

11-26-2018

Comparison of Germline versus Somatic BAP1 Mutations for Risk of Metastasis in Uveal Melanoma.

K. G. Ewens
University of Pennsylvania


E. Lalonde
University of Pennsylvania

J. Richards-Yutz
University of Pennsylvania

C. L. Shields
Thomas Jefferson University

A. Ganguly
University of Pennsylvania

Follow this and additional works at: <https://jdc.jefferson.edu/willsfp>

 Part of the [Oncology Commons](#), and the [Ophthalmology Commons](#)

[Let us know how access to this document benefits you](#)

Recommended Citation

Ewens, K. G.; Lalonde, E.; Richards-Yutz, J.; Shields, C. L.; and Ganguly, A., "Comparison of Germline versus Somatic BAP1 Mutations for Risk of Metastasis in Uveal Melanoma." (2018). *Wills Eye Hospital Papers*. Paper 91.
<https://jdc.jefferson.edu/willsfp/91>


This Article is brought to you for free and open access by the Jefferson Digital Commons. The Jefferson Digital Commons is a service of Thomas Jefferson University's [Center for Teaching and Learning \(CTL\)](#). The Commons is a showcase for Jefferson books and journals, peer-reviewed scholarly publications, unique historical collections from the University archives, and teaching tools. The Jefferson Digital Commons allows researchers and interested readers anywhere in the world to learn about and keep up to date with Jefferson scholarship. This article has been accepted for inclusion in Wills Eye Hospital Papers by an authorized administrator of the Jefferson Digital Commons. For more information, please contact: JeffersonDigitalCommons@jefferson.edu.

RESEARCH ARTICLE

Open Access



Comparison of Germline versus Somatic *BAP1* Mutations for Risk of Metastasis in Uveal Melanoma

K. G. Ewens¹, E. Lalonde¹, J. Richards-Yutz¹, C. L. Shields² and A. Ganguly^{1*} 

Abstract

Background: Germline mutations in *BAP1* have been associated with *BAP1*-Tumor Predisposition Syndrome (*BAP1*-TPDS), a predisposition to multiple tumors within a family that includes uveal melanoma (UM), cutaneous melanoma, malignant mesothelioma and renal cell carcinoma. Alternatively, somatic mutations in *BAP1* in UM have been associated with high risk for metastasis. In this study, we compare the risk of metastasis in UM that carry germline versus somatic *BAP1* mutations and mutation-negative tumors.

Methods: DNA extracted from 142 UM and matched blood samples was sequenced using Sanger or next generation sequencing to identify *BAP1* gene mutations.

Results: Eleven of 142 UM (8%) carried germline *BAP1* mutations, 43 (30%) had somatic mutations, and 88 (62%) were mutation-negative. All *BAP1* mutations identified in blood samples were also present in the matched UM. There were 52 unique mutations in 54 tumors. All were pathogenic or likely pathogenic.

A comparison of tumors carrying somatic vs. germline mutations, or no mutations, showed a higher frequency of metastasis in tumors carrying somatic mutations: 74% vs. 36%, $P=0.03$ and 74% vs. 26% $P<0.001$, respectively. Tumors with a somatic mutation compared to mutation-negative had an older age of diagnosis of (61.8 vs. 52.2 years, $P=0.002$), and shorter time to metastasis (16 vs. 26 months, $P=0.04$). Kaplan-Meier analysis further showed that tumors with somatic (vs. germline) mutations demonstrated a greater metastatic risk ($P=0.03$). Cox multivariate analysis showed in addition to chromosome-3 monosomy and larger tumor diameter, the presence of *BAP1* somatic, but not germline mutations, was significantly associated with risk of metastasis ($P=0.02$). Personal or family history of *BAP1*-TPDS was available for 79 of the cases. All eight cases with germline mutations reported a history of *BAP1*-TPDS, which was significantly greater than what was observed in cases with somatic mutations (10 of 23, $P=0.009$) or mutation-negative cases (11 of 48, $P<0.001$).

Conclusions: Defining germline vs. somatic nature of *BAP1* mutations in UM can inform the individual about both the risk of metastasis, and the time to metastasis, which are critically important outcomes for the individual. This information can also change the cascade screening and surveillance of family members.

Keywords: uveal melanoma, ocular/eye cancer, *BAP1*, germline mutations, somatic mutations

* Correspondence: ganguly@penntestmed.upenn.edu

¹Department of Genetics, Perelman School of Medicine, University of Pennsylvania, 415 Curie Blvd, Philadelphia, Pennsylvania 19104-6145, United States

Full list of author information is available at the end of the article



© The Author(s). 2018 **Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated.

Background

The *BRCA1-associated protein 1* (*BAP1*) gene is located on chromosome 3p21 and encodes a nuclear localized, ubiquitin carboxy-terminal hydrolase tumor suppressor protein [1, 2]. *BAP1* has been shown to bind to the *BRCA1* protein enhancing *BRCA1*-mediated tumor suppression, and is involved in various biological processes including DNA damage response, regulation of the cell cycle and cell growth [2, 3]. Germline *BAP1* mutations have been associated with hereditary predisposition to multiple different cancers that include uveal and cutaneous melanoma, malignant mesothelioma on exposure to asbestos, renal cell carcinoma and other cancer types, such as lung adenocarcinoma and meningioma, that are collectively referred to as *BAP1* Tumor Predisposition Syndrome (*BAP1*-TPDS, OMIM #614327) [4, 5]. In vivo studies in melanocytic cells, have shown that *BAP1* is involved in the maintenance of a melanocytic phenotype; the depletion of *BAP1* protein levels result in dedifferentiation of cells and the acquisition of a more primitive, stem cell like phenotype [6]. It is well established that the *BAP1* region of chromosome 3p is commonly deleted in several cancers including melanomas, breast and lung cancers, among others [1, 7].

Uveal melanoma (UM) is a rare malignant tumor of the eye with a frequency of 5.1 per million in the United States [8], and is the cancer most commonly associated with the *BAP1*-TPDS [9]. The prognosis for UM is poor in approximately 50% of cases primarily due to metastasis to the liver within a short time [10, 11]. Factors such as large tumor size, location and chromosome 3 monosomy [12–18], as well as a specific 12-gene expression pattern [19–21] have all been associated with an increased risk of metastasis. In addition, the presence of inactivating somatic [22–29] or germline [30] *BAP1* mutations often in conjunction with chromosome 3 monosomy, loss of *BAP1* expression, or lack of immunohistochemical staining, have all been associated with metastasizing UM.

Germline *BAP1* mutations have been identified in approximately 2–5% of UM unselected for the presence of any high risk of hereditary cancers [30–33]. These studies reported that metastasis occurred more frequently in UM carrying germline mutations in the blood compared to *BAP1* mutation-negative controls, but the differences were significant only in one study [30], possibly due to the small number of tumors with germline mutations. In studies where DNA from both tumor tissue and blood was sequenced to rule out the presence of germline mutations, the frequency of somatic *BAP1* mutations was considerably higher than that observed in tumors with germline mutations, approaching 50% [23, 27, 34]. In other studies where somatic mutations were detected by *BAP1* expression assays and/or negative immunohistochemical staining

methods, as well as sequencing, the lack of *BAP1* expression was also significantly associated with the risk of metastasis [22–29]. While these studies show a strong association between somatic *BAP1* mutations and metastasis, the effect of germline changes remains unclear.

