRef]
- Liu, J.; Gu, Z.; Tang, Y.; Hao, J.; Zhang, C.; Yang, X. Tumour-suppressive microRNA-424-5p directly targets CCNE1 as potential prognostic markers in epithelial ovarian cancer. *Cell Cycle* 2018, *17*, 309–318. [CrossRef]
- Chen, K.; Zeng, J.; Tang, K.; Xiao, H.; Hu, J.; Huang, C.; Yao, W.; Yu, G.; Xiao, W.; Guan, W.; et al. miR-490-5p suppresses tumour growth in renal cell carcinoma through targeting PIK3CA. *Biol. Cell* 2016, 108, 41–50. [CrossRef]
- Hong, L.; Wang, Y.; Chen, W.; Yang, S. Microrna-508 Suppresses Epithelial-Mesenchymal Transition, Migration, and Invasion of Ovarian Cancer Cells through the Mapk1/Erk Signaling Pathway. J. Cell. Biochem. 2018, 119, 7431–7440. [CrossRef] [PubMed]
- 164. Parmar, M.K.; Ledermann, J.A.; Colombo, N.; Du, B.A.; Delaloye, J.F.; Kristensen, G.B.; Wheeler, S.; Swart, A.M.; Qian, W.; Torri, V.; et al. Paclitaxel Plus Platinum-Based Chemotherapy Versus Conventional Platinum-Based Chemotherapy in Women with Relapsed Ovarian Cancer: The Icon4/Ago-Ovar-2.2 Trial. *Lancet* 2003, 361, 2099–2106. [PubMed]
- 165. Neijt, J.P.; Engelholm, S.A.; Tuxen, M.K.; Sørensen, P.G.; Hansen, M.; Sessa, C.; de Swart, C.A.M.; Hirsch, F.R.; Lund, B.; van Houwelingen, H.C. Exploratory Phase III Study of Paclitaxel and Cisplatin Versus Paclitaxel and Carboplatin in Advanced Ovarian Cancer. J. Clin. Oncol. 2000, 18, 3084–3092. [CrossRef] [PubMed]
- 166. Schoch, S.; Gajewski, S.; Rothfuß, J.; Hartwig, A.; Köberle, B. Comparative Study of the Mode of Action of Clinically Approved Platinum-Based Chemotherapeutics. *Int. J. Mol. Sci.* **2020**, *21*, 6928. [CrossRef]
- 167. Schiff, P.; Fant, J.; Horwitz, S.B. Promotion of microtubule assembly in vitro by taxol. Nature 1979, 277, 665–667. [CrossRef]
- 168. Johnson, S.W.; Laub, P.B.; Beesley, J.S.; Ozols, R.F.; Hamilton, T.C. Increased platinum-DNA damage tolerance is associated with cisplatin resistance and cross-resistance to various chemotherapeutic agents in unrelated human ovarian cancer cell lines. *Cancer Res.* 1997, 57, 850–856.
- 169. Pfisterer, J.; Plante, M.; Vergote, I.; du Bois, A.; Hirte, H.; Lacave, A.J.; Wagner, U.; Stahle, A.; Stuart, G.; Kimmig, R.; et al. Gemcitabine Plus Carboplatin Compared with Carboplatin in Patients with Platinum-Sensitive Recurrent Ovarian Cancer: An Intergroup Trial of the Ago-Ovar, the Ncic Ctg, and the Eortc Gcg. J. Clin. Oncol. 2006, 24, 4699–4707. [CrossRef]

- 170. Sehouli, J.; Camara, O.; Schmidt, M.; Mahner, S.; Seipelt, G.; Otremba, B.; Schmalfeldt, B.; Tesch, H.; Lorenz-Schlüter, C.; Oskay-Ozcelik, G.; et al. Pegylated Liposomal Doxorubicin (Caelyx) in Patients with Advanced Ovarian Cancer: Results of a German Multicenter Observational Study. *Cancer Chemother. Pharmacol.* 2009, 64, 585–591. [CrossRef]
- 171. Ferrandina, G.; Ludovisi, M.; Lorusso, D.; Pignata, S.; Breda, E.; Savarese, A.; Del Medico, P.; Scaltriti, L.; Katsaros, D.; Priolo, D.; et al. Phase III Trial of Gemcitabine Compared with Pegylated Liposomal Doxorubicin in Progressive or Recurrent Ovarian Cancer. *J. Clin. Oncol.* **2008**, *26*, 890–896. [CrossRef]
- 172. Witham, J.; Valenti, M.R.; Alexis, K.; Vidot, S.; Eccles, S.A.; Kaye, S.B.; Richardson, A. The Bcl-2/Bcl-Xl Family Inhibitor Abt-737 Sensitizes Ovarian Cancer Cells to Carboplatin. *Clin. Cancer Res.* **2007**, *13*, 7191–7198. [CrossRef]
- 173. Eliopoulos, A.G.; Kerr, D.J.; Herod, J.; Hodgkins, L.; Krajewski, S.; Reed, J.C.; Young, L. The control of apoptosis and drug resistance in ovarian cancer: Influence of p53 and Bcl-2. *Oncogene* **1995**, *11*, 1217–1228.
