

Thomas Jefferson University [Jefferson Digital Commons](https://jdc.jefferson.edu/)

[Department of Obstetrics and Gynecology](https://jdc.jefferson.edu/obgynfp)

Department of Obstetrics and Gynecology

3-1-2021

FIGO (international Federation of Gynecology and obstetrics) initiative on fetal growth: best practice advice for screening, diagnosis, and management of fetal growth restriction

Nir Melamed University of Toronto

Ahmet Baschat Johns Hopkins University

Yoav Yinon Tel-Aviv University Follow this and additional works at: [https://jdc.jefferson.edu/obgynfp](https://jdc.jefferson.edu/obgynfp?utm_source=jdc.jefferson.edu%2Fobgynfp%2F68&utm_medium=PDF&utm_campaign=PDFCoverPages)

O OBSIA DED SARE ARRIVES and Gynecology Commons Aristotle University of Thessaloniki
L'et US KNOW how access to this document benefits you

Federico Mecacci

University of Florence
Recommended Citation

Francesc; Berghella, Vincenzo; Nazareth, Amala; Tahlak, Muna; McIntyre, H David; Da Silva Costa, Fabrício;
See hext page foi la difficint Melamed, Nir; Baschat, Ahmet; Yinon, Yoav; Athanasiadis, Apostolos; Mecacci, Federico; Figueras, Kihara, Anne B; Hadar, Eran; McAuliffe, Fionnuala; Hanson, Mark; Ma, Ronald C; Gooden, Rachel; Sheiner, Eyal; Kapur, Anil; Divakar, Hema; Ayres-de-Campos, Diogo; Hiersch, Liran; Poon, Liona C; Kingdom, John; Romero, Roberto; and Hod, Moshe, "FIGO (international Federation of Gynecology and obstetrics) initiative on fetal growth: best practice advice for screening, diagnosis, and management of fetal growth restriction" (2021). Department of Obstetrics and Gynecology Faculty Papers. Paper 68. https://jdc.jefferson.edu/obgynfp/68

This Article is brought to you for free and open access by the Jefferson Digital Commons. The Jefferson Digital Commons is a service of Thomas Jefferson University's [Center for Teaching and Learning \(CTL\)](http://www.jefferson.edu/university/teaching-learning.html/). The Commons is a showcase for Jefferson books and journals, peer-reviewed scholarly publications, unique historical collections from the University archives, and teaching tools. The Jefferson Digital Commons allows researchers and interested readers anywhere in the world to learn about and keep up to date with Jefferson scholarship. This article has been accepted for inclusion in Department of Obstetrics and Gynecology Faculty Papers by an authorized administrator of the Jefferson Digital Commons. For more information, please contact: JeffersonDigitalCommons@jefferson.edu.

Authors

Nir Melamed, Ahmet Baschat, Yoav Yinon, Apostolos Athanasiadis, Federico Mecacci, Francesc Figueras, Vincenzo Berghella, Amala Nazareth, Muna Tahlak, H David McIntyre, Fabrício Da Silva Costa, Anne B Kihara, Eran Hadar, Fionnuala McAuliffe, Mark Hanson, Ronald C Ma, Rachel Gooden, Eyal Sheiner, Anil Kapur, Hema Divakar, Diogo Ayres-de-Campos, Liran Hiersch, Liona C Poon, John Kingdom, Roberto Romero, and Moshe Hod

FIGO (International Federation of Gynecology and Obstetrics) initiative on fetal growth: Best practice advice for screening, diagnosis, and management of fetal growth restriction

Nir Melamed1 | **Ahmet Baschat²** | **Yoav Yinon³** | **Apostolos Athanasiadis⁴** | **Federico Mecacci5** | **Francesc Figueras⁶** | **Vincenzo Berghella7** | **Amala Nazareth⁸** | **Muna Tahlak⁹** | **H. David McIntyre10** | **Fabrício Da Silva Costa11** | **Anne B. Kihara12** | **Eran Hadar13,14** | **Fionnuala McAuliffe15** | **Mark Hanson16,17** | **Ronald C. Ma18,19** | **Rachel Gooden20** | **Eyal Sheiner²¹** | **Anil Kapur²²** | **Hema Divakar²³** | **Diogo** Ayres-de-Campos²⁴ | Liran Hiersch²⁵ | Liona C. Poon²⁶ | John Kingdom²⁷ | **Roberto Romero28** | **Moshe Hod13,14***

4 Third Department of Obstetrics and Gynecology, Aristotle University of Thessaloniki, Thessaloniki, Greece

6 Maternal-Fetal Medicine Department, Barcelona Clinic Hospital, University of Barcelona, Barcelona, Spain

7 Division of Maternal-Fetal Medicine, Department of Obstetrics and Gynecology, Thomas Jefferson University, Philadelphia, PA, USA

 8 Jumeira Prime Healthcare Group, Emirates Medical Association, Dubai, United Arab Emirates

 $^{\circ}$ Latifa Hospital for Women and Children, Dubai Health Authority, Emirates Medical Association, Mohammad Bin Rashid University for Medical Sciences, Dubai, United Arab Emirates

¹⁰Mater Research, The University of Queensland, Brisbane, Qld, Australia

 11 Department of Gynecology and Obstetrics, Ribeirão Preto Medical School, University of São Paulo, Ribeirão Preto, São Paulo, Brazil

¹² African Federation of Obstetricians and Gynaecologists, Khartoum, Sudan

¹³Helen Schneider Hospital for Women, Rabin Medical Center, Petach Tikva, Israel

 16 Institute of Developmental Sciences, University Hospital Southampton, Southampton, UK

 18 Department of Medicine and Therapeutics, The Chinese University of Hong Kong, Hong Kong SAR, China

¹⁹Hong Kong Institute of Diabetes and Obesity, The Chinese University of Hong Kong, Hong Kong SAR, China

²⁰FIGO (International Federation of Gynecology and Obstetrics), London, UK

²¹Soroka University Medical Center, Ben-Gurion University of the Negev, Be'er-Sheva, Israel

22World Diabetes Foundation, Bagsværd, Denmark

23Divakars Speciality Hospital, Bengaluru, India

²⁴ Medical School and Santa Maria Hospital, University of Lisbon, Portugal

²⁵Sourasky Medical Center and Sackler Faculty of Medicine, Lis Maternity Hospital, Tel Aviv University, Tel Aviv, Israel

This is an open access article under the terms of the [Creative Commons Attribution](http://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2021 The Authors. *International Journal of Gynecology & Obstetrics* published by John Wiley & Sons Ltd on behalf of International Federation of Gynecology and Obstetrics

¹ Division of Maternal Fetal Medicine, Department of Obstetrics and Gynecology, Sunnybrook Health Sciences Centre, University of Toronto, Toronto, ON, Canada 2 Center for Fetal Therapy, Department of Gynecology and Obstetrics, Johns Hopkins University, Baltimore, MD, USA

 3 Fetal Medicine Unit, Department of Obstetrics and Gynecology, Sheba Medical Center, Tel-Hashomer, Sackler Faculty of Medicine, Tel-Aviv University, Tel Aviv, Israel

⁵ Maternal Fetal Medicine Unit, Division of Obstetrics and Gynecology, Department of Biomedical, Experimental and Clinical Sciences, University of Florence, Florence, Italy

¹⁴Sackler Faculty of Medicine, Tel-Aviv University, Tel Aviv, Israel

¹⁵UCD Perinatal Research Centre, School of Medicine, National Maternity Hospital, University College Dublin, Dublin, Ireland

¹⁷NIHR Southampton Biomedical Research Centre, University of Southampton, Southampton, UK

4 INVITELY GUNECOLOGY (W)

²⁶Department of Obstetrics and Gynecology, Prince of Wales Hospital, The Chinese University of Hong Kong, Shatin, Hong Kong SAR, China

²⁷Division of Maternal Fetal Medicine, Department of Obstetrics and Gynecology, Mount Sinai Hospital, University of Toronto, Toronto, ON, Canada

28Perinatology Research Branch, *Eunice Kennedy Shriver* National Institute of Child Health and Human Development, National Institutes of Health, U.S. Department of Health and Human Services, Bethesda, MD, and Detroit, MI, USA

***Correspondence**

Moshe Hod, Helen Schneider Hospital for Women, Rabin Medical Center, Petah Tikva, Israel. Email: hodroyal@inter.net.il

Funding Information

Publication of this Supplement was funded by FIGO.

Conflicts of Interest

The authors have no conflicts of interest. Dr Romero contributed to this work as part of his official duties as an employee of the United States Federal Government.

Keywords: detection, diagnosis, fetal growth restriction, FIGO initiative, management, monitoring

CONTENTS

 <u>ALLAMED ET AL. ALL CONDUCT ALL CONDUCT </u>

1 | **EXECUTIVE SUMMARY**

Fetal growth restriction (FGR) is defined as the failure of the fetus to meet its growth potential due to a pathological factor, most commonly placental dysfunction. Worldwide, FGR is a leading cause of stillbirth, neonatal mortality, and short- and long-term morbidity. Ongoing advances in clinical care, especially in definitions, diagnosis, and management of FGR, require efforts to effectively translate these changes to the wide range of obstetric care providers. This article highlights agreements based on current research in the diagnosis and management of FGR, and the areas that need more research to provide further clarification of recommendations.

The purpose of this article is to provide a comprehensive summary of available evidence along with practical recommendations concerning the care of pregnancies at risk of or complicated by FGR, with the overall goal to decrease the risk of stillbirth and neonatal mortality and morbidity associated with this condition. To achieve these goals, FIGO (the International Federation of Gynecology and

Obstetrics) brought together international experts to review and summarize current knowledge of FGR.

This summary is directed at multiple stakeholders, including healthcare providers, healthcare delivery organizations and providers, FIGO member societies, and professional organizations. Recognizing the variation in the resources and expertise available for the management of FGR in different countries or regions, this article attempts to take into consideration the unique aspects of antenatal care in low-resource settings (labelled "LRS" in the recommendations). This was achieved by collaboration with authors and FIGO member societies from low-resource settings such as India, Sub-Saharan Africa, the Middle East, and Latin America.

Aspects of FGR addressed in this article include prediction, diagnosis, investigation, management, and postpartum counselling. The main recommendations are given below and are summarized in Table 1 (section 8) and in the management algorithms for high-resource settings (Figure 1a) and low-resource settings (Figure 1b) (section 4).

Prediction and prevention of fetal growth restriction (FGR)

Investigation of fetal growth restriction (FGR) Recommendation Quality of evidence Strength of recommendation 1. Women with suspected FGR should undergo systematic assessment that includes the following: (1) detailed history; (2) detailed sonographic assessment for structural anomalies, soft markers, and sonographic signs related to fetal infection; (3) Doppler studies that include at least the umbilical artery and, when available, also the uterine and middle cerebral arteries; and (4) maternal screening for relevant congenital infections, which should be focused on cytomegalovirus and toxoplasmosis, but may also include rubella, herpes, syphilis, malaria, and Zika virus in cases at high risk. **LRS** The extent of investigation may be limited by available resources. Assessment should include screening for infections such as malaria and Zika virus in endemic areas. ⊕⊕⊕⊕ Strong 2. Confirmation of gestational age should be the first step when FGR is suspected. With the exception of pregnancies achieved by assisted reproductive technology, first-trimester crown–rump length is the most accurate method to date pregnancy when in the range of 7–60 mm. If more than one scan is performed in the first trimester, the earliest scan with a crown–rump length of at least 10 mm should be used. **LRS** In low-resource settings, dating may need to be based on menstrual history or symphysis–fundal height. ⊕⊕⊕⊕ Strong 3. Amniocentesis for karyotype (as well as microarray and polymerase chain reaction for infectious agents when available) should be offered to women with suspected FGR, especially in cases with early-onset severe (estimated fetal weight <3rd percentile) FGR, in the presence of sonographic findings associated with genetic or infectious etiologies, no obvious signs of placental dysfunction, and when the findings are likely to affect management. ⊕⊕⊕○ Strong

LRS The availability of genetic testing may be limited by available resources.