Most studies of *BAP1* mutation status in UM have been carried out using enucleated UM which may not be representative of how UM are currently treated, especially smaller tumors that are treated with globe sparing procedures. In this study of *BAP1* mutations in UM, the presence of germline or somatic mutations was determined using sequence analysis of DNA extracted from biopsies of both enucleated globes and fine needle aspirations (FNA) and from matched blood samples. Demographic and clinical characteristics of the tumors and the frequency of metastasis were compared among tumors with germline or somatic *BAP1* mutations and mutation-negative UM.

Methods

UM cases

A total of 142 UM cases managed by the Ocular Oncology Service at Wills Eye Hospital, Thomas Jefferson University, Philadelphia, PA, USA between 1998 and 2016 were evaluated in this study of somatic and germline *BAP1* mutations. Twenty-three tumor samples were obtained by solid open biopsy of enucleated UM and 119 by fine needle aspirate (FNA) biopsies. Matched peripheral blood samples were obtained from all 142 individuals. Data on demographic and clinical characteristics, metastatic status, location of metastasis and relevant follow-up times were obtained by retrospective chart reviews. Written informed consent was obtained for all individuals at the time of chromosome testing of the tumor DNA. This research was approved by the Institutional Review Board of the University of Pennsylvania, and is in accordance with the Declaration of Helsinki.

The 142 UM were initially collected from two cohorts. Cohort 1 cases (N=90) were chosen from a series of tumors diagnosed between 1998 and 2013 based on availability of matched tumor and blood samples. Cohort 2 consisted of 52 cases diagnosed between 2008 and 2016 for which *BAP1* testing was requested, typically due to a personal and/or family history of cancer. The decision to combine the two sets of tumors into a single cohort for analysis was based on the finding that cohort membership was not a significant variable in either univariate ($P=0.53$) or multivariate ($P=0.65$) Cox regression analysis (Table 1). In addition, there was no significant difference between the two cohorts in the relative number of tumors carrying germline or somatic *BAP1* mutations and mutation-negative tumors ($P=0.76$), or in

Table 1 Demographic and tumor characteristics for 142 UM and assessment of their association with metastasis evaluated by Cox univariate regression for each of the nine variables alone, and multivariate regression that included all variables described in the table

Variables	All tumors N=142 (frequency)	Univariate regression			Multivariate regression		
		P-value	Hazard ratio	95% confidence intervals	P-value	Hazard ratio	95% confidence intervals
Cohort							
1	90 (0.63)		reference			reference	
2	52 (0.37)	0.53	0.82	0.44-1.53	0.65	1.18	0.57-2.48
Source of biopsied sample ^a							
FNA, N=119 (0.84)			reference			reference	
BAP1 mutation negative	73 (0.61)						
BAP1 mutation positive	46 (0.39)						
Enucleated tumor, N= 23 (0.16)		0.01	2.12	1.19-3.76	0.08	2.04	0.92-4.52
BAP1 mutation negative	15 (0.65)						
BAP1 mutation positive	8 (0.35)						
Age (years)		0.04	1.02	1.00-1.04	0.03	1.02	1.00-1.05
Median	56.7						
Mean±SD	55.4±14.7						
Range	14-88						
Sex							
Male	77 (0.54)		reference			reference	
Female	65 (0.46)	0.24	0.73	0.43-1.24	0.73	0.90	0.50-1.63
Chromosome 3							
Disomy	63 (0.44)		reference			reference	
Monosomy, partial monosomy (N=2), mosaic (N=5)	79 (0.56)	<0.001	4.64	2.46-8.78	0.008	2.99	1.33-6.72
Tumor diameter (mm)		<0.001	1.25	1.17-1.35	<0.001	1.20	1.10-1.31
Median	12.0						
Mean±SD	12.5±4.1						
Range	5.0-22.0						
Tumor thickness (mm)		<0.001	1.22	1.13-1.31	0.61	0.97	0.87-1.09
Median	5.7						
Mean±SD	6.2±3.3,						
Range	1.0-16.5						
Ciliary body involvement							
Absent	110 (0.78)	<0.001	reference			reference	
Present	32 (0.22)		3.05	1.79-5.20	0.15	1.54	0.85-2.80
BAP1							
Negative	88 (0.62)		reference			reference	
Somatic	43 (0.30)	<0.001	4.81	2.79-8.28	0.02	2.20	1.13-4.30
Germline	11 (0.08)	0.33	1.70	0.59-4.91	0.81	0.87	0.28-2.72

^aThere was no significant difference in the number of BAP1 mutation negative tumors and those carrying germline or somatic mutations UM biopsies from FNA samples compared to enucleated tumors (P=0.79, Fisher Exact test)
Figures in bold indicate significant P-values <0.05

the number of cases with personal or family history of BAP1-TPDS cancers (P=0.35, see below and Additional file 1: Table S1). There was a significant difference in the time to metastasis (P=0.01) and in the number of

metastases (P=0.003) between the two cohorts, possibly due to a longer follow-up time for the cohort 1 tumors (median=85 months, range 6-191) compared to 24 months (range 4-77 months, P<0.001, Additional file 1:

Table S1) for cohort 2. However, taking both metastasis and follow-up time into account in a Kaplan-Meier analysis, there was no significant association between cohort membership and metastasis ($P=0.52$, Additional file 1: Figure S1).

A personal or family history of *BAP1*-TPDS syndrome, defined following the guidelines found in OMIM #614327 [4] and Pilarski et al [5], was available for a subset of the UM cases. Individuals having at least two *BAP1*-TPDS tumors (UM, cutaneous melanoma, malignant mesothelioma, or renal cell carcinoma) themselves, or one in the UM case plus at least one in a first or second degree relative were considered as indicative of *BAP1*-TPDS syndrome. Cases with non-specific cancer diagnoses were not included as *BAP1*-TPDS syndromic tumors,

***BAP1* mutation screening and chromosome 3 copy number analysis**

Genomic DNA was extracted from peripheral blood lymphocytes using the Gentra Puregene Blood Kit (Cat No.158489, Qiagen) following the manufacturer's protocol, and from tumor samples as described previously [15, 35]. Sanger sequencing of DNA from matched peripheral blood lymphocytes was used to determine the presence of a germline mutation as noted by Abdel-Rahman et al [36]. DNA from enucleated tumor or FNA biopsies was sequenced to identify *BAP1* mutations using either Sanger or next generation sequencing of all coding exons and adjacent intronic regions [37, 38]. Sequence alignment and variant calling was performed as described previously [37]. The 2015 ACMG guidelines were followed in determining the degree pathogenicity of all sequence variants [39].

Chromosome 3 copy number was determined by microsatellite analysis ($N=51$) or whole genome SNP array (Affymetrix Human 100K, SNP-5.0 or SNP-6.0 and Cytoscan HD genotyping arrays, Affymetrix, Santa Clara CA) [15, 35]. Tumors were categorized as either disomy-3 ($N=63$) or monosomy ($N=79$) that included 72 tumors with complete monosomy-3, two with partial monosomy and five with a mosaic monosomy 3 indicating tumor heterogeneity of cells with chromosome-3 disomy and monosomy.

Statistical analyses

Continuous variables (age, tumor diameter and thickness) were compared using a Mann-Whitney U-test, and discrete variables (source of tumor sample, cohort 1 or 2, metastasis status, sex, tumor location and chromosome 3 copy number) using Fisher Exact or chi-square tests (vassarstats.net). Cox univariate and multivariate proportional hazard regression was used to determine the association of clinical and tumor characteristics with metastasis.

