

5-4-2017

Guanylate cyclase C as a target for prevention, detection, and therapy in colorectal cancer.

Allison A. Aka
Thomas Jefferson University

Jeff A. Rappaport
Thomas Jefferson University

Amanda M. Pattison
Thomas Jefferson University

Takami Sato
Thomas Jefferson University

Adam E. Snook
Thomas Jefferson University
Follow this and additional works at: <https://jdc.jefferson.edu/petfp>

 Part of the [Medical Pharmacology Commons](#)
See next page for additional authors

[Let us know how access to this document benefits you](#)

Recommended Citation

Aka, Allison A.; Rappaport, Jeff A.; Pattison, Amanda M.; Sato, Takami; Snook, Adam E.; and Waldman, Scott A., "Guanylate cyclase C as a target for prevention, detection, and therapy in colorectal cancer." (2017). *Department of Pharmacology and Experimental Therapeutics Faculty Papers*. Paper 82.
<https://jdc.jefferson.edu/petfp/82>

This Article is brought to you for free and open access by the Jefferson Digital Commons. The Jefferson Digital Commons is a service of Thomas Jefferson University's [Center for Teaching and Learning \(CTL\)](#). The Commons is a showcase for Jefferson books and journals, peer-reviewed scholarly publications, unique historical collections from the University archives, and teaching tools. The Jefferson Digital Commons allows researchers and interested readers anywhere in the world to learn about and keep up to date with Jefferson scholarship. This article has been accepted for inclusion in Department of Pharmacology and Experimental Therapeutics Faculty Papers by an authorized administrator of the Jefferson Digital Commons. For more information, please contact: JeffersonDigitalCommons@jefferson.edu.

Authors

Allison A. Aka, Jeff A. Rappaport, Amanda M. Pattison, Takami Sato, Adam E. Snook, and Scott A. Waldman

| | |
|----|---|
| 1 | |
| 2 | Review Article |
| 3 | |
| 4 | Guanylate Cyclase C as a Target for Prevention, Detection, and Therapy in Colorectal |
| 5 | Cancer |
| 6 | |
| 7 | Contents |
| 8 | 1.0 Abstract |
| 9 | 2.0 Current challenges in colorectal cancer |
| 10 | 3.0 Genetic basis of colorectal cancer |
| 11 | 4.0 GUCY2C as a target in colorectal cancer |
| 12 | 5.0 GUCY2C agonists for colorectal cancer prevention |
| 13 | 5.1 Endogenous ligands |
| 14 | 5.2 Enterotoxins |
| 15 | 5.3 Synthetic peptides |
| 16 | 6.0 GUCY2C-targeted immunotherapies for metastatic colorectal cancer |
| 17 | 6.1 Vaccines |
| 18 | 6.2 Adoptive T-cell therapies |
| 19 | 6.3 GUCY2C-targeted immunotoxins |
| 20 | 7.0 GUCY2C as a biomarker for colorectal cancer detection |
| 21 | 7.1 GUCY2C mRNA as a biomarker |
| 22 | 7.2 GUCY2C as a target for diagnostic imaging agents |
| 23 | 8.0 Conclusion |
| 24 | 9.0 Expert Commentary |
| 25 | 10.0 Five Year View |
| 26 | 11.0 Key Issues |
| 27 | |
| 28 | |
| 29 | |

30 **1.0 Abstract**

31 **Introduction:** Colorectal cancer remains the second leading cause of cancer death in the
32 United States, and new strategies to prevent, detect, and treat the disease are needed. The
33 receptor, guanylate cyclase C (GUCY2C), a tumor suppressor expressed by the intestinal
34 epithelium, has emerged as a promising target.

35 **Areas Covered:** This review outlines the role of GUCY2C in tumorigenesis, and steps to
36 translate GUCY2C-targeting schemes to the clinic. Endogenous GUCY2C-activating ligands
37 disappear early in tumorigenesis, silencing its signaling axis and enabling transformation.
38 Pre-clinical models support GUCY2C ligand supplementation as a novel disease prevention
39 paradigm. With the recent FDA approval of the GUCY2C ligand, *linaclotide*, and two more
40 synthetic ligands in the pipeline, this strategy can be tested in human trials. In addition to
41 primary tumor prevention, we also review immunotherapies targeting GUCY2C expressed
42 by metastatic lesions, and platforms using GUCY2C as a biomarker for detection and patient
43 staging.

44 **Expert Commentary:** Results of the first GUCY2C targeting schemes in patients will become
45 available in the coming years. The identification of GUCY2C ligand loss as a requirement for
46 colorectal tumorigenesis has the potential to change the treatment paradigm from an
47 irreversible disease of genetic mutation, to a treatable disease of ligand insufficiency.

48

49 **2.0 Current Challenges in Colorectal Cancer**

50 Colorectal cancer (CRC) remains the fourth most diagnosed cancer, and the second leading
51 cause of cancer death in the United States [1]. Worldwide, it accounts for as many as 1.2
52 million new cases and 600,000 deaths per year [2]. CRC incidence and mortality has
53 declined since the 1980s, paralleling adoption of screening; however, available screening
54 methods (e.g. fecal occult blood, flexible sigmoidoscopy, colonoscopy) vary in terms of
55 sensitivity and specificity, risks, and evidence supporting their implementation.
56 Unfortunately, no screening method has proven to reduce all-cause mortality [3].
57 Colonoscopy has become the gold standard, enabling removal of dysplastic lesions before
58 progression to cancer. Yet, sensitivity decreases for lesions <1 cm, and the adenoma
59 detection rate and completeness of polyp removal varies between providers [4, 5]. Indeed,

60 the superiority of colonoscopy over other screening approaches recently was questioned
61 [6].

62
63 Complicating limitations of current screening tools, widespread and ill-defined risk factors
64 for CRC make it difficult to develop screening guidelines. A fraction of new tumors arise in
65 patients with known genetic syndromes (e.g. Lynch syndrome, familial adenomatous
66 polyposis); however >90% of cases are thought to be sporadic [7]. Age and family history
67 play a role, hence colonoscopy is indicated at age 50 for patients with average risk, and at
68 age 40 for patients with a first degree relative diagnosed at a young age [2]. But other risk
69 factors of unclear significance include high-fat diets, tobacco smoking, alcohol
70 consumption, and body mass index [8, 9]. Patients with inflammatory conditions of the
71 bowel, such as ulcerative colitis, are particularly predisposed to CRC [10], and several
72 studies have suggested benefits from low dose non-steroidal anti-inflammatory drugs [11,
73 12, 13]. However, a pathophysiological link between these risk factors and tumorigenesis
74 remains unclear, delaying the development of disease prevention schemes.

75
76 Despite progress in early detection and treatment, ~25% of patients present with late
77 stage disease [14]. Many promising agents for metastatic disease have **become clinically**
78 **available** (e.g. tyrosine kinase inhibitors, epidermal growth factor inhibitors, anti-
79 angiogenesis agents, etc.), but the five-year survival rate for this population remains only
80 11.7% [2, 14]. Hence, strategies for prevention, detection, and treatment of primary and
81 metastatic CRC are needed.

82 83 **3.0 Genetic Basis of Colorectal Cancer**

84 CRCs develop slowly, often requiring over a decade to accumulate mutations required for
85 epithelial transformation (providing a long window for detection). **The average tumor**
86 **contains 90 different mutations [15], but despite this genetic heterogeneity, 70-80% of**
87 **sporadic (non-hereditary) colorectal tumors arise by a series of mutations typically**
88 **described as the adenoma-carcinoma sequence [7, 16].** Canonically, this sequence begins
89 with an inactivating mutation of the *adenomatous polyposis coli* (*APC*) tumor suppressor
90 gene. The APC protein normally inhibits the accumulation of β -catenin, the downstream

91 mediator of the Wnt signaling pathway. APC loss enables phosphorylation and aberrant
92 translocation of β -catenin to the nucleus in the absence of the Wnt ligand, where it
93 participates in oncogenic transcription (activation of c-MYC and cyclin D1) [17].
94 Subsequent activating mutations of oncogenes (e.g. *KRAS*), and deactivating mutations of
95 tumor suppressors (e.g. *TP53*) characterize the progression from normal epithelium, to
96 adenoma, to carcinoma [7, 16]. About 15% of sporadic tumors arise by a different
97 mechanism, characterized by dysfunction of DNA mismatch repair genes, such as *MLH1* and
98 *MSH2*. Here, defective DNA repair permits accumulation of mutations in short repeated
99 genetic sequences (microsatellite sequences), resulting in a microsatellite instability
100 phenotype. Finally, a third tumorigenic pathway is characterized by aberrant gene silencing
101 via CpG island methylation, an epigenetic phenomenon. These adenomas often harbor
102 mutational activation of the *BRAF* oncogene, and exhibit a characteristic sessile serrated
103 architecture [7, 16]. Many tumors contain elements of more than one pathway; for
104 example, hypermethylation of the *MLH1* mismatch repair gene contributes to a large subset
105 of microsatellite unstable tumors.

106
107 By comparison, tumors arising from hereditary CRC syndromes occur less frequently (3-
108 5% of cases), and arise from germline, rather than somatic, mutations in the
109 aforementioned pathways [7, 16]. Hereditary non-polyposis CRC (HNPCC, or Lynch
110 syndrome) occurs most commonly, and arises from mismatch repair gene mutation.
111 Familial adenomatous polyposis (FAP) arises from germline mutations of *APC*, resulting in
112 thousands of colonic polyps early in life and 100% risk of cancer by age 40.