Strength of recommendation

Quality of evidence

⊕⊕○○ Strong

⊕⊕⊕○ Strong

⊕⊕○○ Strong

9. Women with a history of FGR should undergo close surveillance of fetal growth starting at 24-28 weeks. ⊕⊕⊕○ Strong

<u>CINECOLOGY</u> \circledast **WILEY 11**
 ELAMED ET AL. CONSTETRICS

This article is directed at multiple stakeholders with the intention of bringing attention to the assessment of fetal growth, with a particular focus on the screening, diagnosis, and management of FGR, which is a leading cause of stillbirth and neonatal mortality and morbidity. This article proposes to standardize and provide guidance for the screening, prevention, diagnosis, and management of FGR.

The intended target audience includes:

Healthcare providers: all those qualified to care for pregnant women (obstetricians, maternal-fetal medicine specialists, general

practitioners, midwives, nurses, advance practice clinicians, radiologists, sonographers, pediatricians, and neonatologists).

Healthcare delivery organizations and providers: governments, federal and state legislators, healthcare management organizations, health insurance organizations, international development agencies, and nongovernmental organizations.

Professional organizations: international, regional, and national professional organizations of obstetricians and gynecologists, obstetric ultrasound, family practitioners, pediatricians, neonatologists, and worldwide national organizations dedicated to the care of pregnant women and their offspring.

3 | ASSESSMENT OF QUALITY OF **EVIDENCE AND GRADING OF STRENGTH OF RECOMMENDATIONS**

In assessing the quality of evidence and grading of strength of recommendations, the article follows the terminology proposed by the Grading of Recommendations, Assessment, Development and Evaluation (GRADE) Working Group.⁵ This system uses consistent language and graphical descriptions for the strength and quality of the recommendations and the evidence on which they are based.

Recommendations are classified as strong or conditional (weak) (Table S1).⁶ The strength of recommendation is dependent not only on the quality of evidence, but also on factors such as risk–benefit, cost, resource allocation, values, and preferences. Thus, some recommendations may be based on low-quality evidence but still represent a benefit that outweighs the risks and burdens, and therefore may be strongly recommended.

The overall quality of evidence was assessed for each of the recommendations and expressed using four levels of quality: very low, low, moderate, and high (Table $S2$).⁷ Considerations for quality of evidence include primarily the study design and methodology. As such, evidence based on randomized controlled trials is considered high-quality evidence, observational studies provide moderate or low quality of evidence, and all others are very low. However, other parameters must be considered while assessing the level of evidence: risk of bias, study limitations, consistency of results, precision, publication bias, indirectness of evidence, and scarcity of evidence. For the quality of evidence, cross-filled circles are used: ⊕○○○ denotes very low-quality evidence; ⊕⊕⊙⊙ low quality; ⊕⊕⊕⊙ moderate quality; and ⊕⊕⊕⊕ high-quality evidence.

4 | **FETAL GROW TH RESTRIC TION: BACKGROUND, DEFINITION, ETIOLOGY, AND RISKS**

4.1 | **Background**

FGR is a common pregnancy complication that worldwide is a leading cause of stillbirth, neonatal mortality, and short- and long-term neonatal morbidity.⁸⁻¹⁵ The definition, diagnosis, and optimal management of FGR have generated controversy as clinicians strive for more harmonized care.

The purpose of this article is to provide a summary of the available evidence and provide recommendations regarding the early prediction and prevention, diagnosis, investigation, monitoring, and timing of delivery of pregnancies complicated by FGR, with the overall goal to decrease the risk of stillbirth and neonatal mortality and morbidity associated with this pregnancy complication. Given the variation in resources and expertise available for the assessment and monitoring of pregnancies complicated by FGR in different countries or regions, we have included, in addition to the standard of care or "best" recommendations, specific recommendations for low-resource settings, which are marked as **LRS** in the recommendation tables. Management algorithms for women in high-resource and low-resource settings are summarized in Figure 1a and 1b, respectively.

4.2 | **Terminology and definitions**

FGR is defined as the failure of the fetus to meet its growth potential due to a pathological factor, most commonly placental dysfunction. Clinically, this is reflected by a drop in fetal size percentiles over the course of gestation. However, fetal growth potential is difficult to determine, and serial assessments of fetal size to detect a drop in fetal weight percentile are usually not available. Instead, care providers most commonly have only a "snapshot" of fetal weight estimation at a given point in time. Therefore, in clinical practice, small for gestational age (SGA), defined as estimated fetal weight (EFW) or abdominal circumference below a certain threshold such as the 10th or 3rd percentile, is most commonly used to suspect FGR.

The use of SGA as a proxy for FGR has several limitations that need to be recognized. First, most SGA fetuses are constitutionally healthy small fetuses, whose smallness is merely the result of their predetermined growth potential (i.e. false-positive diagnosis of FGR). Second, some growth-restricted fetuses, depending on their original growth potential and timing of insult, may remain above the percentile threshold described above and may thus not be SGA (i.e. false-negative diagnosis of FGR). Third, the use of SGA as a proxy for FGR is limited by the accuracy of sonographic fetal weight estimation, which has an estimation error of up to ±15%–20%. Finally, the diagnosis of SGA is highly dependent on the growth chart being used, which can therefore have a considerable effect on the proportion of fetuses or infants flagged as SGA in a given population.

 <u>Consecution</u> α **B** α **B** α **C** α **B** α **B**

It should be noted that there is inconsistency in the literature regarding the terminology described above, where some use the term FGR to describe a fetus with an estimated weight below the 10th percentile for gestational age and the term SGA to describe an infant with birth weight below the 10th percentile for gestational age. However, for the purpose of this article, the term SGA is used to indicate an EFW or birth weight below the 10th percentile for gestational age, and the term FGR to refer to a small fetus that has failed to achieve its growth potential because of a pathologic process.

4.2.1 | Consensus-based definition of placentarelated FGR

The major member societies of FIGO follow a definition using the 10th percentile as a means of diagnosing an SGA fetus, which then leads to further testing, assessment, and follow-up. There are proposals to address the limitations of this definition, but their validity regarding reduction in adverse outcomes needs to be tested. For example, in an attempt to overcome some of the limitations described above, a consensus-based definition for placenta-mediated FGR has been proposed via a Delphi procedure.¹ To decrease the likelihood of false-positive and false-negative diagnosis of FGR, the consensus definition was based on a combination of measures of fetal size (fetal weight estimation and abdominal circumference) and abnormal Doppler findings in the umbilical, uterine, and middle cerebral arteries, as described in Box 1. The implementation of this definition is limited by the lack of a recommendation on which growth chart should be used to define the 10th and 3rd percentiles for EFW and fetal abdominal circumference. In addition, further research is needed to correlate this definition with adverse perinatal outcomes.

4.2.2 | Early- versus late-onset FGR

It has been suggested that FGR should be broadly classified, based on gestational age at the time of diagnosis, into early-onset FGR (<32 weeks) and late-onset FGR (≥32 weeks). The rationale underlying this classification is based on differences between these two phenotypes of FGR in severity, natural history, Doppler findings, association with hypertensive complications, placental findings, and management.16-18

Early-onset FGR has a prevalence of 0.5%–1%, is usually more severe, and is more likely to be associated with abnormal umbilical artery Doppler than late-onset FGR. The underlying placental pathology is frequently similar to that observed in cases of early-onset pre-eclampsia (maternal vascular malperfusion), which explains the strong association of early-onset FGR with pre-eclampsia. Therefore, early-onset FGR is usually easier to detect, and the natural history tends to follow a predictable sequence of Doppler changes in the umbilical artery and ductus venosus. The main challenge in cases of

early-onset FGR is management (i.e. timing of delivery), by attempting to determine the optimal balance between the opposing risks of stillbirth and prematurity.¹⁹

Late-onset FGR is more common than early-onset FGR with a prevalence of 5%–10%. In contrast to early-onset FGR, it is usually milder, is less likely to be associated with pre-eclampsia, and is usually associated with normal umbilical artery Doppler. Therefore, the main challenge with regard to late-onset FGR is diagnosis, while management (i.e. delivery) is relatively simple given that the diagnosis is commonly made during the late-preterm or term periods, where the risks associated with delivery are relatively small. The diagnosis of late-onset FGR mainly relies on adaptive changes in the cerebral circulation ("redistribution" or "brain-sparing effect"), which is reflected by low resistance to flow in the middle cerebral artery thereby generating a low cerebroplacental ratio, as described in section 8.1.7. Given that the umbilical artery and ductus venosus Doppler studies are usually normal in cases of late-onset FGR, the natural history in these

 • CONFIGURATION CONFIDENTIAL 15 CONFIDENTIAL 15 CONFIDENTIAL 15 CONFIDENTIAL 15 ALLERY 15 Routine patient education: Optimal weight gain in pregnancy, smoking cessation Determine risk of FGR based on history **Low risk High risk Preventive interventions:** Monitor and optimize maternal weight gain **Standard monitoring:** Aspirin 100-150 mg in the evening from ≤16 weeks Symphysis-fundal height (SFH) Consider ultrasound exam in the case of low SFH or Monitoring of fetal growth as available routinely at 32-36 weeks **FGR** is suspected

Investigation:

- Detailed history: medical, family, and obstetric history; confirm pregnancy dating when available
- Detailed anatomy scan: structural anomalies, soft markers, markers of fetal infection
- Doppler studies: umbilical artery
- Maternal screening for fetal infections including malaria and Zika virus when relevant
- Genetic consultation, amniocentesis for karyotype (and microarray and PCR for fetal infections if available)

Monitor:

- Educate mothers regarding fetal kick counting and symptoms of pre-eclampsia, monitor blood pressure
- Nonstress test (NST) and umbilical artery Doppler (see Table 1 for details regarding frequency)
- Fetal growth: every 2-3 weeks if available
- Administer antenatal corticosteroids per standard protocols

Delivery:

- Timing is based on gestational age and umbilical artery Doppler and NST findings (see Table 1 for details)
- Prefer ripening with balloon catheter over prostaglandin preparations when possible
- Continuous fetal heart rate monitoring when available

Postpartum follow-up and counselling for future pregnancies:

- Infant follow-up
- Educate women regarding preventive strategies to decrease the risk of future cardiovascular disease
- Counselling regarding risk of recurrence and management of future pregnancies, including optimization of nutritional status, smoking cessation, aspirin

FIGURE 1B Approach to screening, diagnosis, and management of fetal growth restriction in low-resource settings. Abbreviations: FGR, fetal growth restriction; NST, nonstress test; PCR, polymerase chain reaction; SFH, symphysis–fundal height.