The Kaplan-Meier method was used to generate metastasis-free survival curves that were compared using a log-rank test. SPSS 24 (IBM, New York, NY) was used for survival analysis procedures. All tests were two-tailed and P-values less than 0.05 were considered statistically significant.

Results

UM individuals and tumor characteristics

Of the 142 UM characterized in this study, 54 (38%) carried a *BAP1* gene mutation. Eleven (8%) of the mutations were germline and 43 (30%) were somatic. The remaining 88 (62%) were *BAP1* mutation-negative. The eleven germline mutations were all retained in the matched UM samples. Thus, there were no cases where a germline mutation was lost in tumors, with or without monosomy-3.

Demographic and tumor characteristics for the 142 UM cases are shown in Table 1. The median age of all individuals was 56.7 years (range=14-88), and 77 (54%) were males. Overall, 63 (44%) of the tumors carried two copies of chromosome 3 (disomy), and 79 (56%) carried one copy of chromosome 3 (monosomy), partial monosomy or mosaic. The median tumor diameter and thickness was 12 mm (range=5.0-22.0) and 5.7 mm (range 1.0-16.5), respectively. The majority of the tumors had no ciliary body involvement ($N=110$, 78%). Most tumors were located only in the choroid ($N=109$, 77%), 26 (18%) were in the choroid plus ciliary body, three (2%) in the ciliary body, one (1%) in the iris, two (2%) in the iris plus ciliary body, and one (1%) in the iris plus ciliary body and choroid. Fifty-nine tumors (42%) metastasized within a median time of 19 months (range=0-107) while 83 (58%) did not metastasize during the observed follow-up period (median=56 months, range=4-191 months, Table 2). Thirty-six of 54 tumors with *BAP1* mutations metastasized (67%) compared to only 23 of 88 (26%) mutation-negative tumors ($P<0.001$) within 2-107 months.

BAP1 status was determined for 119 (84%) FNA biopsies and 23 enucleated tumors (Table 1). There was a significant difference in the number of metastases depending on the source of the biopsied sample: 43 of 119 (36%) tumors biopsied from FNA metastasized compared to 16 of 23 from enucleated globes (70%, $P=0.006$). The source of the biopsied sample was also significantly associated with metastasis in a Cox univariate ($P=0.01$), but not in a multivariate analysis (0.08, Table 1). This is not surprising, however, given that some of the classic prognostic factors for UM metastasis, larger tumor diameter and thickness and tumor location which warrant enucleation, were significantly different between tumors sampled from FNAs or enucleated tumor samples ($P=0.002$, >0.001 and 0.02, respectively

Table 2 Description of demographic and tumor variables with pairwise comparisons for 11 tumors with germline BAP1 mutations, 43 with *BAP1* somatic mutations and 88 with no *BAP1* mutations

Variables	<i>BAP1</i> germline N=11 (0.08) (frequency)	<i>BAP1</i> somatic N=43 (0.30) (frequency)	<i>BAP1</i> negative N=88 (0.62) (frequency)	Pairwise comparisons (P-value)		
				Germline/ Somatic	Germline/ Negative	Somatic/ Negative
Metastases						
Yes N=59 (0.42)	4 (0.36)	32 (0.74)	23 (0.26)	0.03^a	0.72 ^a	<0.001^a
No N=83 (0.58)	7 (0.64)	11 (0.26)	65 (0.74)			
Time to metastasis (months) (N=59)	N=4 (0.07)	N=32 (0.54)	N=23 (0.38)	0.07 ^b	0.50 ^b	0.04^c
Median (19.0 months)	37.5	16.0	26.0			
Mean±SD (26.4±20.9)	35.0±15.8	23±22.5	29.7±19.2			
Range (0-107)	16-49	2-45,90,107 ^d	0-84			
Follow-up time (months) in tumors with no metastasis (N=83)	N=7 (0.08)	N=11 (0.13)	N=65 (0.78)	0.59 ^c	0.25 ^c	0.33 ^c
Median (56.0 months)	24	54.0	58.0			
Mean±SD (59.8±39.0)	43.7±39.3	49.9±30.0	63.2±40.1			
Range (4-191)	4-109	8-90	5-191			
Cohort						
1	6 (0.54)	27 (0.63)	57 (0.65)	0.73 ^a	0.74 ^a	0.85 ^a
2	5 (0.46)	16 (0.37)	31 (0.35)			
Source of biopsied sample						
FNA	10 (0.91)	36 (0.84)	73 (0.83)	0.68 ^a	0.69 ^a	1.00 ^a
Enucleated tumor	1 (0.09)	7 (0.16)	15 (0.17)			
<i>BAP1</i> -TPDS personal or family history (N=79 reports) ^e						
Yes, syndromic tumors present (N=29, 0.37)	8 (1.00)	10 (0.44)	11 (0.23)	0.009^a	<0.001^a	0.10
No syndromic tumors (N=50, 0.63)	0	13 (0.56)	37 (0.77)			
Age (years)						
Median	59.0	61.8	52.2	0.07 ^c	0.86 ^c	0.002^c
Mean±SD	51.9±13.6	61.2±12.8	53.1±15.0,			
Range	22-67	28-88	14-84			
Sex						
Male	7 (0.64)	25 (0.58)	45 (0.51)	1.00 ^a	0.53 ^a	0.46 ^a
Female	4 (0.36)	18 (0.42)	43 (0.49)			
Chromosome 3						
Disomy	1 (0.09)	3 (0.07)	59 (0.67)	1.0 ^a	<0.001^a	<0.001^a
Monosomy, partial monosomy (N=2), mosaic (N=5)	10 (0.91)	40 (0.93)	26 (0.33)			
Tumor diameter (mm)						
Median	14.0	14.0	11.5	0.70 ^c	0.23 ^c	0.003^c
Mean±SD	13.4±4.4	13.9±3.9	11.7±4.0,			
Range	8.0-20.0	6.0-21.5	5.0-22.0			
Tumor thickness (mm)						
Median	6.6	7.0	5.0	0.41 ^c	0.51 ^c	0.002^c
Mean±SD	6.5±3.6	7.5±3.5	5.6±2.9,			
Range	1.5-12.3	2.0-16.5	1.0-13.1			

Table 2 Description of demographic and tumor variables with pairwise comparisons for 11 tumors with germline BAP1 mutations, 43 with BAP1 somatic mutations and 88 with no BAP1 mutations (Continued)

Variables	BAP1 germline	BAP1 somatic	BAP1 negative	Pairwise comparisons (P-value)		
	N=11 (0.08) (frequency)	N=43 (0.30) (frequency)	N=88 (0.62) (frequency)	Germline/ Somatic	Germline/ Negative	Somatic/ Negative
Ciliary body involvement						
Absent	8 (0.73)	28 (0.65)	74 (0.84)	0.73 ^a	0.40 ^a	0.02^a
Present	3 (0.27)	15 (0.35)	14 (0.14)			

^aAssociation test performed using two-tailed Fisher Exact or Chi-square tests^bReported P-values are calculated from a normal approximation of the Mann Whitney test statistic^cTests of means of quantitative variables performed using Mann-Whitney U tests^dThe time for metastasis for 30 of the tumors with somatic mutations was 2-45 months. The time to metastasis for the remaining two tumors was 90 and 107 months^eBAP1-TPDS, BAP1-tumor predisposition syndrome as defined in OMIM #614327 [4] and Pilarski et al [5]

Figures in bold indicate significant P-values <0.05

(data not shown). Importantly, there was no difference in the number of tumors carrying a germline or somatic mutation or mutation-negative tumors in enucleated tumors compared to FNA samples ($P=0.79$, Table 1 footnote^a).