113 114 **4.0 GUCY2C as a Target in Colorectal Cancer**

115 Although a wealth of genetic and epigenetic changes have been associated with intestinal
116 transformation, a common causative agent has yet to be found. Recent studies have defined
117 a role for the intestinal surface receptor, guanylate cyclase C (*GUCY2C*), as a tumor
118 suppressor involved in the earliest stages of transformation.

119
120 *GUCY2C* belongs to the particulate guanylate cyclase class of receptors, and appears on the
121 apical brush border of the intestinal epithelium [18]. Early studies defined its role in

122 regulating luminal secretion, specifically as the receptor for the bacterial heat-stable
123 enterotoxin, ST, the causative agent of traveler's diarrhea [19]. ST binding to the
124 extracellular domain of GUCY2C activates the intracellular catalytic domain, converting
125 GTP to cyclic GMP [18]. This second messenger activates cGMP-dependent protein kinase II
126 (PKGII), leading to downstream phosphorylation and activation events, including water
127 and electrolyte secretion via the cystic fibrosis transmembrane conductance regulator
128 (CFTR) [18]. Predictably, persons with activating or deactivating mutations of *GUCY2C*
129 exhibit intestinal hyper- or hypo- secretory syndromes, respectively [20, 21]. Our
130 understanding of GUCY2C-induced signaling has since expanded to include regulatory roles
131 in epithelial renewal along the crypt-villus axis [22, 23], GI barrier integrity [24, 25], injury
132 response [26, 27], and the gut-brain satiety axis [28, 29]. Importantly, GUCY2C is densely
133 expressed throughout the intestine, and overexpressed by tumor tissue, features that can
134 be exploited for diagnostic and therapeutic goals [30, 31, 32].

135

136 Endogenous GUCY2C ligands, the **peptides** guanylin and uroguanylin, are among the most
137 commonly lost gene products in mouse models and human CRC [33, 34, 35]. In a study of
138 300 patients, >85% of colorectal tumors exhibited disappearance of guanylin mRNA and
139 protein compared to normal adjacent tissue [33]. This loss occurs early in intestinal
140 transformation, suggesting that an intact GUCY2C signaling axis opposes tumorigenesis [23,
141 36, 37]. Indeed, mice in which GUCY2C expression is eliminated (*Gucy2c^{-/-}*) exhibit a
142 tumorigenic phenotype, including epithelial dysfunction, DNA mutation, cellular
143 proliferation and migration, and metabolic reprogramming (Figure 1) [24, 37].
144 Interestingly, diet-induced obesity in mice also leads to guanylin loss and tumor formation,
145 suggesting a mechanistic link between CRC and a well-described risk factor, obesity [28].
146 Conversely, GUCY2C activating ligands and downstream mediators suppress oncogenic
147 drivers (e.g. pRb, cyclin D1, B-catenin, pAKT) and increase tumor suppressors (e.g. p21,
148 p27) [23, 25, 38, 39]. These findings underlie the *paracrine hormone hypothesis*, whereby
149 CRC arises from an environment of **ligand** loss and functional GUCY2C inactivation. This
150 pathophysiological paradigm could transform colon cancer from an irreversible disease of
151 genetic origin, to a treatable disorder of **ligand** insufficiency. Recent FDA-approval of the

152 GUCY2C ligand, *linaclotide*, and the entrance of two others in the clinical pipeline, makes it
153 feasible to test **ligand** supplementation for chemoprevention of CRC in humans.

154

155 **5.0 GUCY2C Agonists for Colorectal Cancer Prevention**

156 GUCY2C peptide agonists available for chemoprevention of primary colorectal tumors
157 include the endogenous **peptides** guanylin and uroguanylin, bacterial diarrheagenic heat-
158 stable enterotoxins (STs), and the synthetic peptides linaclotide, plecanatide, and
159 dolcanatide. These ligands share structural homologies and conserved mechanisms of
160 action through GUCY2C activation and downstream cGMP production.

161

162 *5.1 Endogenous Ligands*

163 Guanylin and uroguanylin, the endogenous ligands for GUCY2C in the intestine, were first
164 described in the early 1990s [40, 41]. They are produced and stored as propeptides, and
165 undergo processing to their mature 15-mer (guanylin) or 16-mer (uroguanylin) forms. The
166 mature **peptides** are thought to act on GUCY2C in a paracrine fashion to maintain epithelial
167 homeostasis and fluid secretion [18]. Uroguanylin also serves an endocrine role in gut-
168 brain satiety signaling [28]. The **peptides** exhibit complimentary roles along the axis of the
169 intestine, with maximal uroguanylin expression in the small intestine, and guanylin in the
170 large intestine [42]. However, even after two decades of study, the cells of origin remain
171 controversial, and may continue to evolve as we elucidate the multiple roles of the GUCY2C
172 signaling axis [43, 44, 45, 46]. Interestingly, despite >50% sequence homology, uroguanylin
173 is principally active in acidic pH and guanylin in basic pH, further reflecting regional
174 specificity [47].

175

176 The disappearance of guanylin/uroguanylin early in colorectal tumorigenesis reflects a
177 tumor suppressive function. Preclinical studies in mice have demonstrated the potential of
178 therapeutic **ligand** replacement. For example, in *Apc^{min/+}* mice (a CRC model) oral
179 uroguanylin supplementation inhibited tumorigenesis [48]. In another model, mice
180 genetically modified to overexpress guanylin were resistant to DSS-induced colitis [25].
181 Further, it was recently shown that diet-induced obesity suppressed guanylin expression in
182 mice, leading to tumorigenesis, and specific enforcement of guanylin expression prevented

183 obesity-related tumors [49]. In all of these studies, no adverse effects were observed over
184 the lifetime of the mice.

185

186 5.2 Enterotoxins

187 Heat-stable enterotoxins (STs) are produced by several diarrheagenic bacteria, including
188 enterotoxigenic *E. coli*, *K. pneumonia*, *V. cholera*, and *Y. enterocolitica* [18]. First described
189 as GUCY2C agonists in 1990, STs include a family of peptides with a conserved C-terminal
190 region [19]. Structurally similar to guanylin and uroguanylin, STs contain an additional
191 disulfide bond, contributing to their canonical heat stability and increased receptor binding
192 affinity [18]. Ligand-receptor binding activates GUCY2C, leading to CFTR-driven fluid and
193 electrolyte transport into the intestinal lumen, manifesting as secretory diarrhea.
194 Enterotoxigenic *E. coli* is endemic in developing countries with poor sanitation
195 infrastructure. Interestingly, these regions have a lower incidence of CRC, which may
196 reflect life-long exposure to STs, increased GUCY2C activation, and suppression of epithelial
197 dysplasia [39, 50].

198

199 5.3 Synthetic Peptides

200 Synthetic peptides sharing homology with natural GUCY2C ligands target the secretory
201 function of GUCY2C for therapeutic purposes. The first agent developed, *linaclotide*
202 (Ironwood Pharmaceuticals, Inc., Cambridge, MA), is an ST analog approved by the FDA for
203 the treatment of chronic idiopathic constipation (CIC) and constipation-predominant
204 irritable bowel syndrome (IBS-C). *Linaclotide* binds GUCY2C, inducing cGMP accumulation
205 and fluid secretion. Double-blind, placebo-controlled, phase III clinical trials were
206 completed for patients with IBS-C (MCP-103-302 and LIN-MD-31) and CIC (MCP-103-303
207 and LIN-MD-01) [51, 52, 53]. *Linaclotide* met all primary endpoints, significantly reducing
208 abdominal symptoms and severity of constipation. No differences in serious adverse events
209 were observed between *linaclotide* and placebo. The most commonly reported side effect
210 was diarrhea, an effect predicted by its mechanism of action. New agents, *plecanatide* and
211 *dolcanatide* (Synergy Pharmaceuticals Inc., New York, NY), are uroguanylin analogs with
212 increased potency [54]. Like *linaclotide*, these agents agonize GUCY2C and stimulate cGMP
213 production. They reduced disease severity (e.g. weight loss, inflammatory infiltrate,

214 destruction of crypt architecture) in pharmacologic and genetic murine models of colitis
215 [54]. In a phase I trial of 72 healthy volunteers, up to 48.6 mg of *plecanatide* was safe and
216 well-tolerated [55]. Currently, *plecanatide* is in phase III clinical trials for CIC and IBS-C
217 [56].

218

219 Given their safety in human trials, these compounds could be used as oral-
220 chemopreventive agents for CRC. In principle, exogenous GUCY2C ligand administration
221 would reconstitute the tumor-suppressing GUCY2C signaling axis, preventing colorectal
222 tumorigenesis. A phase I trial is underway to identify oral *linaclotide* dosing regimens that
223 stimulate GUCY2C in the rectum. Study participants receive a single oral dose of *linaclotide*
224 daily for 7 days, and then are assessed for increases in cGMP levels in rectal biopsy, as well
225 as safety and tolerability (Linaclotide Acetate in Preventing Colorectal Cancer in Healthy
226 Volunteers, clinicaltrials.gov NCT01950403).