Abbreviations: AC, fetal abdominal circumference; AREDV, absent or reversed end-diastolic velocity; CPR, cerebroplacental ratio; EFW, estimated fetal weight; PI, pulsatility index; UA, umbilical artery; UtA, uterine artery. Adapted from Gordijn et al.¹

Box 2 Common etiologies of fetal growth restriction.

Suboptimal uteroplacental perfusion of fetal nutrition

- a. Maternal (preplacental) factors
	- Hypoxemia (chronic lung disease, high altitude)
	- Anemia
	- Smoking, substance abuse (cocaine, methamphetamines)
	- Malabsorption, poor weight gain
	- Environmental toxins: air pollution, heavy metals (lead, mercury), perfluorooctanoic acid (PFOA)
- b. Placental factors
	- Maternal vascular malperfusion pathology (infarction, fibrin deposition, chronic abruption)
	- Fetal vascular malperfusion pathology
	- Chronic placental inflammation (e.g. villitis of unknown etiology)
	- Confined placental mosaicism
- c. Umbilical cord (postplacental) factors
	- Increased coiling
	- Increased cord length
	- True cord knot
	- Single umbilical artery
	- Marginal or velamentous cord insertion

Fetal disorders

- Genetic disorders (chromosomal, microdeletions/ duplications, single site mutations, epigenetic disorders)
- Structural anomalies (e.g. congenital heart disease, gastroschisis)
- Congenital infections (cytomegalovirus, toxoplasmosis, herpes, rubella, syphilis, Zika virus, malaria)
- Teratogen exposure (drugs, toxins)

cases is less predictable and there is a risk of sudden decompensation and stillbirth.^{16,19}

4.3 | **Etiology of fetal growth restriction**

FGR is often the result of one or more maternal, placental, or fetal disorders that interfere with the normal mechanisms regulating fetal growth.^{20,21} The most common etiologies of FGR are listed in Box 2. It is important to note that there is often confusion in the literature between "etiologies" (or pathogenetic pathways) and "risk factors" for FGR. For example, although maternal conditions such as chronic hypertension, kidney disease, systemic lupus erythematosus, and long-standing diabetes are often listed as "maternal etiologies" for FGR, these conditions should probably be viewed instead as maternal risk factors for abnormal placentation that may result in placenta-mediated FGR.

Given that maternal nutrition and fetal growth are closely related, $22,23$ maternal undernutrition is an important cause of FGR worldwide.²⁴⁻²⁶ The impact of maternal undernutrition on fetal growth depends on its timing and severity.²⁰ To date, maternal interventions in dietary advice and modifications have lacked significant success in

Box 3 Risks associated with fetal growth restriction.

Antenatal

- Stillbirth
- Pre-eclampsia
- Placental abruption
- Preterm birth
- **Neonatal (short term)**
- Neonatal mortality
- Neonatal morbidity (hypoglycemia, hyperbilirubinemia, hypothermia, necrotizing enterocolitis, respiratory morbidity, intraventricular hemorrhage)

Neonatal (long term)

- Neurodevelopmental disorders
- Metabolic syndrome (obesity, hypertension, diabetes, cardiovascular disease)

preventing FGR. While the mechanisms by which maternal anemia contributes to FGR are unclear, both impaired nutrient transport to the fetus²⁷ and abnormal placental adaptation to low maternal hemoglobin 28 have been suggested as potential mechanisms.

Abnormal placentation is a common cause of FGR, 29 which is often diagnosed by ultrasound Doppler studies ³⁰ and typical histopathological placental findings.31-33

Chromosomal abnormalities have been suggested to contribute to up to 5% of FGR cases; triploidy and trisomy 13 and 18 are important considerations in early-onset FGR and the risk of many aneuploidies is higher in the presence of structural fetal anomalies.34-36 In 1%–6% of cases of FGR with normal karyotype, submicroscopic (micro) duplications/deletions can be found using chromosomal microarray analysis, 35 even when FGR is an apparently isolated finding.³⁷ FGR is also more prevalent in fetuses with structural malformations, and the risk increases when multiple anomalies are present.³⁸

FGR is related to intrauterine infection in up to 5% of cases.^{20,39} Viral agents such as rubella, cytomegalovirus, HIV, and Zika are common causes of infection-related FGR.⁴⁰⁻⁴⁴ Protozoan infections like toxoplasmosis and malaria are another important cause, especially in endemic areas.^{45,46} The main mechanism involved in the pathogenesis of FGR in these cases is a decline in cell population.²⁰ Finally, maternal exposure to teratogens such as radiation,⁴⁷ illicit drugs, $48,49$ and alcohol⁵⁰ is another important etiology of FGR.

4.4 | **Risks associated with fetal growth restriction**

The main short- and long-term risks associated with FGR are listed in Box 3. It is associated with both fetal and obstetric complications. The most devastating complication is stillbirth, 51-53 and there is a well-established inverse relationship between weight percentile and the risk of stillbirth, $54-57$ which is more pronounced in the early **AREQUISTER AND ET AL. CONSIDERED ET AL**

preterm period than at term.⁵⁸ FGR is an important cause of iatrogenic preterm birth,⁵⁹ as early delivery remains the main and perhaps only strategy for the prevention of stillbirth in cases of severe FGR.^{16,60} FGR is also an independent risk factor for spontaneous preterm birth.⁶¹ Other obstetric complications associated with FGR include pre-eclampsia and placental abruption, as the pathophysiology of these conditions is often closely related.^{29,30,62-66}

Despite ongoing improvements in neonatal care, FGR is associated with increased neonatal mortality and short-term morbidity. The risk of perinatal mortality in term FGR is reported to be fiveto 10-fold higher than in appropriately grown neonates.^{57,61,67} The severity of FGR, Doppler abnormalities, and associated prematurity

are independent predictors of neonatal complications.⁶⁸ Among preterm infants, the co-presence of FGR further increases the risk of certain prematurity-related complications such as respiratory morbidity, intraventricular hemorrhage, necrotizing enterocolitis, and metabolic disorders.⁵⁷ Among term infants, FGR increases the risks of low cord artery $pH₁⁶⁹$ low Apgar score,⁶⁹ and neonatal complications such as hypoglycemia, hypothermia, and jaundice.⁷⁰⁻⁷²

Growth-restricted infants are also at risk of long-term complications including neurodevelopmental impairment $11,73-78$ and noncommunicable diseases.15,79-82 This is discussed in greater detail in section 9.1 (Infant follow-up).

4.5 | **Recommendations**

5 | **E ARLY PREDIC TION AND PRE VENTION OF FETAL GROW TH RESTRICTION**

Early prediction of FGR is important as it can identify women at high risk of FGR who may benefit from preventive interventions and close monitoring during pregnancy. Box 4 lists the most common risk factors for FGR. While the predictive value of individual risk factors is low, clinical prediction models that are based on combinations of the risk factors outlined below can considerably improve the prediction of FGR. One important limitation of most of the studies on early prediction of FGR is the lack of a gold standard for the antenatal or postnatal diagnosis of FGR. As such, there is wide variation among studies regarding the outcomes being predicted, including either SGA (birth weight below the 10th or 3rd percentile) or adverse perinatal outcomes that are associated with (but are not specific to) FGR. As many SGA infants are constitutionally small and healthy, differentiating between healthy small fetuses and those that are small due to FGR is critically important. As a rule, the prediction of early-onset severe FGR is better than of late-onset FGR.

Box 4 Risk factors for fetal growth restriction.

History-based risk factors

- a. Maternal demographics
- Advanced age
- Underweight
- Living in high altitude
- Severe anemia, hemoglobinopathies
- Environmental factors (air pollution, heavy metals, heat)
- b. Medical conditions
	- Chronic hypertension
	- Chronic kidney disease
	- Systemic lupus erythematosus
	- Inflammatory bowel disease
	- Antiphospholipid syndrome
	- Pregestational diabetes (long standing)
- c. Obstetric history
- Previous pregnancy affected by FGR or pre-eclampsia **Biochemical markers**
- Low PIGE
- Low PAPP-A
- High AFP
- **Ultrasound-based markers**
	- Uterine artery: pulsatility index >95th percentile
	- Uterine artery: bilateral notching
	- Marginal or velamentous cord insertion
	- Two-vessel cord (single umbilical artery)
	- Abnormal placental morphology^a
	- Decreased fetal growth velocity

Abbreviations: FGR, fetal growth restriction; PlGF, placental growth factor; PAPP-A, pregnancy-associated plasma protein-A; AFP, alpha-fetoprotein.

^aRefers to placental dimension (short-based thick placenta) and texture (calcifications, echogenic cystic lesions).

5.1 | **History-based risk factors**

Several maternal factors influence fetal growth and the risk of FGR: advanced maternal age, racial/ethnic origin (e.g. South Asian), consanguinity, low body mass index, nulliparity, use of recreational drugs and alcohol, assisted reproductive technology, and medical disorders such as chronic hypertension, diabetes mellitus, and autoimmune conditions (Box 4). $83-89$ Cigarette smoking is a common risk factor for FGR and reduces birth weight by an average of 200 g in a dose-response manner.⁹⁰ In a cohort of 33 602 pregnancies, maternal characteristics predicted 37% of women who subsequently delivered SGA neonates (birth weight <5th percentile) at a false-positive rate of 10%.⁸³

Some risk factors for FGR are especially relevant in low-resource countries. In a recent review from Africa, the main risk factors reported were low maternal nutritional status, HIV infection, malaria, and hypertensive diseases. Based on these findings, the authors concluded that to a large extent FGR in Africa is preventable through established interventions for malaria, HIV, and maternal undernutrition.42 In addition, exposure during pregnancy and lactation to toxic environmental chemicals and heavy metals has become a growing problem, especially in low-resource countries.⁹¹

5.2 | **Biochemical markers**

At this point there is no role for routine screening with serum biomarkers for FGR. However, when biochemical markers are available as part of prenatal genetic screening for trisomy 21, it may be reasonable to use this information for the purpose of risk stratification for FGR.