In a Cox univariate analysis, the presence of a somatic, but not a germline BAP1 mutation (hazard ratio (HR) =4.81, $P<0.0001$; HR=1.7, $P=0.33$, respectively, Table 1) was significantly associated with metastasis. Other tumor variables that were significant in the univariate regression analysis included tumor source (FNA or enucleated tumor, HR= 2.12; $P=0.01$), age (HR=1.02, $P=0.04$), chromosome 3 monosomy, larger tumor diameter and thickness, and ciliary body involvement (HR=4.64, 1.25 and 1.22, respectively; all $P<0.001$). Considering all variables in a multivariate analysis, the presence of a somatic BAP1 mutation (HR=2.20, $P=0.02$), age (HR=1.02, $P=0.03$) tumor diameter (HR=1.20, $P<0.001$), and chromosome 3 monosomy (HR=2.99, $P=0.008$) remained significant, while the presence of a germline mutation was not significant (HR=0.87, $P=0.81$).

BAP1 Gene Mutations

Fifty-two unique BAP1 mutations were identified in 54 tumors: 43 tumors (30%) carried somatic mutations, while 11 tumors (8%) carried germline mutations (Table 3). All mutations were defined as pathogenic or likely pathogenic. These included seven (13%) missense and 15 (28%) splice-site mutations, three (6%) in-frame deletions and 29 mutations causing premature truncation, including one large deletion encompassing exons 15-17 and the intervening introns. There was no significant difference in the frequencies of these categories of mutations found in UM with germline compared to somatic mutations ($P=1.0$), nor in tumors that metastasized compared to those that did not ($P=0.53$). Thirty-six (67%) of the mutations were in the catalytic UCH domain. One splice-site mutation, c.438-1A>G was present as a germline mutation in one

tumor (UM-23) and as a somatic mutation in two tumors (UM-11 and UM-27). Thirty-three of these mutations were previously described in UM by Ewens et al [37] and 13 have been identified in UM by others (as noted in Table 3). Although the number of germline mutations was small ($N=11$), the locations of these were spread throughout the gene, similar to the distribution of somatic mutations (Table 3). Therefore, no new information was inferred from the location, or the associated functional domains, that would make the germline mutations have unique characteristic compared to somatic mutations.

Characterizations of UM with and without BAP1 mutations

Clinical and tumor characteristics of UM carrying BAP1 germline or somatic mutations and mutation-negative tumors, as well as pairwise comparisons of the three groups are presented in Table 2. There was no significant difference in the cohort, source of biopsy sample, or sex of the individuals among the three categories of mutations. The age at diagnosis of individuals with somatic mutations was significantly older (61.8 years, range=28-88) than individuals with mutation-negative tumors (median=52.2 years, range=14-84 years, $P=0.002$) and approached significance compared to individuals with a tumor carrying a germline mutation (median=59.0 years, range=22-67, $P=0.07$).

There was a significant difference in the comparison of chromosome 3 monosomy in mutation-negative tumors compared to tumors with either germline or somatic mutations. Only 26 (33%) mutation-negative tumors carried monosomy 3, while ten (91%) tumors with germline mutations and 40 (93%) with somatic mutations were chromosome 3 monosomy ($P<0.001$ in both cases). A comparison of chromosome 3 copy number between tumors with germline vs. somatic mutations (91% vs. 93%) was not significant ($P=1.0$, Table 2). There were four tumors with BAP1 mutations and chromosome 3 disomy (marked with superscript b in Table 3). These would suggest that chromosome 3 loss was not a

Table 3 Fifty-two unique germline and somatic *BAP1* mutations identified in 54 UM

ID	BAP1 mutation	Metastases	chr 3	DNA change	Protein change	Predicted effect	Predicted pathogenic effect ^a	Previously reported mutations in UM
UM_44	germline	yes	monosomy	exon 1	c.3G>A	p.Met1Ile	missense: start site lost	[36]
UM_76	somatic	yes	monosomy	intron 1	c.38-1G>C	p.?	splicing	[36]
UM_9	somatic	yes	monosomy	exon 2	c.40_52del13	p.Leu14SerfsTer54	truncating	[36]
UM_1136	germline	no	monosomy	exon 2	c.58_59insTG	p.Glu20ValfsTer53	truncating	
UM_113	somatic	yes	monosomy	intron 2	c.67+1G>A	p.?	splicing	[27, 36]
UM_1123	somatic	yes	monosomy	intron 2	c.67+2dupT	p.?	splicing	
UM_1080	somatic	no	monosomy	intron 2/exon 3	c.68-12_75del20	p.?	splicing	
UM_802	somatic	yes	monosomy	intron 2/exon 3	c.68-16_69del18	p.?	splicing	[36]
UM_584	germline	no	monosomy	exon 3	c.79delG	p.Val27CysfsTer45	truncating	
UM_17	somatic	yes	monosomy	exon 3	c.82C>T	p.Gln28Ter	truncating	[36]
UM_1207	somatic	no	monosomy	exon 3	c.91_93delGAG	p.Glu31del	in-frame deletion	
UM_1133	germline	no	disomy ^b	intron 3	c.122+1G>C	p.?	splicing	
UM_109	germline	yes	monosomy	intron 3	c.122+1G>T	p.?	splicing	[36]
UM_75	somatic	no	monosomy	exon 4	c.125_145del21	p.Pro42_Phe48del	in-frame deletion	[36]
UM_88	somatic	yes	monosomy	exon 4	c.145delC	p.Leu49CysfsTer23	truncating	[36]
UM_877	somatic	yes	monosomy	exon 4	c.165_180del16	p.Arg57SerfsTer10	truncating	
UM_13	somatic	yes	disomy ^b	exon 4	c.178C>T	p.Arg60Ter	truncating	[36, 45]
UM_62	somatic	yes	monosomy	exon 4	c.202_227del26	p.Asp68CysfsTer3	truncating	[36]
UM_780	somatic	yes	monosomy	exon 4/intron 4	c.234del96	p.?	splicing	
UM_119	somatic	yes	monosomy	exon 4	c.253C>T	p.Gln85Ter	truncating	[36]
UM_106	somatic	yes	monosomy	exon 4	c.254A>C	p.Gln85Pro	missense	[36]
UM_1126	somatic	no	monosomy	exon 5	c.295_312del18	p.Val99_Ser104del	in-frame deletion	
UM_1046	somatic	no	monosomy	exon5/intron5	c.370_375+12del18	p.?	splicing	
UM_1208	somatic	yes	monosomy	intron 5/exon 6	c.376-20_383del28	p.?	splicing	
UM_56	somatic	yes	monosomy	exon 6	c.422A>G	p.His141Arg	missense	[36, 46]
UM_23	germline	yes	monosomy	intron 6	c.438-2A>G	p.?	splicing	[36]
UM_11	somatic	no	disomy ^b	intron 6	c.438-2A>G	p.?	splicing	
UM_27	somatic	yes	monosomy	intron 6	c.438-2A>G	p.?	splicing	
UM_58	germline	no	monosomy	exon 7	c.458_459delCT	p.Pro153ArgfsTer7	truncating	[9, 36]
UM_114	somatic	yes	monosomy	exon 7	c.497_509del13	p.Glu166ValfsTer17	truncating	[36]
UM_51	somatic	no	monosomy	exon 7	c.506A>C	p.His169Pro	missense	[36]
UM_1118	somatic	no	monosomy	exon 7	c.524C>G	p.Pro175Arg	missense	[27]