227

228 **6.0 GUCY2C-Targeted Immunotherapies for Metastatic Colorectal Cancer**

229 While prevention of CRC remains the clinical ideal, therapeutic strategies for advanced
230 disease are also needed. A growing body of literature endorses immunotherapy for cancer
231 treatment. The immune system has a remarkable ability to suppress neoplastic
232 proliferation, as demonstrated by heightened cancer risk in immunocompromised patients
233 [57, 58]. In part, this risk reflects diminished immune control of oncogenic viruses (e.g.
234 human herpes virus 8 and Kaposi sarcoma, hepatitis B and C viruses and liver cancer, or
235 Epstein-Barr virus and Hodgkin's lymphoma) [58]; **however**, these patients also are
236 predisposed to cancers without known infectious etiologies (e.g. melanoma, thyroid, and
237 colorectal cancers) [58]. Instead, these are thought to arise from poor immune surveillance
238 against cancer cells in tumors and the circulation. For example, the presence of
239 lymphocytes in CRC tumors is associated with delayed metastasis and prolonged survival
240 [59]. Tumor cells have a propensity to bypass or overcome these natural defense
241 mechanisms, creating an unmet need for therapies that improve the immune response to
242 cancer antigens (e.g. vaccines, adoptive T cell therapy) or target cancer cells directly (e.g.
243 immunotoxins) [60, 61].

244

245 Effective CRC immunotherapies require antigenic targets that maximize immunogenicity
246 and minimize autoimmunity. The most explored target, the glycoprotein carcinoembryonic
247 antigen (CEA), is upregulated in CRC, but also appears in organs outside the GI tract,
248 leading to **potential** autoimmunity and immunological tolerance [62, 63]. In contrast,
249 GUCY2C has unique anatomic and biological characteristics **that appear to circumvent**
250 **these issues**. GUCY2C is expressed by intestinal mucosa from the small bowel to the rectum,
251 and is overexpressed in primary and metastatic colorectal neoplasms [30, 31, 32]. **Further,**
252 **expression is largely restricted to the luminal aspect of the GI mucosa, and its extracellular**
253 **domain is antigenically distinct from other members of the guanylate cyclase family found**
254 **in other tissues [64, 65, 66]. Importantly, GUCY2C resides in an immune privileged**
255 **compartment, with minimal exposure to the systemic immune response [64, 65, 66].**
256 **Limited cross-talk between systemic and mucosal immune elements protects normal**
257 **mucosa expressing GUCY2C from autoimmune toxicity, while also limiting systemic**
258 **tolerance to the antigen [64, 65, 66].** These **advantages** have led to the exploration of
259 several GUCY2C-targeted immunotherapeutic strategies (Figure 1).

260

261 *6.1 Vaccines*

262 Similar to the yearly-recommended flu vaccine, cancer vaccines stimulate the immune
263 system to destroy cancer cells by targeting tumor-specific antigens, while also generating
264 long-lasting immunity [60]. Viral vector vaccines, engineered to contain the genes for
265 cancer antigens, enhance antitumor immunity by stimulating the expansion of adaptive
266 immune system elements, namely Type 1 CD4⁺ T-helper cells, cytotoxic CD8⁺ T cells and
267 antibodies [61]. This paradigm forms the basis for a GUCY2C-targeted vaccine, designed to
268 **elicit** immune responses to metastatic CRC.

269

270 The first GUCY2C-specific vaccine incorporated replication-deficient type 5 recombinant
271 adenovirus (Ad5) encoding the extracellular domain of GUCY2C (Ad5-GUCY2C) [64, 65, 66].
272 In a murine pre-clinical proof-of-concept study, the vaccine stimulated a GUCY2C-specific
273 CD8⁺ cytotoxic T-cell response, which killed GUCY2C-expressing colon cancer cells.
274 Remarkably, survival in mice with lung and liver metastases improved, **without signs of**
275 **inflammatory bowel disease, organ or metabolic dysfunction, or autoimmune tissue**

276 **damage** [64]. Interestingly, the vaccine produced strong CD4⁺ T-cell, CD8⁺ T-cell, and B-cell
277 responses in *Gucy2c*^{-/-} mice, but produced only a modest CD8⁺ T-cell response in *Gucy2c*^{+/+}
278 mice, which was attributed to **GUCY2C-specific** CD4⁺ T-cell tolerance [66]. **To overcome**
279 **this, the vaccine was modified to include an immunogenic T-helper epitope from foreign**
280 **protein** [66, 67]. This new vector reconstituted CD4⁺ T-cell, CD8⁺ T-cell, and memory
281 responses [66]. This was the first demonstration that selective CD4⁺ T-cell tolerance blocks
282 GUCY2C-specific immunity and memory responses. Importantly, this paradigm may extend
283 to other antigens, including those in melanoma and breast cancer, suggesting that
284 overcoming CD4⁺ T-cell tolerance may be a requirement in many cancer vaccine
285 approaches [66, 68, 69].

286

287 **Preliminary results were recently reported for a phase I clinical trial exploring the safety**
288 **and immunogenicity of this vaccination scheme in stage I/II colon cancer patients**
289 **([clinicaltrials.gov NCT01972737](https://clinicaltrials.gov/ct2/show/study/NCT01972737))**[70]. The vaccine is analogous to the murine vaccine, but
290 encodes the human GUCY2C extracellular domain fused to the **T-helper epitope PAn DR**
291 **Epitope** (Ad5-GUCY2C-PADRE). Preliminary findings are consistent with the pre-clinical
292 studies, with patients responding to the vaccine by producing GUCY2C-specific CD8⁺ T-cell
293 and B-cell responses, but not a CD4⁺ T-cell response, suggesting that selective CD4⁺ T-cell
294 tolerance governs GUCY2C-specific immune responses in humans, as well as mice [70].
295 **Moreover, like preclinical studies, the vaccine did not induce GUCY2C-targeted toxicity in**
296 **any GUCY2C-expressing tissue**. Importantly, these first findings in humans support Ad5-
297 GUCY2C-PADRE as a promising therapeutic approach for patients with GUCY2C-expressing
298 malignancies.

299

300 *6.2 Adoptive T Cell Therapies*

301 The past decade has witnessed remarkable progress in an immunotherapy approach
302 known as adoptive cell therapy (ACT). Rather than employing a vaccine or other drug to
303 induce an immune response within a patient, **this strategy employs ex vivo tissue culture to**
304 **expand naturally-occurring immune effectors or create them *de novo* for administration to**
305 **the patient** [71]. One approach involves boosting the activity of naturally occurring
306 immune responses present in tumors, called tumor-infiltrating lymphocytes (TILs), which

307 are suppressed by the tumor microenvironment [61]. TILs can be isolated from patient
308 tumors, activated and expanded *ex vivo*, and reintroduced to the patient, bypassing
309 immunosuppressive elements. Another approach involves *ex vivo* genetic manipulation of
310 peripheral blood lymphocytes to retarget them to tumors by expressing cancer-specific T-
311 cell receptors (TCRs). Both TIL and TCR-gene transfer approaches have been efficacious in
312 mouse models and humans with metastatic melanoma [72, 73, 74, 75, 76]. An ACT
313 alternative approach employs chimeric antigen receptors (CARs). Here, T lymphocytes are
314 modified to express an engineered receptor comprised of intracellular T-cell signaling
315 motifs and an extracellular antibody domain that recognizes antigens in an MHC/HLA-
316 independent fashion [77, 78]. CD19-targeted CAR-T cells have shown remarkable promise
317 in the treatment of refractory leukemia in humans [79, 80, 81]. Because CARs can
318 theoretically employ antibodies targeting any **cell surface** antigen, ACT approaches may be
319 vastly expanded and personalized for other malignancies, including solid tumors.

320

321 While efficacious for certain cancers, ACT has **had mixed results** in CRC patients. **A recent**
322 **report demonstrated regression of lung metastases in a patient with colorectal cancer**
323 **injected with TILs targeting mutant KRAS [82]. However, prior** trials of ACT targeting CEA
324 and Her-2 resulted in adverse autoimmune effects, including death [83, 84]. In contrast,
325 GUC2YC-targeted CAR-T cells may target metastatic CRC cells without destroying healthy
326 tissue, given the anatomical compartmentalization of GUCY2C on the luminal aspect of the
327 intestine, beyond access by CAR-T cells, which recognize native GUCY2C. As a proof-of-
328 concept, CD8⁺ T cells bearing CARs targeted to mouse GUCY2C lysed murine colon cancer
329 cells, **eliminated colorectal cancer metastases**, and prolonged survival in a mouse model of
330 metastatic CRC, without toxicity [85].

331

332 *6.3 GUCY2C-targeted Immunotoxins*

333 Antibodies offer several advantages as an immunotherapeutic tool, including
334 immunomodulatory capacity, interference in ligand-receptor interactions, and relative ease
335 of mass-production. Indeed, antibody-therapies are well-established in the clinic, with over
336 50 FDA-approved therapeutics [86]. For example, the monoclonal antibody bevacizumab
337 (Avastin), which targets the vascular endothelial growth factor pathway, is FDA-approved

338 as first line treatment for metastatic CRC. Others include cetuximab (Erbix) and
339 panitumumab (Vectibix), antibodies which bind to the extracellular domain of the
340 epidermal growth factor receptor, blocking ligand binding and tumorigenic signaling [87,
341 88]. Still, these agents offer limited improvements in survival: bevacizumab was approved
342 as a first line agent for metastatic CRC in 2004, but only increased median survival from 15
343 to 20 months [87].