The placenta releases multiple factors into maternal circulation from the early stages of pregnancy, and first-trimester serum levels of some of these factors have been shown to be associated with subsequent placenta-mediated complications. $92,93$ Low levels of pregnancy-associated plasma protein-A (PAPP-A), a placental glycoprotein produced by the syncytiotrophoblast layer, have been associated with adverse pregnancy outcomes including SGA. A meta-analysis including 32 studies and 175 240 pregnancies found that PAPP-A levels below the 5th percentile had a moderate association with birth weight below the 10th percentile (OR 2.08, positive predictive value of 18%), while the association was stronger for PAPP-A levels below the 1st percentile (OR 3.4; positive predictive value of 28%).⁹⁴ Thus, although women with low PAPP-A are at increased risk for FGR, the majority of these women will have a normal pregnancy outcome, especially as an isolated biomarker in healthy women. However, a low PAPP-A level is often considered an indication for closer monitoring of fetal growth.⁹⁵ Elevated second-trimester maternal serum levels of alpha-fetoprotein are thought to reflect abnormal placental permeability and are associated with increased risk of placenta-mediated complications including FGR and stillbirth.^{96,97} The combination of low PAPP-A in the first trimester and high alpha-fetoprotein in the second trimester is particularly predictive of severe FGR.⁹⁸ Elevated human chorionic

gonadotropin (hCG) levels greater than 2.5 MoM in the second trimester, alone or combined with high alpha-fetoprotein levels, are also associated with an increased risk of SGA.⁹⁹

Angiogenic factors play a key role in the regulation of placental vascular development.¹⁰⁰ Placental growth factor (PIGF) is a proangiogenic factor highly expressed in the syncytiotrophoblast and the maternal endothelium. Impaired placentation is associated with reduced placental production of this protein. Low first-trimester PlGF levels have been shown to be associated with adverse pregnancy outcome including pre-eclampsia and SGA.¹⁰¹⁻¹⁰⁴ In a casecontrol study of 296 pregnancies with SGA and 609 controls, the detection rate of low PlGF for SGA at a false-positive rate of 5% and 10% was 15% and 21%, respectively. The combined use of PlGF and PAPP-A increased the detection rate to 19% and 27%, respectively.¹⁰³ A multicenter screening study found that the detection rate of a combined screening by maternal factors, fetal biometry, and serum PlGF and alpha-fetoprotein at 19–24 weeks for the delivery of SGA infants below the 5th percentile at less than 32, 32–36, and greater than or equal to 37 weeks of gestation was 100%, 76%, and 38%, respectively, at a false-positive rate of 10%.⁹⁶

Findings are less consistent for soluble fms-like tyrosine kinase-1 (sFlt-1), an antiangiogenic factor released from the placenta that results in maternal endothelial dysfunction characteristic of pre-eclampsia.105 Although maternal serum sFlt-1 levels are known to be elevated in pre-eclamptic pregnancies, a large case–control study demonstrated that high levels of sFlt-1 at 10–14 weeks were actually associated with a slightly reduced risk of SGA (OR 0.92; 95% CI, 0.88-0.96). 101 Therefore, the sFlt-1:PlGF ratio test used to diagnose pre-eclampsia should not be used in the first trimester as a screening test for FGR.¹⁰⁶

5.3 | **Ultrasound markers**

Several ultrasound-based markers have been shown to be predictive of FGR, including uterine artery Doppler, placental morphology, and placental volumes. However, given their modest predictive accuracy, they cannot be recommended for universal screening for FGR.

Increased uterine artery resistance largely reflects a failure of extravillous cytotrophoblast invasion and transformation of the spiral arteries and is associated with the development of pre-eclampsia and FGR due to maternal vascular malperfusion of the placenta.¹⁰⁷

First- and second-trimester abnormal uterine artery Doppler waveforms, defined as mean pulsatility index above the 95th percentile, have been shown to be associated with FGR.¹⁰⁸⁻¹¹⁰ In a large prospective cohort study of 4610 nulliparous women, uterine artery pulsatility index at 11^{+0} to 13^{+6} weeks predicted 60% of preterm and 17% of term SGA infants at a false-positive rate of 10%.¹¹¹ Although uterine artery Doppler shows promise, especially for the prediction of early-onset FGR, current evidence does not support routine screening with uterine artery Doppler for FGR in low- or high-risk pregnancies.112

Sonographic evaluation of the placenta is a routine part of the obstetric ultrasound examination. A method for systematic

two-dimensional (2D) placental ultrasound examination has been described, often in combination with other parameters ^{30,113,114} Abnormal placental morphology is defined by placental dimensions, shape, texture, and cord insertion. Placental shape is considered abnormal when the placental thickness is above 4 cm or greater than 50% of placental length. Placental texture is defined as normal when it is homogenous, and abnormal when the placenta is heterogeneous and contains multiple echogenic cystic lesions or has a jelly-like appearance with turbulent uteroplacental flow.115,116 Placental cord insertion is defined as central (>2 cm from placental disc margin), marginal (within 2 cm of margin), or velamentous (inserting into the surrounding membranes).¹¹⁴ In a cohort of 60 high-risk women with abnormal uterine artery Doppler, women with abnormal placental shape at 19–23 weeks had higher odds of FGR (OR 4.7) than women with normal placental shape.¹⁰⁸ However, the use of 2D placental imaging has significant limitations, including difficulty in assessing nonanterior placentas and a wide variability in the morphology of normal placentas. Furthermore, there are no large-scale prospective

Improvements in ultrasonographic imaging provide a tool for estimating placental volume using three- and four-dimensional scanning techniques. Placental volume has been proposed as a marker for various obstetric complications related to defective placental function, including FGR .^{117,118} A systematic review estimating the value of first-trimester 3D placental volume for the prediction of SGA found a detection rate of 24.7% at a 10% false-positive rate.¹¹⁹ Another parameter is the placental quotient, defined as the ratio of the placental volume to the fetal crown–rump length. The placental quotient was reported to have a high negative predictive value for perinatal complications but was not very useful when used for screening of SGA in a low-risk population, with a sensitivity of 27.1%.¹²⁰ The discriminatory ability of placental volume alone for SGA appears to be modest, but may be integrated into a multivariable screening model. However, the use of 3D placental volume as a routine screening tool for FGR is limited by the need for proper equipment and training required to obtain these measurements in a reproducible manner.

studies validating the use of this modality for prediction of FGR.¹¹⁴

5.4 | **Prediction models**

Currently there is no single screening test sufficiently predictive of FGR to recommend routine clinical use. Investigations are underway to combine various tests, but such prediction models have not been sufficiently validated in terms of outcomes studies and therefore must be considered investigative protocols at this time. In a prospective cohort of 4970 women, the combination of first-trimester maternal serum PAPP-A, beta hCG, maternal blood pressure, and uterine artery Doppler performed in the first trimester had a detection rate of 73% for early SGA (<34 weeks) but only 32% for late SGA (≥34 weeks).¹⁹ A different model that included maternal characteristics, first-trimester blood pressure, uterine artery pulsatility index, PlGF, and sFlt-1 was evaluated in a larger cohort of 9150 women **20 WILEY-CANECOLOGY** (W)

and achieved a detection rate of 86% for early-onset FGR and 66% for late-onset FGR, both at a false-positive rate of 10%.^{19,121} In the second trimester, the SCOPE consortium examined 5606 healthy nulliparous women with singleton pregnancies and found that the combination of clinical risk factors, 15-week biomarkers (53 biomarkers were used), and 20-week ultrasound (fetal biometry and Doppler studies of the umbilical and uterine arteries) had only a moderate detection rate for SGA below the 10th percentile, with a positive predictive value of 32% and a negative predictive value of 91%.¹²²

5.5 | **Prevention of fetal growth restriction in highrisk populations**

5.5.1 | Lifestyle modifications

Ideally, all women should plan their pregnancies, adopting a healthy lifestyle and optimizing any medical conditions and their body mass index. The preconception period provides an opportunity for health promotion with the aim of reducing accepted risk factors, including those associated with FGR.¹²³

Insufficient gestational weight gain has been associated with an increased risk of FGR, especially in women with low body mass index (BMI, calculated as weight in kilograms divided by height in meters squared).¹²⁴ Recognizing that these associations are only based on observational data, we still believe that it would be reasonable to recommend monitoring of weight gain and informing women of the target weight gain range, as recommended by the 2009 Institute of Medicine guidelines.¹²⁵ These guidelines recommend a total gestational weight gain of 12.5–18 kg (28–40 lb) for underweight women (BMI <18.5); 11.5–16 kg (25–35 lb) for the normal weight group (BMI 18.5–24.9); 7–11.5 kg (15–25 lb) for overweight women (BMI 25.0– 29.9); and 5-9 kg (11-20 lb) for obese women (BMI ≥30).¹²⁶

Substance use, including smoking, alcohol, and illicit drugs, is associated with low birth weight and increased perinatal morbidity and mortality.⁹⁰ Interventions to promote smoking cessation during pregnancy have been shown to result in a reduction in low birth weight (RR 0.81) and an increase in mean birth weight $(+33 \text{ g})$.¹²⁷ Women should be advised that smoking cessation at any point in gestation is of benefit, and that the greatest benefit is associated with cessation before 15 weeks of pregnancy.¹²⁸ The risk of SGA with alcohol intake is increased with as little as one drink per day.¹²⁹

5.5.2 | Medical interventions

Most studies on early prevention of placental complications have focused on pre-eclampsia, with the results often being extrapolated to FGR due to the common pathophysiology. However, to date, other than lifestyle modifications, no medical interventions to prevent FGR have been clearly established.

Aspirin is recommended for women at increased risk of pre-eclampsia, but there is some evidence that it may also reduce

the risk of FGR 130,131 In a recent meta-analysis of 45 trials that included 20 909 women at high risk of pre-eclampsia, the administration of aspirin starting at less than or equal to 16 weeks of pregnancy reduced the risk of FGR by nearly half (RR 0.56; 95% CI, 0.44–0.70), with higher dosages of aspirin associated with a greater reduction, favoring a dose of $100-150$ mg.¹³² A second individual patient data meta-analysis also supported earlier initiation of aspirin for the prevention of FGR, with an RR of 0.76 (95% CI, 0.61–0.94) for women randomized before 16 weeks versus an RR of 0.95 (95% CI, 0.84-1.08) for women randomized at 16 weeks or beyond.¹³¹ One randomized trial found that evening but not morning administration of aspirin is associated with reduction in the rate of pre-eclampsia and FGR.¹³³ However, it should be emphasized that most of the available data on aspirin come from studies that focused on the prevention of pre-eclampsia as the primary outcome in women at high risk of pre-eclampsia, with the prevention of FGR considered only as a secondary outcome. Furthermore, in the largest trial to date on the use of aspirin for the prevention of pre-eclampsia (ASPRE trial), aspirin was not associated with a reduction in the risk of SGA below the 10th, 5th, or 3rd percentile.¹³⁰ However, we believe that given the safety of aspirin and the overlap in the risk factors and pathogenesis of pre-eclampsia and FGR, it is reasonable to recommend aspirin to women at high risk of FGR, using the same regimen of aspirin used for women at high risk of pre-eclampsia. Most international guidelines recommend 100–150 mg aspirin to prevent FGR in women at high risk.134

The adjunct role of heparin in combination with aspirin to prevent placenta-mediated complications in high-risk situations was originally attributed to its anticoagulant properties and the speculative prevention of placental thrombosis. However, in vitro and in vivo data suggest heparins may have other biological properties including anti-inflammatory, complement inhibition, and proangiogenic activities.¹³⁵⁻¹³⁸ A study-level meta-analysis of six trials including 848 women showed that low-molecular-weight heparin (LMWH) was associated with a reduction in the composite outcome of pre-eclampsia, birth weight below the 10th percentile, placental abruption, or pregnancy loss after 20 weeks (RR 0.52; 95% CI, 0.32–0.86) with similar risk reduction for SGA below the 10th and 5th percentiles. However, the higher-quality trials suggest no treatment effect,¹³⁹ and a subsequent individual patient data meta-analysis looking at the same composite outcome found no beneficial effect of LMWH treatment (RR 0.64; 95% CI, 0.36-1.11).¹⁴⁰ Likewise, the enoxaparin for pre-eclampsia and intrauterine growth restriction (EPPI) trial included women at high risk for placenta-mediated complications (with a high proportion of women with prior FGR) and showed no difference in the rate of the composite outcome (pre-eclampsia or SGA <5th percentile) between treated and nontreated women.¹⁴¹ Therefore, based on the most up-to-date evidence, LMWH cannot be recommended for the prevention of FGR in women at high risk of placenta-mediated complications. Its use for the prevention of FGR should therefore be limited to research settings, for example in women already on aspirin who are found to have abnormal levels of angiogenic markers prior to fetal viability.¹⁴²

8. Low-molecular-weight heparin is not recommended for the prevention of FGR in women at high risk of FGR and its use should be limited to research settings. ⊕⊕⊕○ Strong Detection of FGR is based on the identification of a fetus that is smaller than expected for gestational age, through either physical examination (symphysis–fundal height, SFH) or ultrasound.