Table 3 Fifty-two unique germline and somatic BAP1 mutations identified in 54 UM (Continued)

ID	BAP1 mutation	Metastases	chr 3	DNA change	Protein change	Predicted effect	Predicted pathogenic effect ^a	Previously reported mutations in UM
UM_1086	somatic	yes	monosomy	intron 7/exon 8	p.?	splicing	P	
UM_60	somatic	yes	monosomy	exon 8	p.Trp196Ter	truncating	P	[23, 47]
UM_1334	germline	no	mosaic	exon 8	p.Arg207GlyfsTer24	truncating	P	
UM_105	somatic	yes	monosomy	intron 8	p.?	splicing	P	[27, 36]
UM_35	somatic	yes	monosomy	exon 9	c.659+1G>A	truncating	P	[36, 48:c.723T>C]
UM_107	somatic	yes	monosomy	exon 9	c.723T>A	truncating	P	[36, 49] rs772448753 ^c
UM_46	somatic	yes	monosomy	exon 9	c.781C>T	truncating	P	
UM_1029	somatic	yes	monosomy	exon 10	c.904_905insT	truncating	P	[36]
UM_115	somatic	yes	monosomy	exon 12	c.1134_1143del10_insAA	truncating	P	
UM_55	somatic	yes	monosomy	exon 12	c.1153C>T	truncating	P	[32, 36, 44, 50]
UM_55	somatic	no	monosomy	exon 12	c.1175_1182delACGAGTAC	truncating	P	[36]
UM_863	somatic	no	monosomy	exon 12	c.1192G>T	truncating	P	
UM_950	germline	yes	mosaic	exon 12	c.1203dupT	truncating	P	
UM_69	somatic	yes	monosomy	exon 12	c.1217_1220delAGGA	truncating	P	[36]
UM_1333	germline	no	mosaic	exon 13	c.1695dupT	truncating	P	
UM_48	somatic	no	monosomy	exon 13	c.1729G>C	missense	P	[36]
UM_61	somatic	yes	monosomy	exon 14	c.1881C>G	truncating	P	[36]
UM_74	germline	no	monosomy	exon 14	c.1882_1885delTCAC	truncating	P	[29, 36, 51]
UM_708	somatic	no	disomy ^b	intron 14-3'UTR	p.Ser628ProfsTer8	truncating (3 exon deletion)	P	
UM_1113	somatic	yes	monosomy	exon 15	c.1890+38_2573del	truncating	P	
UM_104	somatic	yes	monosomy	exon 15	c.1926_1951del 26	truncating	P	[36]
UM_804	somatic	yes	monosomy	exon 16	c.1932_1948del17	truncating	P	[36]
UM_108	somatic	yes	monosomy	exon 16	c.1986_1989delTGAT	truncating	P	[36]
					p.Asp672Gly	missense	LP	[23, 36]

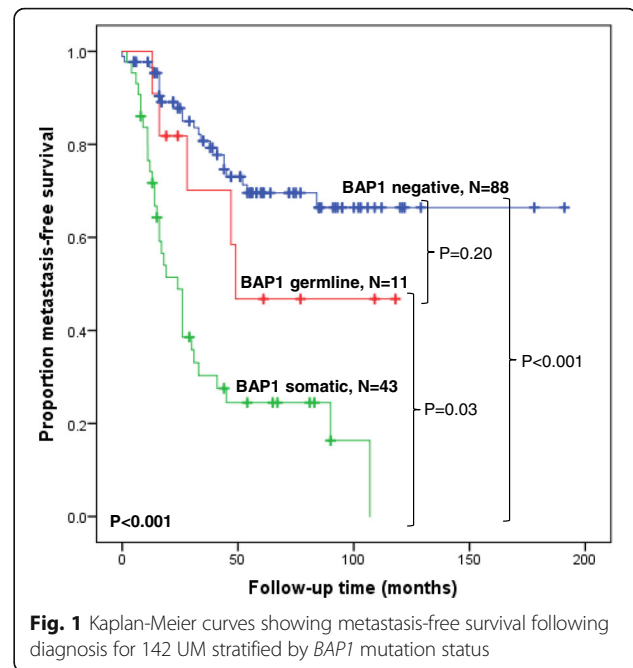
^aPredicted pathogenic effect based on ACMG guidelines [39] P, pathogenic; LP, likely pathogenic^bIndicates UM with chromosome 3 disomy and carrying a germline or somatic BAP1 mutation^cPopulation frequency rs772448753: 8.2E-06

necessary consequence of *BAP1* loss. The one germline mutation was a splice-site mutation, intron 3, c.122+1G>C. The three somatic mutations included one splice-site mutation (intron 6, c.438-2A>G) and two truncating mutations: exon 4, c.178C>T (p.Arg60X) and a large deletion of exons 15-17 and the intervening introns. The tumor carrying the exon 4, c.178C>T (p.Arg60X) truncating mutation metastasized at 11 months, but the other three tumors had not metastasized within 24 months (c.122+1G>C) or within at least 65 months (c.438-2A>G and the large three exon deletion).

Tumors with *BAP1* somatic mutations had a significantly larger median diameter (14.0 mm, range=6.0-21.5) and thickness (7.0 mm, range=2.0-16.5) compared to mutation negative tumors (median diameter=11.5 mm, range=5.0-22.0; thickness=5.0 mm, range=0.0-13.1, $P=0.003$ and $P=0.002$ respectively, Table 2). However, there was no significant difference in the tumor diameter or thickness between *BAP1* germline tumors and tumors with either somatic mutations ($P=0.70$ and $P=0.41$, respectively), or *BAP1*-negative tumors ($P=0.23$ and $P=0.51$, respectively). Table 2 shows that there was also a significant difference in ciliary body involvement of tumors carrying a somatic mutation, where the tumor was more likely to have ciliary body involvement (33%) compared to *BAP1*-negative tumors (14%, $P=0.04$), but not compared with those with a germline mutation (27%, $P=1.0$).

Tumors carrying *BAP1* somatic mutations metastasized significantly more often (32 of 43, 74%) compared to those with either *BAP1* germline mutations (4 of 11, 36%, $P=0.03$) or to *BAP1* mutation-negative tumors (23 of 88, 26%, $P<0.001$, Table 2). However, there was no significant difference in the frequency of metastasis between tumors with germline mutations and *BAP1*-negative tumors ($P=0.72$). This finding was corroborated by the multivariate regression analyses shown in Table 1 indicating that somatic, but not germline *BAP1* mutations, were significantly associated with metastasis ($P=0.03$ and 0.70 , respectively). Kaplan-Meier analysis (Figure 1) also showed that tumors with a *BAP1* somatic mutation had a significantly poorer metastatic outcome compared to those with germline mutations ($P=0.03$) or mutation-negative tumors ($P<0.001$), but the difference between tumors with germline mutations and *BAP1* mutation-negative tumors was not significant ($P=0.23$). The time to metastasis was also significantly shorter in the tumors with somatic mutations (median=16.0 months, range=2-107) compared to *BAP1*-negative tumors (median=26 months, range=0-84 months, $P=0.04$, Table 2).