344 The next generation of antibody therapies, antibody-drug conjugates (ADCs) enable
345 targeted delivery of cytotoxic agents to specific tissues [89, 90]. ADCs are engineered by
346 linking a cytotoxin to a monoclonal antibody, facilitating targeting to cells expressing
347 cancer antigens, endocytic uptake, and intracellular delivery of the toxic payload.
348 Conceptually, the targeted nature of ADCs reduces systemic exposure, and endocytic
349 uptake reduces drug resistance by P-glycoprotein efflux pump, two of the pitfalls of existing
350 chemotherapeutics [89]. However, as a relatively new drug class, ADCs historically have
351 proven difficult to optimize, and have been associated with significant side effects due to
352 non-specific targeting [90]. For this reason, only two have achieved FDA approval,
353 adotrastuzumab emtansine and brentuximab vedotin, although several others have entered
354 clinical trials.

355
356 Recently, a model GUCY2C-targeted ADC was devised, consisting of a GUCY2C antibody,
357 ricin toxin payload, and cleavable disulfide linker (4-succinimidylloxycarbonyl- α -methyl- α -
358 [2-pyridyldithio]- toluene; SMPT) [91]. The ADC specifically targeted GUCY2C, underwent
359 endocytosis, trafficked to lysosomes, and delivered a toxic payload to colon cancer cells
360 [91]. In mice with CRC lung metastases, the ADC prolonged survival without compromising
361 normal tissue [91]. A subsequent phase I clinical trial was recently completed, examining a
362 human IgG1 monoclonal antibody to GUCY2C conjugated via a protease-cleavable linker to
363 monomethyl auristatin E, an anti-microtubule agent. The ADC (TAK-264) was tested for
364 safety and tolerability in 41 patients with GUCY2C-expressing metastatic gastrointestinal
365 disease. Four patients in the highest dose group experienced dose-limiting toxicity
366 (neutropenia), but the safety profile was deemed manageable, and preliminary data
367 suggest antitumor activity [92].

368 **7.0 GUCY2C as a Biomarker in Colorectal Cancer Detection**

369 Features that elevate GUCY2C as a target for immunotherapy (overexpression by tumors,
370 limited expression outside the gastrointestinal tract [30, 31, 32]) also have value for cancer
371 detection and staging. Disease stage remains a key prognostic and therapeutic factor in the
372 management of patients with CRC [93]. Whereas the resection of tumors restricted to the
373 bowel wall (stage II) is often curative, patients with metastasis of tumor cells to lymph
374 nodes (stage III) experience recurrence rates of up to 50% with surgery alone [2]. Although
375 adjuvant chemotherapy remains controversial at stage II, progression to stage III is an
376 indication for chemotherapy, increasing survival as much as 15% [2, 94]. Unfortunately,
377 traditional staging by histopathological examination of lymph node tissue remains
378 insensitive, leading to missed metastases, patient under-staging, and inappropriate patient
379 management. For example, less than 0.01% of available tissue is typically reviewed, and as
380 many as 25% of supposedly lymph node-negative (pN0) patients die of disease recurrence
381 [93], suggesting undetected metastatic cells.

382

383 *7.1 GUCY2C mRNA as a biomarker*

384 The expression profile of GUCY2C makes it uniquely suited for the staging of primary
385 colorectal tumors and occult metastases [32, 95]. In a blinded multicenter prospective trial,
386 2570 lymph nodes from 257 pN0 colorectal cancer patients were examined for GUCY2C
387 mRNA by quantitative real-time PCR [96]. Patients were followed for 24 months, and the
388 primary outcome measure was time to recurrence. Remarkably, 87% of patients
389 considered stage II by traditional histopathological techniques were found to harbor occult
390 metastases by GUCY2C molecular staging, correlating with earlier time to recurrence.
391 Furthermore, qRT-PCR was used to stratify patients by tumor burden, based on the
392 number of positive nodes and relative GUCY2C expression across nodes [97]. For the first
393 time, it was shown that patients with greater occult tumor burden had a greater risk of
394 recurrence, and this method could be used to stratify patients based on prognostic risk.
395 Importantly, molecular staging by GUCY2C RT-PCR has been validated across multiple
396 users and laboratories and may replace conventional histopathologic evaluation for staging
397 and therapeutic decision making in colorectal cancer [98, 99, 100].

398

399 7.2 GUCY2C as a target for diagnostic imaging agents

400 Positron emission tomography has become a mainstay for staging CRC and monitoring
401 treatment response [101]. This method capitalizes on the increased metabolic demand, and
402 therefore increased glycolysis, by cancer cells. Cancer cells take up the glucose analog, 2-
403 [¹⁸F]fluoro-2-deoxy-D-glucose (FDG) to a greater extent than surrounding normal tissue,
404 allowing visualization by PET. However, glucose requirements by other tissues decreases
405 specificity; false positives (due to inflammation, surgery, diverticulitis, etc.) lead to
406 unnecessary follow-up colonoscopy or inappropriate staging [101]. Alternative imaging
407 modalities using molecular targets, rather than metabolic patterns, may address these
408 issues [102]. Targeting imaging probes to GUCY2C offers a sensitive means of detecting
409 tumors derived from intestinal epithelium. Conjugates of radionuclides and GUCY2C
410 ligands (e.g. ST, uroguanylin analogs) specifically target GUCY2C-expressing xenografts
411 [103, 104]. These agents can be visualized with gamma camera scintigraphy, and
412 accurately differentiate tumors of gastrointestinal origin from surrounding tissue [103,
413 104]. Further, GUCY2C-directed antibodies accumulate in cells via clathrin-mediated
414 endocytosis of the antibody-receptor complex, with the potential to amplify delivery of
415 imaging agents or therapeutic cargo [91].

416

417 **8.0 Conclusion**

418 Despite improvements in CRC screening, incidence and mortality are among the highest of
419 all cancers, and while the genetic basis has been well described, therapeutic targets remain
420 elusive. The intestinal receptor GUCY2C has emerged as a target uniquely suited for
421 prevention, therapy, and diagnostics. Its role as a tumor suppressor, inactivated by **ligand**
422 loss early in tumorigenesis, suggests a novel disease prevention paradigm focused on
423 GUCY2C ligand replacement. A clinical program is underway ultimately to test this strategy
424 with the FDA-approved agent, *linaclotide*, and other promising agents are emerging.
425 Further, its expression profile in the intestinal lumen and metastatic CRC tumors offers an
426 ideal target for a rapidly expanding array of cancer immunotherapies, including vaccines,
427 T-cell therapies, and antibody-drug conjugates. Finally, GUCY2C can be exploited as a
428 sensitive biomarker for the detection and staging of CRC. Translation to the clinics is

429 underway on multiple fronts. Novel approaches targeting GUCY2C could revolutionize the
430 treatment of CRC.

431

432 9.0 Expert Commentary

433

434 Cancer research remains an ever-changing field, with exciting advances in the past few
435 decades that have shifted traditional treatment approaches. Preventative strategies are the
436 clinical ideal and successes have been achieved for several neoplasms, such as the
437 decreased incidence of gastric cancer following the identification and reduction of *H. pylori*
438 infections [1]. Likewise, colonoscopy has reduced the incidence of colorectal cancers by
439 eliminating lesions before they become invasive and metastatic.

440

441 While screening has reduced the incidence of colorectal cancer, it remains the fourth most
442 diagnosed cancer, and the second leading cause of cancer death, with a 5-year survival
443 <15% in metastatic disease. GUCY2C appears to play a pivotal role in epithelial
444 homeostasis, including intestinal barrier integrity and obesity, known risk factors for colon
445 cancer, suggesting novel molecular pathways that may be pharmacologically targetable.
446 The revelation of GUCY2C ligand loss and receptor silencing early in tumorigenesis may
447 have a transformative impact, supported by the exploration of multiple translational
448 avenues. With regards to cancer prevention, reactivation of the GUCY2C tumor suppressor
449 pathway with exogenous peptides has shown promise in pre-clinical models. Though
450 initially formulated for the treatment of irritable bowel disease and chronic constipation,
451 the translation of the FDA-approved GUCY2C ligand, *linaclotide*, to CRC is feasible, as safety
452 and efficacy are already established. However, long-term effects of *linaclotide* and other
453 synthetic GUCY2C ligands have not yet been defined and longitudinal chemoprevention
454 trials are required.

455

456 In the context of CRC treatment, the identification of GUCY2C as a biomarker and cell-
457 surface target of metastatic CRC cells may usher in new biologics and immunotherapies.
458 GUCY2C-targeted vaccines and antibody-drug conjugates have advanced into clinical
459 testing. Further, detection of GUCY2C mRNA in lymph nodes offers a sensitive means of

460 staging the disease, enabling more accurate identification of patients at risk for disease
461 recurrence. Appropriate intervention in patients with previously unrecognized occult
462 metastases may improve survival, especially as targeted therapeutics enter the clinic.

463
464 Another area of interest and debate is the nature of cancer inception, and implications for
465 targeting strategies. Traditionally, disease recurrence and treatment failure are thought to
466 result from the inevitable acquisition of mutations and epigenetic changes that allow
467 cancer cells to evade destruction [105]. However, evidence increasingly indicates the
468 presence of “cancer stem cells”, a subpopulation of cancer cells with stem-like
469 characteristics (e.g., tumorigenesis, self-renewal, and differentiation) that underlie
470 metastasis, recurrence, and chemoresistance [106]. Identification and targeting of cancer
471 stem cell markers could enhance CRC therapies. For example, a recent study demonstrated
472 co-expression of CD133 and the breast cancer resistance protein (BCRP)/ATP-binding
473 cassette subfamily G member 2 (ABCG2) by human colorectal tumors [107].
474 Downregulation of ABCG2 inhibited self-renewal capabilities and enhanced
475 chemotherapeutic effects in double-positive colon adenocarcinoma cells. Dual-therapies,
476 potentially targeting a universal CRC marker like GUCY2C as well as a marker of the stem
477 cell subpopulation may be a new translational avenue. As we better-characterize these
478 neoplastic markers, therapeutic strategies will continue to evolve.