6.1 | **Symphysis–fundal height**

Measurement of SFH using a tape is a simple, inexpensive, and widely used strategy to screen for FGR.¹⁴³⁻¹⁴⁶ SFH is measured with the woman in a supine position using a nonelastic metric tape after she has emptied her bladder. To decrease the interobserver variability, a standardized technique for measuring SFH should be followed.^{144,145} SFH is defined as the distance from the upper border of the symphysis pubis bone to the top of the uterine fundus. 145 SFH measured in centimeters between 24 and 38 weeks of gestation approximates the gestational age. 147 Numerous local charts are currently used worldwide.¹⁴⁸⁻¹⁵⁶ with the recent addition of an international standard for SFH based on serial measurements.¹⁴⁵ However, the accuracy of SFH measurement in predicting SGA (EFW <10th percentile) is limited, and there are no randomized controlled trials that compare SFH measurement with serial ultrasound evaluation of fetal biometry.¹⁵⁷ In a meta-analysis of 34 observational studies, SFH was reported to have a sensitivity of 58% and a specificity of 87% for predicting birth weight below the 10th percentile. There was marked heterogeneity between studies, mainly due to the use of different SFH charts.¹⁵⁸ A single SFH measurement at 32–34 weeks of pregnancy has been reported to be approximately 65%–85% sensitive and 96% specific for detecting FGR.¹⁴³ It is important to acknowledge that factors such as maternal obesity, uterine leiomyomas, and polyhydramnios may further limit the accuracy of SFH as a screening tool.^{144,159}

6.2 | **Sonographic fetal weight estimation**

Sonographic fetal biometry is the cornerstone for detection of fetal growth disorders. Standard fetal biometry includes assessment of head circumference (HC), biparietal diameter, abdominal circumference (AC), and femur length (FL). Measurement of these biometric indices should be obtained by an experienced individual and in a standardized manner, as has been previously described.¹⁶⁰ Fetal weight is estimated based on various combinations of the four biometric indices described above, using one of many published equations.¹⁶¹⁻¹⁶⁵ The accuracy of most equations falls within the range of ±10%, and the error has been shown to be greater at the extremes of fetal weight, and to be affected by factors such as fetal sex, presentation, and plurality (greater in twin gestations).^{162-164,166-171} Several studies have compared the accuracy of various equations. Most studies concluded that equations that are based on 3–4 biometric indices (rather than only 1–2 indices) provide the most consistent

and accurate results. A recent systematic review ¹⁶⁵ found that the Hadlock equation, based on three indices (HC, AC, and FL: Log10 we ight = 1.326 − 0.00326*AC*FL + 0.0107*HC + 0.0438*AC + 0.158*F L), 2 provided the greatest accuracy. Since the accuracy of the various equations may vary between different populations, it may be reasonable for radiologists, sonographers, or care providers to choose an equation that has been validated within their local population and within the gestational age range in which it will be used. However, if such information is not available—a very frequent scenario—it seems reasonable to use the Hadlock equation as described above.

6.3 | **Is there a role for routine third-trimester ultrasound to assess fetal growth?**

In many countries, measurement of SFH is the primary screening tool for FGR in low-risk pregnancies and ultrasound measurement of fetal biometry is performed only when indicated on the basis of risk factors or abnormal SFH. $134,143,172$ -174 However, this approach fails to identify the majority of FGR infants,¹⁴⁶ a concerning finding given that undetected FGR is associated with increased risk of adverse perinatal outcome and stillbirth.53,175

An alternative approach is to perform a routine third-trimester ultrasound for fetal weight estimation. However, a strategy for routine third-trimester ultrasound in low-risk pregnancies is not supported by available data and cannot be recommended.¹⁷⁶⁻¹⁷⁸

A meta-analysis of 13 trials assessed the effect of routine sonographic weight estimation at more than 24 weeks of gestation on pregnancy outcomes in both unselected and low-risk pregnancies.178 The authors found no association between routine sonographic EFW and adverse pregnancy outcomes including perinatal mortality, preterm birth, induction of labor, or cesarean section. In a recent randomized controlled trial of women with uncomplicated pregnancies, the use of serial (every 4 weeks) third-trimester ultrasound was superior to routine care in the detection of a composite outcome of fetal growth or amniotic fluid abnormalities (RR 3.43; 95% CI, 1.64-7.17).¹⁷⁹ However, it is important to note that the incidence of maternal or fetal morbidity was not significantly different between the groups. Similar results were reported by others.¹⁸⁰ In contrast, the Pregnancy Outcome Prediction (POP) study prospectively assessed 3977 women and compared the detection of SGA (birth weight <10th percentile) by routine ultrasound versus clinically indicated ultrasound in the third trimester.¹⁸¹ The detection rate of SGA was nearly tripled in the routine ultrasound group (57% vs 20%). The risk of neonatal morbidity was increased only in the subset of SGA fetuses with fetal abdominal circumference growth velocity in the lowest decile (RR 3.9; 95% CI, 1.9–8.1), emphasizing the importance of combined analysis of fetal biometry and fetal growth velocity for better detection of fetuses at risk.¹⁸² Furthermore, it has been suggested that the prediction of FGR based on routine third-trimester ultrasound can be improved by integrating EFW with additional biomarkers. A combined screening model that included maternal characteristics, third-trimester EFW and placental **BELAMED ET AL. ALL PROPERTY AND RELANSED ET AL. CONSTRES AND INCORPORATION CONSTRES AND INCORPORATION CONSTRESS AND INCORPORATION CONSTRESS AND INCORPORATION CONSTRESS AND INCORPORATION CONSTRESS AND INCORPORATION CO**

Doppler, and biochemical markers (PlGF and estriol) achieved better performance than EFW alone in the detection of FGR (77% vs 64%) at a 10% false-positive rate.¹⁸³

There are many conceptual explanations to support thirdtrimester ultrasound as it can assist in the diagnosis of clinically significant findings other than FGR, including fetal malpresentation,¹⁸⁴ disorders of amniotic fluid, and fetal anomalies, $185,186$ especially when combined with Doppler measurements and biochemical markers.^{95,187-189} However, there is no evidence that this information improves outcomes when performed routinely in lowrisk pregnancies.

6.4 | **Which growth chart should be used to determine fetal weight percentile?**

The interpretation of sonographic EFW depends on gestational age and is commonly classified as appropriate for gestational age, SGA, or large for gestational age, based on the calculation of EFW percentile using one of the many available growth charts. The choice of growth chart has been shown to have a considerable impact on the proportion of fetuses classified as either SGA or large for gestational age.^{190,191} Over the past several years there has been an ongoing debate regarding the optimal growth chart that should be used, and numerous studies have compared the performance of a wide variety of charts in different populations with conflicting results. Prior to further discussion of specific charts, it is important to clarify the terminology and the types of charts that are currently available.

6.4.1 | Growth references versus growth standards

Growth references are descriptive charts that provide information on the distribution of weight of *all* newborns in a given population, and as such they include both normal and complicated pregnancies. Although growth references are useful as they provide information on the overall distribution of birth weight in the population, their use for the purpose of antenatal detection of FGR may be challenging as they are affected by the rate of pathologies in the population. For example, in populations with a high rate of large fetuses (e.g. due to a high rate of obesity and diabetes), the reference would be shifted upward. Similarly, in populations with a high rate of FGR (e.g. due to a high rate of malnutrition), the reference would be shifted downward.

For that reason, it may be reasonable to prefer growth standards over growth references for the antenatal detection of FGR. Growth standards are prescriptive charts that are based only on low-risk or uncomplicated pregnancies, and as such provide information on what is the optimal fetal growth. There is variation between different growth standards with regard to the definition of "low-risk" pregnancies; while some standards excluded women with pre-existing medical conditions and pregnancy complications, others also excluded

women below or above certain height or weight, women with suboptimal nutrition, low socioeconomic status, exposure to air pollution, high altitude etc. Since growth standards include only low-risk uncomplicated pregnancies, their distribution is usually narrower (i.e. the 10th and 90th percentiles are closer to the mean) compared with growth references.

One important and practical aspect regarding the use of reference versus standard charts relates to the weight percentile threshold that should be used to trigger further evaluation for FGR. When using a growth reference, it is reasonable to use the 10th percentile for that purpose, as a considerable proportion of infants below the10th percentile will be affected by pathology. In the case of growth standard, however, using the same threshold of the 10th percentile would, per definition, identify 10% of the low-risk pregnancies as suspected for FGR, which is not practical. Therefore, when using a growth standard, a lower threshold—such as the 5th or 3rd percentile—should be used to indicate further evaluation for FGR.

6.4.2 | Charts based on birth weight versus sonographic fetal weight estimation

A second important distinction is between growth charts that are based on birth weight versus those that are based on sonographic EFW. Birth weight-based charts rely on cross-sectional data of infant birth weights across the full range of gestational ages, usually obtained from large databases. Different types of regression techniques are then used to calculate the mean and various percentiles of birth weight across gestation. These charts are commonly used as they are easy to develop. However, their main limitation is that infants born prematurely (before 37 weeks) are more likely to be affected by placental dysfunction and to be growth restricted. Therefore, these charts are likely to underestimate the optimal weight of fetuses during the preterm period, which in turn may lead to an underdiagnosis of FGR before 37 weeks. This is illustrated in Figure 2, where the birth weightbased chart of Alexander (USA)¹⁹² is compared with several ultrasound-based charts.

Therefore, it seems reasonable to prefer growth charts that are based on sonographic EFW over those that are based on birth weight. Ultrasound-based growth charts are more difficult and expensive to develop, as they are usually based on data from prospective longitudinal studies where women undergo several sonographic weight estimations during pregnancy. However, these charts do not share the limitation of birth weight-based charts, described above, and are thus more likely to reflect the optimal fetal growth throughout pregnancy (Figure 2). Another reason why ultrasound-based charts should be preferred is that the measure used during pregnancy to assess fetal growth is sonographic EFW; it is therefore more appropriate to compare it to charts based on the same measure (i.e. sonographic EFW) rather than to charts based on birth weight. Some of the commonly used ultrasound based charts are presented in Figure 2.193

FIGURE 2 Comparison of the 10th percentile curves of common growth charts. Key: Hadlock: ultrasound-based chart 193 ; NICHD, National Institute of Child Health and Human Development $chart^{198}$; IG21, Intergrowth-21st chart¹⁹⁶; WHO, World Health Organization chart¹⁹⁷ Alexander: birth weight-based chart.¹⁹² [Colour figure can be viewed at [wileyonlinelibrary.com\]](www.wileyonlinelibrary.com)

6.4.3 | Universal versus customized charts

One final distinction is between universal and customized growth charts, which represent a spectrum of approaches towards the similarity of the genetic growth potential of different fetuses across the world. At one end of this spectrum there are universal charts that are based on the assumption that under optimal conditions, all fetuses are expected to have the same growth potential, irrespective of their country of origin or race and that the only reason for the differences currently observed between different countries or races are purely due to environmental factors, such as malnutrition and environmental toxins. These ultrasound-based charts are developed through multicenter, multinational, prospective longitudinal studies, where data on sonographic fetal growth from multiple countries are pooled into a single international universal chart. The best examples of such universal charts are the recently published Intergrowth-21st¹⁹⁴⁻¹⁹⁶ and World Health Organization (WHO) charts.197

Others, however, believe that the variation in fetal growth between countries and races is not solely the result of environmental factors. Instead, it is suggested that genetic variation in growth potential contributes to the observed differences in fetal growth between race groups, and that race-specific charts should therefore be preferred over universal charts. Examples of such race-specific charts are the National Institute of Child Health and Human Development (NICHD) charts which include separate charts for white, black, Hispanic, and Asian women, 198 and the recently

published PRB/NICHD customized standard for African American women.199

According to the third approach, growth charts should be adjusted not only for maternal race but also for other physiologic factors that are thought to determine fetal growth potential, such as maternal height, weight, parity, and fetal sex. One such example is the Gestation Related Optimal Weight (GROW) software for customized growth percentiles.^{200,201}

At the other end of the spectrum is the individualized growth assessment (IGA) approach, which is based on estimation of the growth potential of the individual fetus, calculated from the secondtrimester growth velocity of that fetus. These estimates are used to generate individualized trajectories that are used to interpret fetal growth during the third trimester ([https://igap.research.bcm.](https://igap.research.bcm.edu) [edu](https://igap.research.bcm.edu)).²⁰²⁻²⁰⁴ While compelling, this approach requires earlier ultrasound exams during pregnancy, as well as appropriate software, and is therefore challenging at present for the purpose of FGR screening in the general population, and especially in low-resource settings.