A positive personal and/or family history of *BAP1*-TPDS tumors was reported for 29 (37%) of the 79 cases that provided information of other cancers in themselves



or their families (Table 2). There were eight cases that carried a germline mutation with information on personal or familial cancers; all had a positive history of *BAP1*-TPDS. Comparisons of *BAP1*-TPDS tumors in UM cases with a germline (8, 100%) vs. somatic mutation (10, 44%) or germline vs. mutation-negative (11, 23%) were significant ($P=0.009$ and $P<0.001$, respectively), while a comparison of those with somatic mutations was not significantly different from those with mutation-negative tumors ($P=0.10$).

Discussion

Inactivating somatic mutations in the *BAP1* gene in UM were first identified and associated with metastatic disease in 2010 [23]. Subsequent to this original publication, there have been numerous studies characterizing *BAP1* somatic [22, 24–29, 40–43] and germline [9, 30, 32, 33] mutations in UM. However, to our knowledge, this is the first report directly comparing risk of metastasis in tumors carrying germline or somatic *BAP1* mutations and mutation-negative tumors. In this study, the presence of somatic mutations was strictly defined as those found in tumors, and not in matched blood samples. A second important difference in this study is that, compared to most other reports where UM tumors analyzed for *BAP1* mutations were comprised primarily of samples biopsied from larger, enucleated tumors, 84% of the samples analyzed in this study were from FNA biopsies. We found no significant difference in the frequency of somatic and germline mutations in biopsies from FNAs compared to enucleated tumors. Since the

majority of UM are currently being treated by globe sparing procedures, it was important to determine whether the consequences of carrying a *BAP1* mutation identified in larger enucleated tumors could be applied to smaller UM biopsied by FNA sampling.

It is well established that tumor characteristics such as tumor diameter, ciliary body involvement and chromosome 3 monosomy are all associated with poor prognosis as measured by the development of metastases within 48 months after the primary is treated [12–18]. It has also been shown that these same variables are significantly associated with *BAP1* mutation status in tumors with somatic or germline mutations [24, 25, 28, 30]. In this study, multivariate regression analysis showed that in addition to these classic tumor variables, the presence of somatic, but not germline, *BAP1* mutations was significantly associated with metastasis (HR=2.20, $P=0.02$; HR=0.87, $P=0.81$, respectively). Kaplan-Meier analysis further showed that tumors with somatic mutations were associated with the poorest metastatic outcome, while there was no significant difference between tumors carrying a germline mutation and mutation-negative tumors. Furthermore, the time to metastasis was seven months shorter for individuals whose tumors carried somatic mutations compared to mutation-negative tumors ($P=0.04$) suggesting a more aggressive phenotype. Thus, in a previous publication [37] we showed that tumors with any mutation in the *BAP1* gene, in combination with chromosome 3-monosomy, have the highest risk of metastasis (HR=11.5). In this manuscript we show that monosomy 3 plus a somatic mutation in the *BAP1* gene, as opposed to a germline mutation, have a significantly higher risk of metastasis ($P=0.01$, H.R.=2.81) and a shorter time to metastasis compared to the ones with a germline mutation.

Given that somatic *BAP1* mutations appear to have a highly significant association with poor metastatic prognosis, it is surprising that the age of diagnosis of the individuals whose tumors carried somatic mutations was significantly older (median=61.8 years) compared to those with mutation-negative tumors (median=52.2 years, $P=0.002$). Previous studies have also reported that tumors with somatic mutations were diagnosed at least 8 years later than mutation-negative controls [24, 27, 29], but the significance of this observation is not clear.

Among the 54 tumors with *BAP1* mutations, there were four with chromosome 3 disomy. It cannot be ruled out that although these tumors appeared to be disomy-3, small deletions not detectable on the SNP arrays leading to partial monosomy or heterogeneity of the tumor sample could be responsible for this finding. Alternatively, in some cases it is possible that during tumor evolution, *BAP1* mutation on one copy of chromosome 3 could precede the loss of the other copy of chromosome 3. While several studies have found total

concordance between the presence *BAP1* mutations and monosomy 3 [23, 27, 40], others using sequencing, gene expression, and/or immunostaining methods have identified *BAP1* mutations in a small number of tumors with disomy 3 [24, 25, 28, 44].

One interesting observation in this study was the presence of *BAP1*-TPDS history in 44% of somatic cases and 23% of mutation negative tumors. The data on *BAP1*-TPDS was based on self-reported personal and family history of cancer. We classified individuals as having *BAP1*-TPDS if they carried at least two relevant tumors in their personal or family history. Since it is self-reported, the information can be imprecise, and there is a possibility for over/under representation of the *BAP1*-TPDS cases as well.

While the presence of germline *BAP1* mutations in cancers associated with *BAP1*-TPSD is well documented, the role of somatic mutations in this syndrome is not well studied. It is possible that the presence of TPDS in our cohort of UM cases that have a somatic *BAP1* mutation or are mutation-negative could be interpreted as evidence for the existence of a second type of the syndrome associated with mutations in a second gene different from *BAP1*.

A major strength of this study is that it includes UM sampled from enucleated tumors and FNAs with both somatic and germline *BAP1* mutations. However, it is limited by the small number of tumors with germline mutations. It is important to confirm these findings in a larger cohort of UM.

Conclusions

This study identified significant differences in the risk of metastasis for individuals whose tumors carry somatic vs. germline *BAP1* mutations. While overall 36 of 59 (61%) metastasizing tumors carried a *BAP1* mutation, only 7% carried germline mutations, compared to 54% with somatic mutations. From the viewpoint of counseling individuals with UM, there are different implications depending on whether the mutation is classified as germline or somatic. The most relevant information for the individuals with UM is the risk of metastasis, as well as an estimate of metastasis-free survival. The presence of germline mutations can provide the individual with information concerning the risk of development of other cancers themselves or in other family members, while information about the presence of somatic mutations is relevant to their individual risk of developing metastases within a shorter time period. Thus it is important to determine whether a tumor carries a *BAP1* mutation, and if positive, to also evaluate a matched blood sample to establish whether the mutation is germline or somatic. In addition, this information can also change the cascade screening and surveillance of the family members.

Additional file

Additional file 1: Table S1. Comparison of demographic and tumor variables of two cohorts of 142 UM. Description of demographic and tumor variables with pairwise comparisons for 142 divided into two cohorts depending on whether tumors were selected from archived samples for which both tumor and blood samples were available (cohort 1, N=90) or tumors for which *BAP1* sequencing was specifically requested (cohort 2, N=52). References: 1. OMIM: Tumor predisposition syndrome; TPDS. <https://www.omim.org/entry/614327>. 2. Pilarski R, Rai K, Cebulla C, Abdel-Rahman M. BAP1 Tumor Predisposition Syndrome. 2016 Oct 13 In: Adam MP, Ardinger HH, Pagon RA, Wallace SE, Bean LJH, Stephens K, Amemiya A, editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2018. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK390611/>. **Figure S1.** Kaplan-Meier analysis comparing metastasis-free survival of two cohorts of 142 UM. Kaplan-Meier curves showing metastasis-free survival following treatment for 142 UM stratified by cohort, as described in the legend for Additional file 1: Table S1. (DOCX 81 kb)

Abbreviations

BAP1: BRCA1-associated protein 1; *BAP1*-TPDS: *BAP1* tumor predisposition syndrome; FNA: Fine needle aspirate; GDL: Genetic Diagnostic Laboratory, University of Pennsylvania, Perelman School of Medicine; UM: Uveal melanoma; vs: Versus

Acknowledgements

The authors wish to thank Jessica Ebrahimzadeh for her service in reviewing case charts.