- 174. Wang, H.; Zhang, Z.; Wei, X.; Dai, R. Small-molecule inhibitor of Bcl-2 (TW-37) suppresses growth and enhances cisplatin-induced apoptosis in ovarian cancer cells. *J. Ovarian Res.* 2015, *8*, 3. [CrossRef]
- 175. Zeitlin, B.D.; Zeitlin, I.J.; Nör, J.E. Expanding Circle of Inhibition: Small-Molecule Inhibitors of Bcl-2 as Anticancer Cell and Antiangiogenic Agents. J. Clin. Oncol. 2008, 26, 4180–4188. [CrossRef]
- 176. Safe, S.; Abdelrahim, M. Sp transcription factor family and its role in cancer. Eur. J. Cancer 2005, 41, 2438–2448. [CrossRef]
- 177. Stavrovskaya, A.A. Cellular Mechanisms of Multidrug Resistance of Tumor Cells. Biochemistry 2000, 65, 95–106.
- 178. Kruh, G.D.; Belinsky, M.G. The Mrp Family of Drug Efflux Pumps. Oncogene 2003, 22, 7537–7552. [CrossRef]
- 179. Liu, N.; Zhong, L.; Zeng, J.; Zhang, X.; Yang, Q.; Liao, D.; Wang, Y.; Chen, G.; Wang, Y. Upregulation of Microrna-200a Associates with Tumor Proliferation, Cscs Phenotype and Chemosensitivity in Ovarian Cancer. *Neoplasma* **2015**, *62*, 550–559. [CrossRef]
- Miyamoto, M.; Sawada, K.; Nakamura, K.; Yoshimura, A.; Ishida, K.; Kobayashi, M.; Shimizu, A.; Yamamoto, M.; Kodama, M.; Hashimoto, K.; et al. Paclitaxel exposure downregulates miR-522 expression and its downregulation induces paclitaxel resistance in ovarian cancer cells. *Sci. Rep.* 2020, *10*, 16755. [CrossRef]
- Wang, Y.; Chen, P.; Chen, X.; Gong, D.; Wu, Y.; Huang, L.; Chen, Y. ROS-Induced DCTPP1 Upregulation Contributes to Cisplatin Resistance in Ovarian Cancer. *Front. Mol. Biosci.* 2022, *9*, 838006. [CrossRef]
- 182. Li, J.; Zheng, C.; Wang, M.; Umano, A.D.; Dai, Q.; Zhang, C.; Huang, H.; Yang, Q.; Yang, X.; Lu, J.; et al. ROS-regulated phosphorylation of ITPKB by CAMK2G drives cisplatin resistance in ovarian cancer. *Oncogene* **2022**, *41*, 1114–1128. [CrossRef]
- 183. Kim, B.; Jung, J.W.; Jung, J.; Han, Y.; Suh, D.H.; Kim, H.S.; Dhanasekaran, D.N.; Song, Y.S. PGC1α induced by reactive oxygen species contributes to chemoresistance of ovarian cancer cells. *Oncotarget* 2017, *8*, 60299–60311. [CrossRef]
- 184. Uusi-Kerttula, H.; Davies, J.A.; Thompson, J.M.; Wongthida, P.; Evgin, L.; Shim, K.G.; Bradshaw, A.; Baker, A.T.; Rizkallah, P.J.; Jones, R.; et al. Ad5null-A20: A Tropism-Modified, Alphavbeta6 Integrin-Selective Oncolytic Adenovirus for Epithelial Ovarian Cancer Therapies. *Clin. Cancer Res.* 2018, 24, 4215–4224. [CrossRef]