6.4.4 | Description of commonly available charts

The 10th percentile curves of some of the charts described above are compared in Figure 2. The Hadlock chart (1991), one of the most commonly used growth charts, is an ultrasound-based standard. It is based on a cohort of 392 low-risk, primarily white women from Texas. The Alexander chart (1996) is based on over 3 million singleton live births in the USA and is included as an example of a birth weight-based reference to illustrate their limitation, which is the underestimation of optimal fetal growth during the preterm period.

The goal of the Intergrowth-21st project (2014) was to develop a universal ultrasound-based prescriptive growth chart. This was a prospective longitudinal study of 4321 low-risk women from eight centers located in eight high- and middle-income countries.¹⁹⁴ The study had strict inclusion and exclusion criteria to ensure that participants were not exposed to environmental factors known to affect fetal growth, and it therefore aimed to reflect optimal fetal growth. Based on predetermined criteria, the authors concluded that the differences between participants from different countries in measures of skeletal growth (crown–rump length and head circumference) were similar enough to justify pooling the data, and they therefore generated a single universal chart. No information was provided on the differences between countries with respect to measures such as fetal weight estimation and abdominal circumference, which are used in clinical practice to detect FGR and are known to be associated with adverse perinatal outcomes. Interestingly, there were considerable differences in birth weight between infants from different countries, even in this highly selected group of women free from the negative influence of environmental factors known to affect fetal growth. For example, the mean birth weight at term in India was 2.9 kg, which was approximately 600 g lower than the mean birth weight in the UK (3.5 kg).¹⁹⁵ These differences have led some to question the validity of the pooled chart and of the hypothesis that underlies the Intergrowth-21st project.^{205,206} As demonstrated in Figure 2, the 10th percentile of the Intergrowth-21st chart is significantly lower throughout gestation than most other ultrasound-based standards.

At around the same time, the results of the NICHD growth study (2015) were published.¹⁹⁸ The overall design of this study was similar to that of the Intergrowth-21st study. It was a prospective longitudinal study of 2334 low-risk women from 12 centers in the USA. The authors found substantial differences in fetal weight between different race groups, and therefore developed separate race-specific growth charts for white, black, Hispanic, and Asian women. The 10th percentile of the NICHD growth chart for white women is included as an example in Figure 2.

The WHO fetal growth charts were published in 2017.¹⁹⁷ Similar to the Intergrowth-21st project, this study aimed to develop a prescriptive universal chart to extend the previously published WHO child growth standard 207 to the fetal period. The design of this study was also similar to that of Intergrowth-21st—a prospective longitudinal study of 1387 low-risk women from 10 centers in 10 high- and middle-income countries. Despite this, the results of the WHO study differed from those of Intergrowth-21st in two aspects. First, the 10th percentile of the WHO chart is considerably higher than the Intergrowth-21st chart, and is in fact almost identical to the 10th percentile of the Hadlock standard (Figure 2). Second, unlike Intergrowth-21st, the investigators of the WHO study found substantial differences in fetal growth between the various countries, and concluded that "…populations, even under optimal nutritional conditions and environment, vary and that fetal growth varies and should be considered when the WHO fetal growth charts or any growth references are applied".²⁰⁸ They expressed concern that use of a universal chart carries a risk of misclassification of FGR, 209-211 and recommended that their chart should be adjusted in each country to the local population.

The benefit of customized charts remains a matter of debate. The GROW software incorporates certain factors that are believed to determine fetal growth potential (maternal race, height, weight, parity, and fetal sex) to calculate the predicted optimal (customized) weight at 40 weeks for each individual fetus.^{200,201} The customized fetal growth curve is then determined retrospectively, based on a proportionality growth function derived from the ultrasound-based Hadlock standard.¹⁹³ The use of customized charts is appealing, especially in the setting of ethnically mixed populations where their use has been shown to decrease over- and underestimation of FGR rates in certain race groups.²¹² A large number of studies investigated the association of customized charts with adverse pregnancy outcomes compared with other birth weight- and ultrasound-based charts, with conflicting results. Several studies found that customized charts performed better at predicting stillbirth and adverse neonatal outcomes,^{66,201,211,213-216} while others found no benefit and concluded that the benefit reported by others is merely because they are based on an ultrasound-based chart and are thus more likely to reflect optimal fetal growth, while the act of customization has a minimal contribution to the stronger association with adverse

 BELAMED ET AL. ALL PROPERTY AND RELANSED ET AL. CONSTRUCT AND RELANSED ET AL. CONSTRUCT AND RELANSED ET AL. 25

outcome.217-219 Another criticism is that the GROW approach assumes that all fetuses follow the same growth trajectory (which is derived from the Hadlock chart)—an assumption that may not be true. Finally, it has been suggested that the required adjustment for multiple factors may be too complex for low-resource countries and, in that setting, a simple adjustment to only one factor—mean birth weight at 40 weeks in the local population—is as predictive for adverse perinatal outcomes as the fully customized GROW charts.²²⁰ It may thus be reasonable for care providers to compare the performance of customized growth charts in their population with that of noncustomized charts (as discussed below), especially in regions or countries with a mixed population where the benefit of customization is expected to be greatest.

6.4.5 | How to choose the best chart

The conflicting results and conclusions regarding the growth charts described above have led to an ongoing debate about the best approach (i.e. universal versus customized charts), as well as to considerable confusion among care providers over which chart they should be using in their local population.

The FIGO Safe Motherhood and Newborn Health Committee recently published a position paper on the choice of reference charts for fetal growth and size at birth. $³$ In that paper, the commit-</sup> tee reviewed in detail the commonly available charts and the available data on their predictive accuracy. The main conclusions were as follows: (1) local or regional charts are likely to be best to identify the 10th percentile of infants at highest risk, given that universal charts such as Intergrowth-21st are likely to under detect SGA fetuses in high-resource countries and, at the same time, over detect SGA in low- and middle-income countries; (2) as an alternative, universal standards such as Intergorwth-21st and WHO may be used with locally adjusted thresholds (e.g. 3rd or 5th percentile in lowor middle-income countries versus 15th or 20th percentile in highincome countries) to avoid under or overdetection of SGA; and (3) when assessing fetal size antenatally by ultrasound, fetal (i.e. ultrasound-based) charts should be used rather than birth weight-based charts. We fully endorse and support these recommendations.

Furthermore, we as well as others, $160,208$ believe that the decision on which chart to use can be further based on a comparison of performance of the various charts in the population of interest, using a local data set. This can be achieved by the following approaches: (1) statistical validation: finding the chart that matches best the distribution of fetal weight in low-risk pregnancies in the local population. That is, identifying the chart that when applied to the local population yields weight percentiles that follow a normal distribution centered at approximately the 50th percentile, and identifies approximately 10% of the low-risk population as being below the 10th percentile and above the 90th percentile, and approximately 5% of the population as being below the 5th percentile and above the 95th percentile. An example of this approach is provided in Figure 3; (2) outcome-based validation: finding the chart for which

FIGURE 3 Illustration of the statistical validation of two charts in a local population. The left chart shows a good match to the population of interest: the distribution of fetal weight percentiles based on this chart follows a normal distribution that is centered at the 50th percentile, with approximately 10% of the population below the 10th and above the 90th percentile. The right chart shows a poor fit for the population of interest as it is skewed to the right: it overdiagnoses fetuses as large for gestational age and underdiagnoses small-forgestational-age fetuses. [Colour figure can be viewed at [wileyonlinelibrary.com\]](www.wileyonlinelibrary.com)

the diagnosis of SGA has the best predictive value for adverse outcomes related to FGR..^{211,221} While this approach seems compelling, interpretation of the predictive value of the different charts for adverse outcomes may be challenging, as there is a trade-off between detection rate and false positive rate for adverse outcomes.²²¹ Thus, charts that are shifted upward (e.g. Hadlock, WHO) would have a higher detection rate but also a high false-positive rate, while charts that are shifted downward (e.g. Intergrowth-21st) would have a lower false-positive rate but would also have a lower detection rate for SGA fetuses at risk of adverse outcomes (Figure 4). Finding the chart that provides the best balance between these two measures requires careful consideration and should be based on a clear definition of the goals of screening.

6.5 | **How to assess fetal growth in twin gestations**

Twin fetuses grow more slowly than singletons, starting from 28-32 weeks of gestation onward.²²²⁻²²⁵ At term, approximately 30%–50% of twins would be identified as SGA (EFW <10th percentile) using singleton growth standards.^{223,226,227} The mechanisms underlying the relative smallness of twins remain unclear. While some believe that this represents a pathological phenomenon due to failure of the uteroplacental circulation to meet the demands of two fetuses (i.e. twins are more likely to be growth restricted due to the same mechanism responsible for FGR in singletons), ^{224,228-230} others suggest that this represents an early benign physiological adaptation of twins to the "crowded" intrauterine environment in an effort to delay the onset of labor (by decreasing uterine distension) and gain maturation at the expense of size. 231 One important implication of this question relates to the growth standard that should be used in

FIGURE 4 Illustration of the impact of the growth chart chosen on the trade-off between detection rate and false-positive rate of fetuses at risk of adverse outcome. Charts that are shifted upward (light blue dotted line) will have a higher detection rate for pregnancies at risk of adverse outcomes (red circles) but would also have a higher false-positive rate (i.e. identify normal pregnancies [green circle] as being at risk). In contrast, charts that are shifted downward (dark blue solid line) will have a lower false-positive rate (i.e. identify fewer normal pregnancies [green circle] as being at risk) but will also have a lower detection rate for pregnancies at risk of adverse outcomes (red circles). [Colour figure can be viewed at [wileyonlinelibrary.com\]](www.wileyonlinelibrary.com)

twins. If the slower growth of twins represents FGR, it would be reasonable to use singleton growth standards to identify the small twin fetus that, like SGA singletons, may be at increased risk for perinatal mortality and morbidity. However, if the relative smallness of twins is due to a benign adaptive mechanism, it may be preferable to use twin-specific growth charts ^{223,232-235} to avoid overdiagnosis of FGR in twin gestations, $63,227$ which is associated with increased use of resources, ultrasound exams, interventions, and patient anxiety.