Funding

This work was supported in part by a grant from the National Cancer Institute (5R21CA181935) to AG, and internal Research and Development funds of the Genetic Diagnostic Laboratory.

Availability of data and materials

The datasets analyzed for this study are available from the corresponding author upon reasonable request.

Authors' contributions

KGE participated in data collection and analysis and manuscript preparation; EL contributed to statistical analysis and manuscript review; J R-Y was responsible for BAP1 sequence data and analysis; CLS provided all case samples and access to charts, participated in the design of the project and review of the manuscript; AG conceived the study and was involved in all aspects of data collection, analysis and manuscript preparation. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Written informed consent was obtained at the time of testing the tumor samples from all individuals. This research was approved by the Institutional Review Board of the University of Pennsylvania, and is in accordance with the Declaration of Helsinki.

Consent for publication

Not applicable

Competing interests

The authors declare they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Department of Genetics, Perelman School of Medicine, University of Pennsylvania, 415 Curie Blvd, Philadelphia, Pennsylvania 19104-6145, United States. ²Oncology Services, Wills Eye Hospital, Thomas Jefferson University, 840 Walnut St, Suite #1440, Philadelphia, Pennsylvania 19107, United States.

Received: 6 March 2018 Accepted: 12 November 2018

Published online: 26 November 2018

References

- Jensen DE, Proctor M, Marquis ST, Gardner HP, Ha SI, Chodosh LA, et al. BAP1: a novel ubiquitin hydrolase which binds to the BRCA1 RING finger and enhances BRCA1-mediated cell growth suppression. *Oncogene*. 1998; 16(9):1097–112.
- Jensen DE, Rauscher FJ. BAP1, a candidate tumor suppressor protein that interacts with BRCA1. *Ann N Y Acad Sci*. 1999;886(1):191–4.
- Ventii KH, Devi NS, Friedrich KL, Chernova TA, Tighiouart M, Van Meir EG, et al. BRCA1-associated protein-1 is a tumor suppressor that requires deubiquitinating activity and nuclear localization. *Can Res*. 2008;68(17): 6953–62.
- OMIM: Tumor predisposition syndrome; TPDS. <https://www.omim.org/entry/614327>.
- Pilarski R, Rai K, Cebulla C, Abdel-Rahman M. BAP1 Tumor Predisposition Syndrome. 2016 Oct 13 In: Adam MP, Ardinger HH, Pagon RA, Wallace SE, Bean LJH, Stephens K, Amemiya A, editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2018. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK390611/>.
- Matatall KA, Agapova OA, Onken MD, Worley LA, Bowcock AM, Harbour JW. BAP1 deficiency causes loss of melanocytic cell identity in uveal melanoma. *BMC Cancer*. 2013; 13:371–371.
- Angeloni D. Molecular analysis of deletions in human chromosome 3p21 and the role of resident cancer genes in disease. *Brief Funct Genomics*. 2007;6(1):19–39.
- Singh AD, Turell ME, Topham AK. Uveal Melanoma: Trends in incidence, treatment, and survival. *Ophthalmology*. 2011;118(9):1881–5.
- Rai K, Pilarski R, Cebulla CM, Abdel-Rahman MH. Comprehensive review of BAP1 tumor predisposition syndrome with report of two new cases. *Clin Genet*. 2016;89(3):285–94.
- Kujala E, Mäkitie T, Kivelä T. Very long-term prognosis of patients with malignant uveal melanoma. *Invest Ophthalmol Vis Sci*. 2003;44(11):4651–9.
- Lorigan JG, Wallace S, Mavligit GM. The prevalence and location of metastases from ocular melanoma: imaging study in 110 patients. *Am J Roentgenol*. 1991;157(6):1279–81.
- Aalto Y, Eriksson L, Seregard S, Larsson O, Knuutila S. Concomitant loss of chromosome 3 and whole arm losses and gains of chromosome 1, 6, or 8 in metastasizing primary uveal melanoma. *Invest Ophthalmol Vis Sci*. 2001;42(2):313–7.
- Damato B, Coupland SE. A reappraisal of the significance of largest basal diameter of posterior uveal melanoma. *Eye*. 2009;23(12):2152–62.
- Damato B, Dopierala J, Klaasen A, van Dijk M, Sibbring J, Coupland SE. Multiplex ligation-dependent probe amplification of uveal melanoma: correlation with metastatic death. *Invest Ophthalmol Vis Sci*. 2009;50(7):3048–55.
- Ewens KG, Kanetsky PA, Richards-Yutz J, Al-Dahmash S, De Luca MC, Bianciotto CG, et al. Genomic profile of 320 uveal melanoma cases: Chromosome 8p-loss and metastatic outcome. *Invest Ophthalmol Vis Sci*. 2013;54(8):5721–9.
- Shields CL, Furuta M, Thangappan A, Nagori S, Mashayekhi A, Lally DR, et al. Metastasis of uveal melanoma millimeter-by-millimeter in 8033 consecutive eyes. *Arch Ophthalmol*. 2009;127(8):989–98.
- Shields CL, Say EAT, Hasanreisoglu M, Saktanasate J, Lawson BM, Landy JE, et al. Personalized prognosis of uveal melanoma based on cytogenetic profile in 1059 patients over an 8-year period: The 2017 Harry S. Gradle Lecture. *Ophthalmology*. 2017;124(10):1523–31.
- Shields CL, Say EAT, Hasanreisoglu M, Saktanasate J, Lawson BM, Landy JE, et al. Cytogenetic abnormalities in uveal melanoma based on tumor features and size in 1059 patients: The 2016 W. Richard Green Lecture. *Ophthalmology*. 2017;124(5):609–18.
- Onken MD, Worley LA, Ehlers JP, Harbour JW. Gene expression profiling in uveal melanoma reveals two molecular classes and predicts metastatic death. *Can Res*. 2004;64(20):7205–9.
- Onken MD, Worley LA, Tuscan MD, Harbour JW. An accurate, clinically feasible multi-gene expression assay for predicting metastasis in uveal melanoma. *J Mol Diagn*. 2010;12(4):461–8.
- Worley LA, Onken MD, Person E, Robirds D, Branson J, Char DH, et al. Transcriptomic versus chromosomal prognostic markers and clinical outcome in uveal melanoma. *Clin Can Res*. 2007;13(5):1466–71.