479

480 **10.0 Five Year View**

481 A large body of work across multiple laboratories supports the hypothesis that GUCY2C
482 ligand loss is a necessary step in tumorigenesis. In the next five years, the molecular steps
483 in this process likely will be defined, potentially leading to new clinical targets.
484 Furthermore, results of the first trials translating GUCY2C-targeting schemes to the clinic
485 will become available, including the effectiveness of GUCY2C ligand supplementation with
486 *linaclotide*, a GUCY2C-targeted vaccine, a GUCY2C-targeted antibody-drug conjugate, and
487 GUCY2C-targeted CAR-T cells. Additional GUCY2C ligands (*dolcanatide* and *plecanatide*)
488 entering the pipeline will likely be explored for similar use as chemoprevention agents.
489 Ultimately, the next five years should provide the first insights into the potential for
490 GUCY2C-targeting to influence human colorectal cancer outcomes.

491
492
493
494
495
496
497
498
499
500
501
502
503
504
505
506

11.0 Key Issues

- The gastrointestinal epithelial receptor, guanylate cyclase C (GUCY2C) has been described as a novel tumor suppressor and reliable biomarker of colorectal cancer.
- Endogenous GUCY2C ligand loss has been widely-described as an early step in colorectal tumorigenesis, suggesting a therapeutic strategy of **ligand** replacement for chemoprevention. The GUCY2C agonist *linaclotide* is FDA approved for other indications and a phase I clinical trial examining its use for colorectal cancer prevention is underway.
- GUCY2C is overexpressed in colorectal cancer metastases and several immunotherapies targeting GUCY2C are being explored, including adoptive T-cell therapy with GUCY2C-targeted CAR-T cells, a viral vector vaccine, and a GUCY2C-targeted antibody-drug conjugate. The latter two are currently in early human trials.
- Cancer staging and imaging strategies targeting GUCY2C also are being explored. GUCY2C mRNA is a sensitive biomarker of occult lymph node metastases, improving cancer detection and staging.

507 **References**

508 **Papers of interest (*)**

509 **Papers of considerable interest (**)**

510

511 1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. *CA Cancer J Clin.* 2016; 66:7-30.

512 2. Brenner H, Kloor M, Pox CP. Colorectal cancer. *Lancet.* 2014; 383:1490-502.

513 3. Lin JS, Piper MA, Perdue LA, Rutter CM, Webber EM, O'Connor E, Smith N, Whitlock

514 EP. Screening for Colorectal Cancer: Updated Evidence Report and Systematic

515 Review for the US Preventive Services Task Force. *JAMA.* 2016; 315:2576-94.

516 4. Corley DA, Jensen CD, Marks AR, Zhao WK, Lee JK, Doubeni CA, Zauber AG, de Boer J,

517 Fireman BH, Schottinger JE, Quinn VP, Ghai NR, Levin TR, Quesenberry CP. Adenoma

518 detection rate and risk of colorectal cancer and death. *N Engl J Med.* 2014;

519 370:1298-306.

520 5. Pohl H, Srivastava A, Bensen SP, Anderson P, Rothstein RI, Gordon SR, Levy LC, Toor

521 A, Mackenzie TA, Rosch T, Robertson DJ. Incomplete polyp resection during

522 colonoscopy-results of the complete adenoma resection (CARE) study.

523 *Gastroenterology.* 2013; 144:74-80.e1.

524 6. Weinberg DS, Barkun A, Turner BJ. Colorectal Cancer Screening in the United States:

525 What Is the Best FIT? *Ann Intern Med.* 2016.

526 7. Bogaert J, Prenen H. Molecular genetics of colorectal cancer. *Ann Gastroenterol.*

527 2014; 27:9-14.

528 8. Marley AR, Nan H. Epidemiology of colorectal cancer. *Int J Mol Epidemiol Genet.*

529 2016; 7:105-14.

530 9. Calle EE, Rodriguez C, Walker-Thurmond K, Thun MJ. Overweight, obesity, and

531 mortality from cancer in a prospectively studied cohort of U.S. adults. *N Engl J Med.*

532 2003; 348:1625-38.

533 10. Kinugasa T, Akagi Y. Status of colitis-associated cancer in ulcerative colitis. *World J*

534 *Gastrointest Oncol.* 2016; 8:351-7.

535 11. Ruder EH, Laiyemo AO, Graubard BI, Hollenbeck AR, Schatzkin A, Cross AJ. Non-

536 steroidal anti-inflammatory drugs and colorectal cancer risk in a large, prospective

537 cohort. *Am J Gastroenterol.* 2011; 106:1340-50.

538 12. Friis S, Riis AH, Erichsen R, Baron JA, Sorensen HT. Low-Dose Aspirin or

539 Nonsteroidal Anti-inflammatory Drug Use and Colorectal Cancer Risk: A Population-

540 Based, Case-Control Study. *Ann Intern Med.* 2015; 163:347-55.

541 13. Hamoya T, Fujii G, Miyamoto S, Takahashi M, Totsuka Y, Wakabayashi K, Toshima J,

542 Mutoh M. Effects of NSAIDs on the risk factors of colorectal cancer: a mini review.

543 *Genes Environ.* 2016; 38:6.

544 14. Pai SG, Fuloria J. Novel therapeutic agents in the treatment of metastatic colorectal

545 cancer. *World J Gastrointest Oncol.* 2016; 8:99-104.

546 15. Sjoblom T, Jones S, Wood LD, Parsons DW, Lin J, Barber TD, Mandelker D, Leary RJ,

547 Ptak J, Silliman N, Szabo S, Buckhaults P, Farrell C, Meeh P, Markowitz SD, Willis J,

548 Dawson D, Willson JK, Gazdar AF, Hartigan J, Wu L, Liu C, Parmigiani G, Park BH,

549 Bachman KE, Papadopoulos N, Vogelstein B, Kinzler KW, Velculescu VE. The

550 consensus coding sequences of human breast and colorectal cancers. *Science.* 2006;

551 314:268-74.

- 552 16. Fearon ER. Molecular genetics of colorectal cancer. *Annu Rev Pathol.* 2011; 6:479-
553 507.
- 554 17. Gerlach JP, Emmink BL, Nojima H, Kranenburg O, Maurice MM. Wnt signaling
555 induces accumulation of phosphorylated beta-catenin in two distinct cytosolic
556 complexes. *Open Biol.* 2014; 4:140120.
- 557 18. Kuhn M. Molecular physiology of membrane guanylyl cyclase receptors. *Physiol Rev.*
558 2016; 96:751-804.
- 559 19. Schulz S, Green CK, Yuen PS, Garbers DL. Guanylyl cyclase is a heat-stable
560 enterotoxin receptor. *Cell.* 1990; 63:941-8.
- 561 20. Fiskerstrand T, Arshad N, Haukanes BI, Tronstad RR, Pham KD, Johansson S, Havik
562 B, Tonder SL, Levy SE, Brackman D, Boman H, Biswas KH, Apold J, Hovdenak N,
563 Visweswariah SS, Knappskog PM. Familial diarrhea syndrome caused by an
564 activating GUCY2C mutation. *N Engl J Med.* 2012; 366:1586-95.
- 565 21. Romi H, Cohen I, Landau D, Alkrinawi S, Yerushalmi B, Hershkovitz R, Newman-
566 Heiman N, Cutting GR, Ofir R, Sivan S, Birk OS. Meconium ileus caused by mutations
567 in GUCY2C, encoding the CFTR-activating guanylate cyclase 2C. *Am J Hum Genet.*
568 2012; 90:893-9.
- 569 22. Basu N, Saha S, Khan I, Ramachandra SG, Visweswariah SS. Intestinal cell
570 proliferation and senescence are regulated by receptor guanylyl cyclase C and p21. *J*
571 *Biol Chem.* 2014; 289:581-93.
- 572 23. Li P, Lin JE, Chervoneva I, Schulz S, Waldman SA, Pitari GM. Homeostatic control of
573 the crypt-villus axis by the bacterial enterotoxin receptor guanylyl cyclase C
574 restricts the proliferating compartment in intestine. *Am J Pathol.* 2007; 171:1847-
575 58.
- 576 24. Han X, Mann E, Gilbert S, Guan Y, Steinbrecher KA, Montrose MH, Cohen MB. Loss of
577 guanylyl cyclase C (GCC) signaling leads to dysfunctional intestinal barrier. *PLoS*
578 *One.* 2011; 6:e16139.
- 579 25. Lin JE, Snook AE, Li P, Stoecker BA, Kim GW, Magee MS, Garcia AV, Valentino MA,
580 Hyslop T, Schulz S, Waldman SA. GUCY2C opposes systemic genotoxic tumorigenesis
581 by regulating AKT-dependent intestinal barrier integrity. *PLoS One.* 2012; 7:e31686.
- 582 26. Brenna O, Bruland T, Furnes MW, Granlund A, Drozdov I, Emgard J, Bronstad G, Kidd
583 M, Sandvik AK, Gustafsson BI. The guanylate cyclase-C signaling pathway is down-
584 regulated in inflammatory bowel disease. *Scand J Gastroenterol.* 2015; 50:1241-52.
- 585 27. Garin-Laflam MP, Steinbrecher KA, Rudolph JA, Mao J, Cohen MB. Activation of
586 guanylate cyclase C signaling pathway protects intestinal epithelial cells from acute
587 radiation-induced apoptosis. *Am J Physiol Gastrointest Liver Physiol.* 2009;
588 296:G740-9.
- 589 28. Valentino MA, Lin JE, Snook AE, Li P, Kim GW, Marszalowicz G, Magee MS, Hyslop T,
590 Schulz S, Waldman SA. A uroguanylin-GUCY2C endocrine axis regulates feeding in
591 mice. *J Clin Invest.* 2011; 121:3578-88.
- 592 29. Kim GW, Lin JE, Snook AE, Aing AS, Merlino DJ, Li P, Waldman SA. Calorie-induced
593 ER stress suppresses uroguanylin satiety signaling in diet-induced obesity. *Nutr*
594 *Diabetes.* 2016; 6:e211.
- 595 30. Birbe R, Palazzo JP, Walters R, Weinberg D, Schulz S, Waldman SA. Guanylyl cyclase
596 C is a marker of intestinal metaplasia, dysplasia, and adenocarcinoma of the
597 gastrointestinal tract. *Hum Pathol.* 2005; 36:170-9.