Most current guidelines do not provide clear recommendations as to which type of charts should be used to monitor the growth of twins,^{143,236,237} while other guidelines specifically recommend the use of singleton-based charts ²³⁸ or twin-specific charts.²³⁹ As a result, singleton-based standards are used by default in most centers to assess the growth of twins. However, recent data provide support to the hypothesis that the relative smallness of twins is a benign adaptive mechanism and, therefore, for the use of twin-specific charts. For example, several studies suggest that the slower growth of twins is the result of differences in programming that is determined as early as the first trimester..^{229,240-243} In addition, it was found that the use of twin-specific (versus singleton-based) charts was associated with a marked decrease in the rate of twins classified as SGA, without affecting the detection rate of stillbirth, suggesting that twin-specific charts can be used safely.^{227,244,245} Similar findings were reported in studies that investigated the association between the type of chart used (twin versus singleton charts) and other outcomes such as perinatal complications and long-term morbidity.^{246,247} Studies that investigated placental pathology findings reported that SGA twins (based on singleton charts) are less likely

6.6 | **Recommendations**

Society of Ultrasound in Obstetrics and Gynecology (ISUOG).²³⁹ Of note, the diagnosis of FGR in twin gestations should also take into consideration intertwin size discordance, especially in the case of monochorionic placentation.²⁵⁰

FIGO recommends the following for detection of fetal growth restriction (FGR)

 ARELAMED ET AL. ARELAMED ET AL. ARELAMED ET AL. CONSTRETRICS \bigcirc **B** \bigcirc **CONSTRETRICS** \bigcirc **B** \bigcirc **WILEY** \bigcirc **27**

to have placental histopathological evidence of placental insufficiency when compared with SGA singletons.^{248,249} In another recent study on the association between SGA and pre-eclampsia, it was found that in contrast to singletons, the diagnosis of SGA in twins based on singleton charts was not associated with a greater risk of pre-eclampsia, while the association of SGA in twins diagnosed using twin-specific charts had the same magnitude of association with pre-eclampsia to that observed between SGA and pre-eclampsia in singletons.⁶³ Overall, these findings provide support to the hypothesis that the relative smallness of twins is less likely to be the result of placental insufficiency and, thus, less likely to reflect true growth restriction. Based on that, we believe that it seems reasonable to use twin-specific charts for the assessment of fetal growth in twin gestations, as this has the potential to avoid overdiagnosis of FGR and the consequences associated with this diagnosis.²⁴⁶ This approach is supported by the recent guidelines of the International

7 | **WHAT KIND OF INVESTIGATIONS SHOULD BE PERFORMED WHEN FETAL**

GROWTH RESTRICTION IS SUSPECTED?

Once FGR is suspected, a systematic investigation should be performed aimed at identifying the underlying etiology for fetal smallness, with the most important reasons being constitutional SGA, placental dysfunction, and fetal conditions such as genetic or infectious disorders. Establishing the most likely etiology is essential to allow for proper counseling, surveillance, and interventions. The investigation should consist of detailed history, evaluation of screening test results for trisomy 21 and biochemical markers, detailed sonographic assessment for structural anomalies and Doppler studies, and additional testing directed at genetic or infectious etiologies when they are suspected.

7.1 | **Detailed history**

A detailed maternal and family history is essential to correctly identify the etiology of FGR. This should include information on maternal age, racial/ethnic group, height and weight, nutritional status, socioeconomic status, medications, cigarette smoking and use of recreational drugs, chronic medical conditions, personal or family history suggestive of thrombophilia, genetic disorders or consanguinity, obstetric history including birth weight of previous children, and confirmation of pregnancy dating by first-trimester ultrasound.¹⁴³

Advanced maternal age has been associated with FGR, with risk increasing for women over the age of 35 years. $251,252$ Maternal social issues, low income, and domestic violence during pregnancy have been shown to be associated with low birth weight.^{253,254} Poor nutritional status due to conditions such as celiac disease ²⁵⁵ and eating disorders is a potentially treatable cause of FGR.^{256,257} Maternal smoking is an important and potentially modifiable risk factor for FGR.^{258,259}

History should also address the risk of congenital fetal infection with cytomegalovirus, toxoplasmosis, syphilis, Zika virus, and varicella-zoster virus. Relevant questions include a history of febrile disease or rash in pregnancy or the periconceptional period, recent travel history to endemic areas (e.g. for Zika virus), and frequent exposure to young children (cytomegalovirus) or to domestic animals (toxoplasmosis).

Accurate dating of pregnancy is essential for the correct interpretation of estimated fetal size and to avoid a false diagnosis of FGR. Determining gestational age based on menstrual history is often unreliable.^{260,261} Therefore, with the exception of pregnancies achieved by assisted reproductive technology, the crown–rump length measured at the time of first-trimester ultrasound is the most accurate method to date pregnancy, and establishes gestational age with a precision of 5 days in 95% of cases. $262-265$ Crown-rump length is most accurate for the purpose of dating when in the range of 7-60 mm.^{266,267} Therefore, confirmation of gestational age based on first-trimester ultrasound (when available) should be the first step when FGR is suspected. If more than one scan is performed in the first trimester, the earliest scan with a crown–rump length of at least 10 mm should be used. 268

7.2 | **Detailed anatomy scan**

Detailed anatomy scan should be routinely performed when FGR is suspected, especially in cases of early-onset severe FGR. The presence of major structural anomalies, soft sonographic markers, or disorders of amniotic fluid (e.g. polyhydramnios) may raise the possibility of chromosomal, subchromosomal, or single gene abnormalities as the cause of FGR .^{269,270} The presence of very shortened fetal long bones (shorter than –2SD and especially –4SD below the mean) should raise the possibility of skeletal dysplasia and indicates targeted genetic assessment.²⁷¹⁻²⁷³ Attention should also be given to findings that are associated with congenital infections, especially in women with a relevant history, as described above. Examples of such sonographic findings include small head circumference, ventriculomegaly, brain or liver calcifications, periventricular hyperechogenicity, cortical brain malformations, echogenic bowel, hydrops, or placentomegaly.40,274

7.3 | **Doppler studies**

Doppler assessment is an integral part of the diagnostic process and management of FGR. The presence of abnormal Doppler findings in the uterine, umbilical, or middle cerebral arteries is highly suggestive of placental dysfunction as the underlying etiology of FGR. A more detailed description of the different types of Dopplers studies and their application in monitoring and timing of delivery in pregnancies complicated by FGR is provided in section 8 (Management of FGR).

It should be noted that umbilical artery Doppler findings may be normal in the early stages of placental FGR. Therefore, normal umbilical artery Doppler studies do not rule out placental dysfunction, and therefore serial monitoring is recommended in all cases of suspected FGR.^{275,276} At the same time, abnormal umbilical artery Doppler is not pathognomonic of placental dysfunction, as certain genetic conditions (e.g. triploidy) may mimic early-onset placental FGR, including the presence of abnormal umbilical artery Doppler, most likely due to concomitant placental insufficiency secondary to the abnormal placental karyotype. $34,277-279$ In contrast to umbilical artery Doppler, uterine artery Doppler is less likely to be abnormal among fetuses with FGR and abnormal karyotype, and should therefore be considered to be more specific for primary placental FGR, especially in the presence of abnormal angiogenic markers in maternal blood.34,107,280

7.4 | **Additional testing**

Screening for congenital infections should be offered when FGR is suspected, especially in cases of early-onset FGR or when infection is possible based on history of ultrasound findings. Testing should be focused on cytomegalovirus and toxoplasmosis, but may also include rubella, varicella, and syphilis in cases at high risk for these infections. Testing for Zika virus and malaria should also be considered in the relevant travel history or location context. However, it should be noted that interpretation of serology results may be challenging due to limited specificity and cross-reactivity of some of the assays, especially when baseline serology results prior to pregnancy or from early pregnancy are not available.²⁸¹ When fetal infection is highly suspected based on serology results or clinical findings, further testing should be offered by means of amniocentesis for the detection of viral DNA in the amniotic fluid using polymerase chain reaction. In these cases, amniocentesis should be delayed until after 21 weeks of gestation and at least 6–8 weeks following the estimated onset of maternal infection to minimize the risk of falsenegative results.274,282

Genetic consultation and genetic testing by amniocentesis should be offered to women with FGR, especially in cases of early-onset or severe FGR (<3rd percentile), co-presence of sonographic findings (such as structural anomalies, soft markers, or

FIGO recommends the following for investigation of fetal growth restriction (FGR)

7.5 | **Recommendations**

an incremental yield of 4% (95% CI, 1%–6%) over karyotyping: 17 of 376 fetuses with isolated FGR and normal karyotype had significant findings in microarray, most commonly 22q11.2 duplication, Xp22.3 deletion, and 7q11.23 deletion. The incremental yield of microarray over karyotyping was even higher at 10% (95% CI, 6%–14%) in the presence of associated fetal malformations.³⁵ Based on these data, it seems reasonable to offer amniocentesis with karyotype and microarray analysis (when available) to women with FGR, with the decision based on factors such as ultrasound findings, gestational age, lack of evidence of placental dysfunction, and whether the results of the amniocentesis would affect management. These data also suggest that the temptation to substitute amniocentesis by noninvasive prenatal testing (NIPT) using cell-free fetal DNA analysis in this context should be strongly resisted.

Recommendation Quality of evidence Strength of recommendation 1. Women with suspected FGR should undergo systematic assessment that includes the following: (1) detailed history; (2) detailed sonographic assessment for structural anomalies, soft markers, and sonographic signs related to fetal infection; (3) Doppler studies that include at least the umbilical artery and, when available, also the uterine and middle cerebral arteries; and (4) maternal screening for relevant congenital infections, which should be focused on cytomegalovirus and toxoplasmosis, but may also include rubella, herpes, syphilis, malaria, and Zika virus in cases at high risk. **LRS** The extent of investigation may be limited by available resources. Assessment should include screening for infections such as malaria and Zika virus in endemic areas. ⊕⊕⊕⊕ Strong 2. Confirmation of gestational age should be the first step when FGR is suspected. With the exception of pregnancies achieved by assisted reproductive technology, first-trimester crown–rump length is the most accurate method to date pregnancy when in the range of 7–60 mm. If more than one scan is performed in the first trimester, the earliest scan with a crown–rump length of at least 10 mm should be used. **LRS** In low-resource settings, dating may need to be based on menstrual history or symphysis–fundal height. ⊕⊕⊕⊕ Strong 3. Amniocentesis for karyotype (as well as microarray and polymerase chain reaction for infectious agents when available) should be offered to women with suspected FGR, especially in cases with early-onset severe (estimated fetal weight <3rd percentile) FGR, in the presence of sonographic findings associated with genetic or infectious etiologies, no obvious signs of placental dysfunction, and when the findings are likely to affect management. ⊕⊕⊕○ Strong

LRS The availability of genetic testing may be limited by available resources.

polyhydramnios), and the absence of obvious signs of placental dysfunction such as abnormal uterine or umbilical artery Doppler. In addition, women should be counselled about the risk of a genetic etiology even in the presence of "isolated" FGR (i.e. without associated fetal anomalies).^{37,270,283-285} A recent meta-analysis of 10 studies found that in cases of isolated FGR, chromosomal microarray had

\mathbf{WII} \mathbf{FV} $\overline{\text{OINECOLOGN}}$ $\overline{\text{OIV}}$

8 | **MANAGEMENT OF PREGNANCIES WITH FETAL GROWTH RESTRICTION**

Management of pregnancies with FGR depends in part on the results of the investigation described in section 7. In cases of fetal abnormalities (genetic or infectious) the management (expectant versus pregnancy termination) should be individualized based on the nature of the disorder, the expected prognosis, gestational age, parental wishes, and local policies.