22. Decatur CL, Ong E, Garg N, Anbunathan H, Bowcock AM, Field MG, et al. Driver mutations in uveal melanoma: Associations with gene expression profile and patient outcomes. *JAMA Ophthalmol*. 2016;134(7):728–33.
23. Harbour JW, Onken MD, Roberson EDO, Duan S, Cao L, Worley LA, et al. Frequent mutation of BAP1 in metastasizing uveal melanomas. *Science*. 2010;330(6009):1410–3.
24. Kalirai H, Dodson A, Faqir S, Damato BE, Coupland SE. Lack of BAP1 protein expression in uveal melanoma is associated with increased metastatic risk and has utility in routine prognostic testing. *Br J Cancer*. 2014;111(7):1373–80.
25. Koopmans AE, Verdijk RM, Brouwer RW, van den Bosch TP, van den Berg MM, Vaarwater et al. Clinical significance of immunohistochemistry for detection of BAP1 mutations in uveal melanoma. *Mod Pathol*. 2014;27(10):1321–30.
26. Shah AA, David Bourne T, Murali R. BAP1 protein loss by immunohistochemistry: A potentially useful tool for prognostic prediction in patients with uveal melanoma. *Pathology*. 2013;45(7):651–6.
27. van de Nes JA, Nelles J, Kreis S, Metz CH, Hager T, Lohmann DR, et al. Comparing the prognostic value of BAP1 mutation pattern, chromosome 3 status, and BAP1 immunohistochemistry in uveal melanoma. *Am J Surg Pathol*. 2016;40(6):796–805.
28. van Essen TH, van Pelt SI, Versluis M, Bronkhorst IHG, van Duinen SG, Marinkovic M, et al. Prognostic parameters in uveal melanoma and their association with BAP1 expression. *Br J Ophthalmol*. 2014;98(12):1738–43.
29. Yavuzigitoglu S, Koopmans AE, Verdijk RM, Vaarwater J, Eussen B, van Bodegom A, et al. Uveal melanomas with SF3B1 mutations: A distinct subclass associated with late-onset metastases. *Ophthalmology*. 2016;123(5):1118–28.
30. Gupta MP, Lane AM, DeAngelis MM, Mayne K, Crabtree M, Gragoudas ES, et al. Clinical characteristics of uveal melanoma in patients with germline BAP1 mutations. *JAMA Ophthalmol*. 2015;133(8):881–7.
31. Aoude LG, Vajdic CM, Krickler A, Armstrong B, Hayward NK. Prevalence of germline BAP1 mutation in a population-based sample of uveal melanoma cases. *Pigment Cell Melanoma Res*. 2013;26(2):278–9.
32. Njauw C-NJ, Kim I, Piris A, Gabree M, Taylor M, Lane AM, et al. Germline BAP1 inactivation is preferentially associated with metastatic ocular melanoma and cutaneous-ocular melanoma families. *PLoS ONE*. 2012;7(4):e35295.
33. Turunen JA, Markkinen S, Wilska R, Saarinen S, Raivio V, Täll M, Lehesjoki A-E, et al. BAP1 germline mutations in Finnish patients with uveal melanoma. *Ophthalmology*. 2016;123(5):1112–7.
34. Field MG, Durante MA, Anbunathan H, Cai LZ, Decatur CL, Bowcock AM, et al. Punctuated evolution of canonical genomic aberrations in uveal melanoma. *Nature Commun*. 2018;9(1):116.
35. Shields CL, Ganguly A, Materin MA, Teixeira L, Mashayekhi A, Swanson LA, et al. Chromosome 3 analysis of uveal melanoma using fine-needle aspiration biopsy at the time of plaque radiotherapy in 140 consecutive cases: the Deborah Iverson, MD. *Lectureship. Arch Ophthalmol*. 2007;125(8):1017–24.
36. Abdel-Rahman MH, Rai K, Pilarski R, Davidorf FH, Cebulla CM. Germline BAP1 mutations misreported as somatic based on tumor-only testing. *Familial Cancer*. 2016;15(2):327–30.
37. Ewens KG, Kanetsky PA, Richards-Yutz J, Purrazzella J, Shields CL, Ganguly T, et al. Chromosome 3 status combined with BAP1 and EIF1AX mutation profiles are associated with metastasis in uveal melanoma. *Invest Ophthalmol Vis Sci*. 2014;55(8):5160–7.
38. Chen Z, Moran K, Richards-Yutz J, Toorens E, Gerhart D, Ganguly T, et al. Enhanced sensitivity for detection of low-level germline mosaic RB1 mutations in sporadic retinoblastoma cases using deep semiconductor sequencing. *Hum Mut*. 2014;35(3):384–91.
39. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and Guidelines for the Interpretation of Sequence Variants: A Joint Consensus. *Genet Med*. 2015;17(5):405–24.
40. Dono M, Angelini G, Cecconi M, Amaro A, Esposito AI, Mirisola V, et al. Mutation frequencies of GNAQ, GNA11, BAP1, SF3B1, EIF1AX and TERT in uveal melanoma: detection of an activating mutation in the TERT gene promoter in a single case of uveal melanoma. *Br J Cancer*. 2014;110(4):1058–65.
41. Johansson P, Aoude LG, Wadt K, Glasson WJ, Warrier SK, Hewitt AW, et al. Deep sequencing of uveal melanoma identifies a recurrent mutation in PLCB4. *Oncotarget*. 2016;7(4):4624–31.
42. Robertson AG, Shih J, Yau C, Gibb EA, Oba J, Mungall KL, et al. Integrative analysis identifies four molecular and clinical subsets in uveal melanoma. *Cancer Cell*. 2017;32(2):204–20.
43. Staby KM, Grøvdal K, Mørk SJ, Heegaard S, Vintermyr OK, Krohn J. Prognostic impact of chromosomal aberrations and GNAQ, GNA11 and BAP1 mutations in uveal melanoma. *Acta Ophthalmol*. 2018;96(1):31–8.
44. Martin M, Maszhofer L, Temming P, Rahmann S, Metz C, Bornfeld N, et al. Exome sequencing identifies recurrent somatic mutations in EIF1AX and SF3B1 in uveal melanoma with disomy 3. *Nat Genet*. 2013;45(8):933–6.
45. Wadt KA, Aoude LG, Johansson P, Solinas A, Pritchard A, Crainic O, et al. A recurrent germline BAP1 mutation and extension of the BAP1 tumor predisposition spectrum to include basal cell carcinoma. *Clin Genet*. 2015;88(3):267–72.
46. van Engen-van Grunsven ACH, Baar MP, Pfundt R, Rijntjes J, Küsters-Vandeveldt HVN, Delbecq A-L, et al. Whole-genome copy-number analysis identifies new leads for chromosomal aberrations involved in the oncogenesis and metastatic behavior of uveal melanomas. *Melanoma Res*. 2015;25(3):200–9.
47. Popova T, Hebert L, Jacquemin V, Gad S, Caux-Moncoutier V, Dubois-d'Enghien C, et al. Germline BAP1 mutations predispose to renal cell carcinomas. *Am J Hum Genet*. 2013;92(6):974–80.
48. Cheung M, Talarchek J, Schindeler K, Saraiva E, Penney LS, Ludman M, et al. Further evidence for germline BAP1 mutations predisposing to melanoma and malignant mesothelioma. *Cancer Genet*. 2013;206(5):206–10.
49. Laurent C, Gentien D, Piperno-Neumann S, Némati F, Nicolas A, Tesson B, et al. Patient-derived xenografts recapitulate molecular features of human uveal melanomas. *Mol Oncology*. 2013;7(3):625–36.
50. Wiesner T, Obenaus AC, Murali R, Fried I, Griewank KG, Ulz P, et al. Germline mutations in BAP1 predispose to melanocytic tumors. *Nat Genet*. 2011;43(10):1018–21.
51. Pilarski R, Cebulla CM, Massengill JB, Rai K, Rich T, Strong L, et al. Expanding the clinical phenotype of hereditary BAP1 cancer predisposition syndrome, reporting three new cases. *Genes Chromosomes Cancer*. 2014;53(2):177–82.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