- 598 31. Carrithers SL, Barber MT, Biswas S, Parkinson SJ, Park PK, Goldstein SD, Waldman
599 SA. Guanylyl cyclase C is a selective marker for metastatic colorectal tumors in
600 human extraintestinal tissues. Proc Natl Acad Sci U S A. 1996; 93:14827-32.
- 601 32. Cagir B, Gelmann A, Park J, Fava T, Tankelevitch A, Bittner EW, Weaver EJ, Palazzo
602 JP, Weinberg D, Fry RD, Waldman SA. Guanylyl cyclase C messenger RNA is a
603 biomarker for recurrent stage II colorectal cancer. Ann Intern Med. 1999; 131:805-
604 12.
- 605 33. **Wilson C, Lin JE, Li P, Snook AE, Gong J, Sato T, Liu C, Gironde MA, Rui H, Hyslop T,
606 Waldman SA. The paracrine hormone for the GUCY2C tumor suppressor, guanylin, is
607 universally lost in colorectal cancer. Cancer Epidemiol Biomarkers Prev. 2014;
608 23:2328-37.
- 609 **Demonstrates near universal loss of the GUCY2C ligand, guanylin, in primary**
610 **colorectal tumors compared to matched normal adjacent tissue**
- 611 34. Steinbrecher KA, Tuohy TM, Heppner Goss K, Scott MC, Witte DP, Groden J, Cohen
612 MB. Expression of guanylin is downregulated in mouse and human intestinal
613 adenomas. Biochem Biophys Res Commun. 2000; 273:225-30.
- 614 35. Zhang L, Zhou W, Velculescu VE, Kern SE, Hruban RH, Hamilton SR, Vogelstein B,
615 Kinzler KW. Gene expression profiles in normal and cancer cells. Science. 1997;
616 276:1268-72.
- 617 36. Steinbrecher KA, Wowk SA, Rudolph JA, Witte DP, Cohen MB. Targeted inactivation
618 of the mouse guanylin gene results in altered dynamics of colonic epithelial
619 proliferation. Am J Pathol. 2002; 161:2169-78.
- 620 37. *Li P, Schulz S, Bombonati A, Palazzo JP, Hyslop TM, Xu Y, Baran AA, Siracusa LD,
621 Pitari GM, Waldman SA. Guanylyl cyclase C suppresses intestinal tumorigenesis by
622 restricting proliferation and maintaining genomic integrity. Gastroenterology. 2007;
623 133:599-607.
- 624 **Describes the mechanism of GUCY2C as a colorectal tumor suppressor**
- 625 38. *Lin JE, Li P, Snook AE, Schulz S, Dasgupta A, Hyslop TM, Gibbons AV, Marszlowicz G,
626 Pitari GM, Waldman SA. The hormone receptor GUCY2C suppresses intestinal tumor
627 formation by inhibiting AKT signaling. Gastroenterology. 2010; 138:241-54.
- 628 **Describes the mechanism of GUCY2C as a colorectal tumor suppressor**
- 629 39. Pitari GM, Di Guglielmo MD, Park J, Schulz S, Waldman SA. Guanylyl cyclase C
630 agonists regulate progression through the cell cycle of human colon carcinoma cells.
631 Proc Natl Acad Sci U S A. 2001; 98:7846-51.
- 632 40. Currie MG, Fok KF, Kato J, Moore RJ, Hamra FK, Duffin KL, Smith CE. Guanylin: an
633 endogenous activator of intestinal guanylate cyclase. Proc Natl Acad Sci U S A. 1992;
634 89:947-51.
- 635 41. Hamra FK, Forte LR, Eber SL, Pidhorodeckyj NV, Krause WJ, Freeman RH, Chin DT,
636 Tompkins JA, Fok KF, Smith CE, et al. Uroguanylin: structure and activity of a second
637 endogenous peptide that stimulates intestinal guanylate cyclase. Proc Natl Acad Sci
638 U S A. 1993; 90:10464-8.
- 639 42. Qian X, Prabhakar S, Nandi A, Visweswariah SS, Goy MF. Expression of GC-C, a
640 receptor-guanylate cyclase, and its endogenous ligands uroguanylin and guanylin
641 along the rostrocaudal axis of the intestine. Endocrinology. 2000; 141:3210-24.

- 642 43. Brenna O, Furnes MW, Munkvold B, Kidd M, Sandvik AK, Gustafsson BI. Cellular
643 localization of guanylin and uroguanylin mRNAs in human and rat duodenal and
644 colonic mucosa. *Cell Tissue Res.* 2016; 365:331-41.
- 645 44. Ikpa PT, Sleddens HF, Steinbrecher KA, Peppelenbosch MP, de Jonge HR, Smits R,
646 Bijvelds MJ. Guanylin and uroguanylin are produced by mouse intestinal epithelial
647 cells of columnar and secretory lineage. *Histochem Cell Biol.* 2016.
- 648 45. Perkins A, Goy MF, Li Z. Uroguanylin is expressed by enterochromaffin cells in the
649 rat gastrointestinal tract. *Gastroenterology.* 1997; 113:1007-14.
- 650 46. Cohen MB, Witte DP, Hawkins JA, Currie MG. Immunohistochemical localization of
651 guanylin in the rat small intestine and colon. *Biochem Biophys Res Commun.* 1995;
652 209:803-8.
- 653 47. Hamra FK, Eber SL, Chin DT, Currie MG, Forte LR. Regulation of intestinal
654 uroguanylin/guanylin receptor-mediated responses by mucosal acidity. *Proc Natl*
655 *Acad Sci U S A.* 1997; 94:2705-10.
- 656 48. Shailubhai K, Yu HH, Karunanandaa K, Wang JY, Eber SL, Wang Y, Joo NS, Kim HD,
657 Miedema BW, Abbas SZ, Boddupalli SS, Currie MG, Forte LR. Uroguanylin treatment
658 suppresses polyp formation in the Apc(Min/+) mouse and induces apoptosis in
659 human colon adenocarcinoma cells via cyclic GMP. *Cancer Res.* 2000; 60:5151-7.
- 660 49. Lin JE, Colon-Gonzalez F, Blomain E, Kim GW, Aing A, Stoecker B, Rock J, Snook AE,
661 Zhan T, Hyslop TM, Tomczak M, Blumberg RS, Waldman SA. Obesity-Induced
662 Colorectal Cancer Is Driven by Caloric Silencing of the Guanylin-GUCY2C Paracrine
663 Signaling Axis. *Cancer Res.* 2016; 76:339-46.
- 664 50. Pitari GM, Zingman LV, Hodgson DM, Alekseev AE, Kazerounian S, Bienengraeber M,
665 Hajnoczky G, Terzic A, Waldman SA. Bacterial enterotoxins are associated with
666 resistance to colon cancer. *Proc Natl Acad Sci U S A.* 2003; 100:2695-9.
- 667 51. Rao S, Lembo AJ, Shiff SJ, Lavins BJ, Currie MG, Jia XD, Shi K, MacDougall JE, Shao JZ,
668 Eng P, Fox SM, Schneier HA, Kurtz CB, Johnston JM. A 12-week, randomized,
669 controlled trial with a 4-week randomized withdrawal period to evaluate the
670 efficacy and safety of linaclotide in irritable bowel syndrome with constipation. *Am J*
671 *Gastroenterol.* 2012; 107:1714-24.
- 672 52. Chey WD, Lembo AJ, Lavins BJ, Shiff SJ, Kurtz CB, Currie MG, MacDougall JE, Jia XD,
673 Shao JZ, Fitch DA, Baird MJ, Schneier HA, Johnston JM. Linaclotide for irritable bowel
674 syndrome with constipation: a 26-week, randomized, double-blind, placebo-
675 controlled trial to evaluate efficacy and safety. *Am J Gastroenterol.* 2012; 107:1702-
676 12.
- 677 53. Lembo AJ, Schneier HA, Shiff SJ, Kurtz CB, MacDougall JE, Jia XD, Shao JZ, Lavins BJ,
678 Currie MG, Fitch DA, Jeglinski BI, Eng P, Fox SM, Johnston JM. Two randomized trials
679 of linaclotide for chronic constipation. *N Engl J Med.* 2011; 365:527-36.
- 680 54. Shailubhai K, Palejwala V, Arjunan KP, Saykhedkar S, Nefsky B, Foss JA, Comiskey S,
681 Jacob GS, Plevy SE. Plecanatide and dolcanatide, novel guanylate cyclase-C agonists,
682 ameliorate gastrointestinal inflammation in experimental models of murine colitis.
683 *World J Gastrointest Pharmacol Ther.* 2015; 6:213-22.
- 684 55. Shailubhai K, Comiskey S, Foss JA, Feng R, Barrow L, Comer GM, Jacob GS.
685 Plecanatide, an oral guanylate cyclase C agonist acting locally in the gastrointestinal
686 tract, is safe and well-tolerated in single doses. *Dig Dis Sci.* 2013; 58:2580-6.