- 185. Lanitis, E.; Dangaj, D.; Hagemann, I.; Song, D.-G.; Best, A.; Sandaltzopoulos, R.; Coukos, G.; Powell, D.J., Jr. Primary Human Ovarian Epithelial Cancer Cells Broadly Express HER2 at Immunologically-Detectable Levels. *PLoS ONE* 2012, 7, e49829. [CrossRef]
- Chung, Y.W.; Kim, S.; Hong, J.H.; Lee, J.K.; Lee, N.W.; Lee, Y.S.; Song, J.Y. Overexpression of Her2/Her3 and Clinical Feature of Ovarian Cancer. J. Gynecol. Oncol. 2019, 30, e75. [CrossRef]
- 187. Mizuno, T.; Kojima, Y.; Yonemori, K.; Yoshida, H.; Sugiura, Y.; Ohtake, Y.; Okuma, H.S.; Nishikawa, T.; Tanioka, M.; Sudo, K.; et al. Neoadjuvant chemotherapy promotes the expression of HER3 in patients with ovarian cancer. *Oncol. Lett.* 2020, 20, 336. [CrossRef]
- 188. Jansson, M.D.; Lund, A.H. Microrna and Cancer. Mol. Oncol. 2012, 6, 590-610. [CrossRef]
- Babar, I.A.; Cheng, C.J.; Booth, C.J.; Liang, X.; Weidhaas, J.B.; Saltzman, W.M.; Slack, F.J. Nanoparticle-based therapy in an in vivo microRNA-155 (miR-155)-dependent mouse model of lymphoma. *Proc. Natl. Acad. Sci. USA* 2012, 109, E1695–E1704. [CrossRef]
- Corsten, M.F.; Miranda, R.; Kasmieh, R.; Krichevsky, A.M.; Weissleder, R.; Shah, K. MicroRNA-21 Knockdown Disrupts Glioma Growth In vivo and Displays Synergistic Cytotoxicity with Neural Precursor Cell–Delivered S-TRAIL in Human Gliomas. *Cancer Res.* 2007, 67, 8994–9000. [CrossRef]
- 191. Yan, L.X.; Wu, Q.N.; Zhang, Y.; Li, Y.Y.; Liao, D.Z.; Hou, J.H.; Fu, J.; Zeng, M.S.; Yun, J.P.; Wu, Q.L.; et al. Knockdown of Mir-21 in Human Breast Cancer Cell Lines Inhibits Proliferation, in Vitro Migration and in Vivo Tumor Growth. *Breast Cancer Res.* 2011, 13, R2. [CrossRef]
- 192. Ji, W.; Sun, B.; Su, C. Targeting MicroRNAs in Cancer Gene Therapy. Genes 2017, 8, 21. [CrossRef]
- Yang, H.; Liu, Y.; Qiu, Y.; Ding, M.; Zhang, Y. MiRNA-204-5p and oxaliplatin-loaded silica nanoparticles for enhanced tumor suppression effect in CD44-overexpressed colon adenocarcinoma. *Int. J. Pharm.* 2019, 566, 585–593. [CrossRef]
- 194. Gandham, S.K.; Rao, M.; Shah, A.; Trivedi, M.S.; Amiji, M.M. Combination Microrna-Based Cellular Reprogramming with Paclitaxel Enhances Therapeutic Efficacy in a Relapsed and Multidrug-Resistant Model of Epithelial Ovarian Cancer. *Mol. Ther.-Oncolytics* 2022, 25, 57–68. [CrossRef]
- 195. Aagaard, L.; Rossi, J.J. Rnai Therapeutics: Principles, Prospects and Challenges. Adv. Drug Deliv. Rev. 2007, 59, 75–86. [CrossRef]
- Rupaimoole, R.; Slack, F.J. MicroRNA therapeutics: Towards a new era for the management of cancer and other diseases. *Nat. Rev. Drug Discov.* 2017, 16, 203–222. [CrossRef]