- 687 56. ClinicalTrials.gov Registry [Published 2016 [cited October 24, 2016]. Available from:
688 <http://www.clinicaltrials.gov>.
- 689 57. Collett D, Mumford L, Banner NR, Neuberger J, Watson C. Comparison of the
690 incidence of malignancy in recipients of different types of organ: a UK Registry
691 audit. *Am J Transplant*. 2010; 10:1889-96.
- 692 58. Engels EA, Pfeiffer RM, Fraumeni JF, Jr., Kasiske BL, Israni AK, Snyder JJ, Wolfe RA,
693 Goodrich NP, Bayakly AR, Clarke CA, Copeland G, Finch JL, Fleissner ML, Goodman
694 MT, Kahn A, Koch L, Lynch CF, Madeleine MM, Pawlish K, Rao C, Williams MA,
695 Castenson D, Curry M, Parsons R, Fant G, Lin M. Spectrum of cancer risk among US
696 solid organ transplant recipients. *JAMA*. 2011; 306:1891-901.
- 697 59. Pagès F, Berger A, Camus M, Sanchez-Cabo F, Costes A, Molitor R, Mlecnik B,
698 Kirilovsky A, Nilsson M, Damotte D, Meatchi T, Bruneval P, Cugnenc P-H,
699 Trajanoski Z, Fridman W-H, Galon J. Effector memory T cells, early metastasis, and
700 survival in colorectal cancer. *N Engl J Med*. 2005; 353:2654-66.
- 701 60. Pernot S, Terme M, Voron T, Colussi O, Marcheteau E, Tartour E, Taieb J. Colorectal
702 cancer and immunity: what we know and perspectives. *World J Gastroenterol*. 2014;
703 20:3738-50.
- 704 61. Snook AE, Waldman SA. Advances in cancer immunotherapy. *Discov Med*. 2013;
705 15:120-5.
- 706 62. Chen J, Raju GS, Jogunoori W, Menon V, Majumdar A, Chen JS, Gi YJ, Jeong YS, Phan L,
707 Belkin M, Gu S, Kundra S, Mistry NA, Zhang J, Su X, Li S, Lin SH, Javle M, McMurray JS,
708 Rahlfs TF, Mishra B, White J, Rashid A, Beauchemin N, Weston BR, Shafi MA,
709 Stroehlein JR, Davila M, Akbani R, Weinstein JN, Wu X, Mishra L. Mutational profiles
710 reveal an aberrant TGF-beta-CEA regulated pathway in colon adenomas. *PLoS One*.
711 2016; 11:e0153933.
- 712 63. McCann KJ, Mander A, Cazaly A, Chudley L, Stasakova J, Thirdborough SM, King A,
713 Lloyd-Evans P, Buxton E, Edwards C, Halford S, Bateman A, O'Callaghan A, Clive S,
714 Anthoney A, Jodrell DI, Weinschenk T, Simon P, Sahin U, Thomas GJ, Stevenson FK,
715 Ottensmeier CH. Targeting carcinoembryonic antigen with DNA vaccination: on-
716 target adverse events link with immunologic and clinical outcomes. *Clin Cancer Res*.
717 2016; 22:4827-36.
- 718 64. Snook AE, Stafford BJ, Li P, Tan G, Huang L, Birbe R, Schulz S, Schnell MJ, Thakur M,
719 Rothstein JL, Eisenlohr LC, Waldman SA. Guanylyl cyclase C-induced
720 immunotherapeutic responses opposing tumor metastases without autoimmunity. *J*
721 *Natl Cancer Inst*. 2008; 100:950-61.
- 722 65. Snook AE, Li P, Stafford BJ, Faul EJ, Huang L, Birbe RC, Bombonati A, Schulz S, Schnell
723 MJ, Eisenlohr LC, Waldman SA. Lineage-specific T-cell responses to cancer mucosa
724 antigen oppose systemic metastases without mucosal inflammatory disease. *Cancer*
725 *Res*. 2009; 69:3537-44.
- 726 66. **Snook AE, Magee MS, Schulz S, Waldman SA. Selective antigen-specific CD4(+) T-
727 cell, but not CD8(+) T- or B-cell, tolerance corrupts cancer immunotherapy. *Eur J*
728 *Immunol*. 2014; 44:1956-66.
- 729 **Describes the Ad5-GUCY2C vaccine, and offers the first demonstration of CD4+ T-cell**
730 **tolerance as a mechanism of cancer vaccine resistance**

- 731 67. Snook AE, Baybutt TR, Hyslop T, Waldman SA. Preclinical evaluation of a replication-
732 deficient recombinant adenovirus serotype 5 vaccine expressing guanylate cyclase C
733 and the PADRE T-helper epitope. *Human gene therapy methods*. 2016; 27:238-50.
- 734 68. Snook AE. Could targeting T-helper cells aid the development of effective cancer
735 vaccines? *Immunotherapy*. 2014; 6:959-61.
- 736 69. Anderson CC. Application of central immunologic concepts to cancer: Helping T cells
737 and B cells become intolerant of tumors. *Eur J Immunol*. 2014; 44:1921-4.
- 738 70. Snook A, Baybutt T, Mastrangelo M, Lewis N, Goldstein S, Kraft W, Oppong Y, Hyslop
739 T, Myers R, Alexeev V, Eisenlohr L, Sato T, Waldman S. A Phase I study of Ad5-
740 GUCY2C-PADRE in stage I and II colon cancer patients. *Journal for ImmunoTherapy
741 of Cancer*. 2015; 3:P450.
- 742 71. Magee MS, Snook AE, Waldman SA. Adoptive cell therapy: at the forefront of cancer
743 immunotherapy. In: Watanabe HS, editor. *Horizons in Cancer Research*. 51.
744 Hauppauge, NY: Nova Science Publishers, Inc.; 2013.
- 745 72. Rosenberg SA, Yang JC, Sherry RM, Kammula US, Hughes MS, Phan GQ, Citrin DE,
746 Restifo NP, Robbins PF, Wunderlich JR, Morton KE, Laurencot CM, Steinberg SM,
747 White DE, Dudley ME. Durable complete responses in heavily pretreated patients
748 with metastatic melanoma using T-cell transfer immunotherapy. *Clin Cancer Res*.
749 2011; 17:4550-7.
- 750 73. Phan GQ, Rosenberg SA. Adoptive cell transfer for patients with metastatic
751 melanoma: the potential and promise of cancer immunotherapy. *Cancer Control*.
752 2013; 20:289-97.
- 753 74. Dudley ME, Wunderlich JR, Yang JC, Sherry RM, Topalian SL, Restifo NP, Royal RE,
754 Kammula U, White DE, Mavroukakis SA, Rogers LJ, Gracia GJ, Jones SA, Mangiameli
755 DP, Pelletier MM, Gea-Banacloche J, Robinson MR, Berman DM, Filie AC, Abati A,
756 Rosenberg SA. Adoptive cell transfer therapy following non-myeloablative but
757 lymphodepleting chemotherapy for the treatment of patients with refractory
758 metastatic melanoma. *J Clin Oncol*. 2005; 23:2346-57.
- 759 75. Johnson LA, Morgan RA, Dudley ME, Cassard L, Yang JC, Hughes MS, Kammula US,
760 Royal RE, Sherry RM, Wunderlich JR, Lee CC, Restifo NP, Schwarz SL, Cogdill AP,
761 Bishop RJ, Kim H, Brewer CC, Rudy SF, VanWaes C, Davis JL, Mathur A, Ripley RT,
762 Nathan DA, Laurencot CM, Rosenberg SA. Gene therapy with human and mouse T-
763 cell receptors mediates cancer regression and targets normal tissues expressing
764 cognate antigen. *Blood*. 2009; 114:535-46.
- 765 76. Morgan RA, Dudley ME, Wunderlich JR, Hughes MS, Yang JC, Sherry RM, Royal RE,
766 Topalian SL, Kammula US, Restifo NP, Zheng Z, Nahvi A, de Vries CR, Rogers-Freezer
767 LJ, Mavroukakis SA, Rosenberg SA. Cancer regression in patients after transfer of
768 genetically engineered lymphocytes. *Science*. 2006; 314:126-9.
- 769 77. *Gross G, Eshhar Z. Therapeutic potential of T cell chimeric antigen receptors
770 (CARs) in cancer treatment: counteracting off-tumor toxicities for safe CAR T cell
771 therapy. *Annu Rev Pharmacol Toxicol*. 2016; 56:59-83.
772 **Review of CAR-T-cell therapy and strategies to avoid off-target toxicity**
- 773 78. Eshhar Z, Waks T, Gross G, Schindler DG. Specific activation and targeting of
774 cytotoxic lymphocytes through chimeric single chains consisting of antibody-
775 binding domains and the gamma or zeta subunits of the immunoglobulin and T-cell
776 receptors. *Proc Natl Acad Sci U S A*. 1993; 90:720-4.

- 777 79. Maude SL, Frey N, Shaw PA, Aplenc R, Barrett DM, Bunin NJ, Chew A, Gonzalez VE,
778 Zheng Z, Lacey SF, Mahnke YD, Melenhorst JJ, Rheingold SR, Shen A, Teachey DT,
779 Levine BL, June CH, Porter DL, Grupp SA. Chimeric antigen receptor T cells for
780 sustained remissions in leukemia. *N Engl J Med*. 2014; 371:1507-17.
- 781 80. Grupp SA, Kalos M, Barrett D, Aplenc R, Porter DL, Rheingold SR, Teachey DT, Chew
782 A, Hauck B, Wright JF, Milone MC, Levine BL, June CH. Chimeric antigen receptor-
783 modified T cells for acute lymphoid leukemia. *N Engl J Med*. 2013; 368:1509-18.
- 784 81. Porter DL, Levine BL, Kalos M, Bagg A, June CH. Chimeric antigen receptor-modified
785 T cells in chronic lymphoid leukemia. *N Engl J Med*. 2011; 365:725-33.
- 786 82. Tran E, Robbins PF, Lu YC, Prickett TD, Gartner JJ, Jia L, Pasetto A, Zheng Z, Ray S,
787 Groh EM, Kriley IR, Rosenberg SA. T-Cell Transfer Therapy Targeting Mutant KRAS
788 in Cancer. *N Engl J Med*. 2016; 375:2255-62.
- 789 83. Morgan RA, Yang JC, Kitano M, Dudley ME, Laurencot CM, Rosenberg SA. Case report
790 of a serious adverse event following the administration of T cells transduced with a
791 chimeric antigen receptor recognizing ERBB2. *Mol Ther*. 2010; 18:843-51.
- 792 84. Parkhurst MR, Yang JC, Langan RC, Dudley ME, Nathan DA, Feldman SA, Davis JL,
793 Morgan RA, Merino MJ, Sherry RM, Hughes MS, Kammula US, Phan GQ, Lim RM,
794 Wank SA, Restifo NP, Robbins PF, Laurencot CM, Rosenberg SA. T cells targeting
795 carcinoembryonic antigen can mediate regression of metastatic colorectal cancer
796 but induce severe transient colitis. *Mol Ther*. 2011; 19:620-6.
- 797 85. ******Magee MS, Kraft CL, Abraham TS, Baybutt TR, Marszalowicz GP, Li P, Waldman SA,
798 Snook AE. GUCY2C-directed CAR-T cells oppose colorectal cancer metastases
799 without autoimmunity. *Oncol Immunology*. 2016; 5:e1227897.
800 **First pre-clinical study of a GUCY2C-directed CAR-T cell therapy for metastatic**
801 **colorectal cancer**
- 802 86. Ayyar BV, Arora S, O'Kennedy R. Coming-of-age of antibodies in cancer therapeutics.
803 *Trends Pharmacol Sci*. 2016.
- 804 87. Chee CE, Sinicrope FA. Targeted therapeutic agents for colorectal cancer.
805 *Gastroenterol Clin North Am*. 2010; 39:601-13.
- 806 88. Martinelli E, De Palma R, Orditura M, De Vita F, Ciardiello F. Anti-epidermal growth
807 factor receptor monoclonal antibodies in cancer therapy. *Clin Exp Immunol*. 2009;
808 158:1-9.
- 809 89. Peters C, Brown S. Antibody-drug conjugates as novel anti-cancer
810 chemotherapeutics. *Biosci Rep*. 2015; 35.
- 811 90. Papachristos A, Pippa N, Demetzos C, Sivolapenko G. Antibody-drug conjugates: a
812 mini-review. The synopsis of two approved medicines. *Drug Deliv*. 2016; 23:1662-6.
- 813 91. Marszalowicz GP, Snook AE, Magee MS, Merlino D, Berman-Booty LD, Waldman SA.
814 GUCY2C lysosomotropic endocytosis delivers immunotoxin therapy to metastatic
815 colorectal cancer. *Oncotarget*. 2014; 5:9460-71.
- 816 92. *****Almhanna K, Kalebic T, Cruz C, Faris JE, Ryan DP, Jung J, Wyant T, Fasanmade AA,
817 Messersmith W, Rodon J. Phase I study of the investigational anti-guanlyl cyclase
818 antibody-drug conjugate TAK-264 (MLN0264) in adult patients with advanced
819 gastrointestinal malignancies. *Clin Cancer Res*. 2016; 22:5049-57.
820 **First clinical trial results of TAK-264, a GUCY2C targeted antibody-drug conjugate**
- 821 93. Rahbari NN, Bork U, Motschall E, Thorlund K, Buchler MW, Koch M, Weitz J.
822 Molecular detection of tumor cells in regional lymph nodes is associated with

- 823 disease recurrence and poor survival in node-negative colorectal cancer: a
824 systematic review and meta-analysis. *J Clin Oncol.* 2012; 30:60-70.
- 825 94. Varghese A. Chemotherapy for stage II colon cancer. *Clin Colon Rectal Surg.* 2015;
826 28:256-61.
- 827 95. Waldman SA, Cagir B, Rakinic J, Fry RD, Goldstein SD, Isenberg G, Barber M, Biswas
828 S, Minimo C, Palazzo J, Park PK, Weinberg D. Use of guanylyl cyclase C for detecting
829 micrometastases in lymph nodes of patients with colon cancer. *Dis Colon Rectum.*
830 1998; 41:310-5.
- 831 96. *Waldman SA, Hyslop T, Schulz S, Barkun A, Nielsen K, Haaf J, Bonaccorso C, Li Y,
832 Weinberg DS. Association of GUCY2C expression in lymph nodes with time to
833 recurrence and disease-free survival in pN0 colorectal cancer. *JAMA.* 2009; 301:745-
834 52.
- 835 **First clinical trial defining the utility of GUCY2C qRT-PCR for molecular staging of**
836 **colorectal cancer**
- 837 97. Hyslop T, Weinberg DS, Schulz S, Barkun A, Waldman SA. Occult tumor burden
838 predicts disease recurrence in lymph node-negative colorectal cancer. *Clin Cancer*
839 *Res.* 2011; 17:3293-303.
- 840 98. Beaulieu M, Desaulniers M, Bertrand N, Deschesnes RG, Beaudry G, Garon G, Haince
841 JF, Houde M, Holzer TJ. Analytical performance of a qRT-PCR assay to detect
842 guanylyl cyclase C in FFPE lymph nodes of patients with colon cancer. *Diagn Mol*
843 *Pathol.* 2010; 19:20-7.
- 844 99. Sargent DJ, Resnick MB, Meyers MO, Goldar-Najafi A, Clancy T, Gill S, Siemons GO,
845 Shi Q, Bot BM, Wu TT, Beaudry G, Haince JF, Fradet Y. Evaluation of guanylyl cyclase
846 C lymph node status for colon cancer staging and prognosis. *Ann Surg Oncol.* 2011;
847 18:3261-70.
- 848 100. Haince JF, Houde M, Beaudry G, L'Esperance S, Garon G, Desaulniers M, Hafer LJ,
849 Heald JL, Lyle S, Grossman SR, Tetu B, Sargent DJ, Fradet Y. Comparison of
850 histopathology and RT-qPCR amplification of guanylyl cyclase C for detection of
851 colon cancer metastases in lymph nodes. *J Clin Pathol.* 2010; 63:530-7.
- 852 101. Van Cutsem E, Verheul HMW, Flamen P, Rougier P, Beets-Tan R, Glynne-Jones R,
853 Seufferlein T. Imaging in colorectal cancer: progress and challenges for the
854 clinicians. *Cancers (Basel).* 2016.
- 855 102. Weissleder R, Schwaiger MC, Gambhir SS, Hricak H. Imaging approaches to optimize
856 molecular therapies. *Sci Transl Med.* 2016; 8:355ps16.
- 857 103. Wolfe HR, Mendizabal M, Lleonart E, Cuthbertson A, Desai V, Pullan S, Fujii DK,
858 Morrison M, Pither R, Waldman SA. In vivo imaging of human colon cancer
859 xenografts in immunodeficient mice using a guanylyl cyclase C--specific ligand. *J*
860 *Nucl Med.* 2002; 43:392-9.
- 861 104. Liu D, Overbey D, Watkinson LD, Daibes-Figueroa S, Hoffman TJ, Forte LR, Volkert
862 WA, Giblin MF. In vivo imaging of human colorectal cancer using radiolabeled
863 analogs of the uroguanylin peptide hormone. *Anticancer Res.* 2009; 29:3777-83.
- 864 105. Maugeri-Sacca M, Vici P, Di Lauro L, Barba M, Amoreo CA, Gallo E, Mottolese M, De
865 Maria R. Cancer stem cells: are they responsible for treatment failure? *Future Oncol.*
866 2014; 10:2033-44.

- 867 106. Ricci-Vitiani L, Lombardi DG, Pilozzi E, Biffoni M, Todaro M, Peschle C, De Maria R.
868 Identification and expansion of human colon-cancer-initiating cells. *Nature*. 2006;
869 445:111-5.
- 870 107. Ma L, Liu T, Jin Y, Wei J, Yang Y, Zhang H. ABCG2 is required for self-renewal and
871 chemoresistance of CD133-positive human colorectal cancer cells. *Tumour Biol*.
872 2016; 37:12889-96.
- 873
- 874