The most common underlying etiology of FGR is placental dysfunction. In early-onset FGR (<32 weeks), increased resistance in umbilical artery Doppler is the primary rate-limiting step to subsequent deterioration of cardiovascular and biophysical parameters.²⁸⁶⁻²⁹² The primary management challenge arises from the risk of fetal deterioration and stillbirth in pregnancies undergoing surveillance versus the neonatal morbidity and mortality associated with preterm delivery.^{68,293-297} In late-onset FGR (≥32 weeks), cardiovascular deterioration in response to fetal hypoxia is predominantly confined to the cerebral circulation with little umbilical artery Doppler changes.^{289,291,292,298} Pregnancies complicated by late-onset FGR are major contributors to adverse perinatal outcome attributable to FGR because of misdiagnosis and challenges in detecting deterioration during fetal surveillance.^{299,300}

There is no effective antenatal treatment for placental dysfunction and therefore once FGR has been identified, the principal management steps are institution of fetal surveillance and determination of appropriate thresholds for delivery. Perinatal outcome in early-onset FGR is improved when pregnancies are managed in a high-level fetal medicine and neonatology unit utilizing a uniform management protocol.³⁰¹ Likewise, the optimal management setting for late-onset FGR is a unit that has access and experience in interpretation of surveillance tests, together with an appropriate level neonatal unit. The recommendations for monitoring, timing and mode of delivery, and potential treatments for placentamediated FGR are described below and summarized in Table 1 and Figure 5.

8.1 | **Monitoring**

The primary goal of fetal monitoring is prevention of stillbirth by detection of fetal deterioration that precedes irreversible compromise. To achieve this goal, monitoring tests need to be accurate in identifying fetal risks that favor delivery and, for pregnancies where delivery thresholds are not met, follow-up monitoring needs to be frequent enough to provide a safety net against unanticipated deterioration or stillbirth. Fetal surveillance tests include maternal monitoring of fetal movements, cardiotocography, ultrasound evaluation of amniotic fluid volume and fetal activity, and Doppler ultrasound of the fetal arterial and venous circulations. With progressive compromise, abnormal fetal activity and fetal heart rate patterns are observed, independent of gestational age at diagnosis of FGR.^{288,292,302-304} In contrast, cardiovascular manifestations

of fetal compromise are driven by placental blood flow resistance in the umbilical artery, and therefore differ significantly between early- and late-onset FGR.^{286,289-291,298,299} The surveillance tests that have been evaluated in the management of FGR pregnancies are described below.

8.1.1 | Fetal movement counting

Fetal activity is established from the first trimester onward and as gestational age advances becomes organized into coordinated behavioral states. Progressive fetal hypoxemia is accompanied by a reduction of fetal activity that can be perceived most accurately by the mother when she is lying down and paying focused attention to fetal movements.305 Decreased fetal movement is often defined as less than 10 movements in 2 hours during focused maternal counting.³⁰⁶ Although reports on whether quality improvement tools to promote awareness and management of reduced fetal movements can effectively decrease the risk of stillbirth have been conflicting, most of these interventions were focused on unselected populations rather than in pregnancies with suspected FGR.³⁰⁷⁻³⁰⁹ Given that fetal movement counting is a simple and inexpensive tool that may provide a safety net between scheduled outpatient monitoring visits, it seems reasonable to use movement counting as an adjunct to monitoring in FGR. The mother should be provided with clear behavioral instructions and confirmatory monitoring should be performed for patients presenting with decreased fetal movements.305,308

8.1.2 | Fetal heart rate monitoring

Fetal heart rate monitoring is universally recommended to monitor pregnancies complicated with FGR.^{134,310-312} Antepartum cardiotocography (CTG), also known as a nonstress test (NST), can be performed as a standalone evaluation or in conjunction with measurement of amniotic fluid volume (modified biophysical profile), or a five-component biophysical profile (BBP).

Some heart rate characteristics reflect fetal oxygenation, gestational age, and maturational state of the nervous and cardiovascular systems. The normal heart rate baseline is between 110 and 160 beats per minute (bpm) and decreases with advancing gestation. Periodic accelerations of fetal heart rate (FHR) usually coincide with fetal movements, are observed from the early second trimester, and increase in magnitude and duration with advancing gestation. These are defined as increases in FHR over the baseline of at least 15 bpm and 15 seconds' duration. Two or more of these accelerations define a "reactive" pattern. Recognizing that the frequency of reactivity increases from 50% at 24–28 weeks to 85% at 28–32 weeks of gestation, criteria of greater than or equal to 10 bpm amplitude and greater than 10 seconds' duration are recommended at earlier gestational ages.³¹³⁻³¹⁶ A "nonreactive" FHR pattern is one that does not display accelerations over an observation period of 40 minutes.

TABLE 1 Recommendations for monitoring, timing, and mode of delivery in cases with suspected fetal growth restriction.

Abbreviations: AEDV/REDV, absent or reversed diastolic velocity in the umbilical artery; BPP, biophysical profile; CPR, cerebroplacental ratio; DV, ductus venosus; FGR, fetal growth restriction; MCA, middle cerebral artery; NST, nonstress test; OR, odds ratio; PI, pulsatility index; PIV, pulsatility index for veins; SGA, small for gestational age; UA, umbilical artery; UtA, uterine artery.

^aMonitoring should be based on integration of multiple modalities (Doppler, BPP, NST).

 $^{\rm b}$ Absolute indications for delivery at any gestational age and birth weight combination that are considered to be viable include: BPP or NST abnormalities or severe pre-eclampsia with uncontrolled hypertension or end-organ damage (section 8.2.3). In addition, timing of delivery should be individualized based on factors such as parental decision regarding threshold for intervention.

c There is lack of evidence on the appropriate test to predict the risk of fetal deterioration and on the optimal monitoring strategy in cases of uncomplicated SGA fetuses, especially at term. Given this, there are differences in practice in various regions of the world regarding use of BPP/NST for fetal monitoring in this context, and some of the authors of these guidelines do not use BPP or NST for monitoring of fetuses with uncomplicated SGA as long as Doppler studies are normal. We suggest that the decision regarding use of BPP/NST should be based on local practices, the risk profile of the local population, and the available resources in each particular setting.

 $^{\rm d}$ Timing should be individualized based on local neonatal outcomes. Before 26 weeks, careful and shared decision making with the parents and neonatology team is recommended.

In addition to reactivity, FHR patterns display "variability"—the average oscillations in the FHR signal, evaluated in bpm in 1-minute windows. Reduced variability appears later than absent reactivity in the process of progressive fetal hypoxia. It reflects reduced sympathetic–parasympathetic activity, secondary to diminished brainstem oxygenation.

The FHR pattern reflects fetal oxygenation and acid-base status at the time of evaluation but does not predict deterioration

FIGURE 5 Delivery criteria for fetal growth restriction. Delivery criteria are based on monitoring with umbilical artery, ductus venosus, and middle cerebral artery Doppler at specified gestational ages with traditional nonstress testing or computerized CTG (cCTG) if available. Abbreviations: NICU, neonatal intensive care unit; FHR, fetal heart rate; CTG, cardiotocogram; STV, short-term variation; ms, milliseconds; EFW, estimated fetal weight; PI, pulsatility index. [Colour figure can be viewed at [wileyonlinelibrary.com](www.wileyonlinelibrary.com)]

in FGR. In unselected pregnancies the rate of stillbirth in the week following a reactive CTG/NST is 1.9/1000 (negative predictive rate of 99.8%).³¹⁷ Without any additional information, the empirically recommended minimum frequency is twice weekly CTG/NST. The frequency may be increased when evaluation of amniotic fluid or Doppler parameters indicate a more advanced degree of fetal compromise and delivery criteria have not yet been met. A nonreactive CTG/NST has low specificity for hypoxia and requires additional tests to determine fetal status and distinguish FHR pattern variations caused by fetal behavior, while reduced variability is a much stronger predictor of central nervous system hypoxia.

8.1.3 | Computerized fetal heart rate monitoring

Some professional societies recommend computerized fetal heart rate monitoring (cCTG) as the preferred modality to analyze CTG/ NST tracings.134,308 Inconsistency in visual assessment, particularly FHR variability, is a major contributor to interobserver variations in interpretation of CTG/NST tracings.³¹⁸ cCTG evaluates FHR parameters such as baseline, accelerations, decelerations, and variability in an objective and quantifiable way. The Sonicaid cCTG system (Huntleigh Healthcare, Cardiff, UK) provides the parameter "shortterm variation" (STV) in milliseconds, while others quantify variability in a more traditional way in ppm .³¹⁹ In contrast to visual FHR analysis, cCTG decreases observer variability and allows longitudinal numerical analysis of variability.³²⁰

FHR variability increases with gestational age; after 29 weeks of gestation, below 4.0 ms or below 3.0 ms meet criteria for reduced or very low STV, respectively.³²¹ Before 29 weeks of gestation, STV below 3.5 ms is considered reduced, and below 2.6 ms is considered very low. STV below 3 ms has a 77% positive predictive value for fetal acidemia.322,323

Similar to CTG/NST, cCTG does not predict fetal deterioration. In early-onset FGR the daily risks for abnormal STV are 4%–5% but are unpredictable by additional monitoring tests. Accordingly, CTG/NST or cCTG monitoring needs to be performed more frequently than Doppler assessments. In patients receiving inpatient monitoring, a minimum frequency of daily cCTG/CTG/NST is recommended.³²⁰

8.1.4 | Ultrasound measurement of amniotic fluid volume

Professional societies do not recommend inclusion of isolated amniotic fluid volume assessment into management decisions for FGR. A decrease in amniotic fluid volume can occur as a result of fetal oliguria in response to progressive placental dysfunction and hypoxia, as well as rupture of membranes.^{287,288,324} Accordingly. additional evaluation is required to determine the significance of decreased amniotic fluid volume. Oligohydramnios can be defined as an ultrasound measured four-quadrant amniotic fluid index below or equal to 5 cm, or a maximum vertical amniotic fluid pocket below or equal to 2 cm.³²⁵ Use of the latter reduces overdiagnosis of oligohydramnios and is preferred. Oligohydramnios is associated with an increased rate of intrapartum FHR abnormalities, need for cesarean section, and low 5-minute Apgar scores, but not acidosis at birth.³²⁶

8.1.5 | Biophysical profile scoring

Biophysical profile (BPP) scoring is not universally recommended as the primary surveillance tool for FGR and is predominantly utilized in Canada and North America where the concept was first developed for fetal surveillance in the later part of the third trimester. The modified BPP refers to combined use of the CTG/NST as a shortterm indicator of fetal acid-base balance and the maximum amniotic fluid pocket as an indicator of long-term placental function. 327 The five-component BPP comprises fetal breathing movements, gross body movements, and tone, in addition to CTG/NST and maximum amniotic fluid pocket, and therefore includes four indicators of short-term acid-base balance.³⁰¹

The modified BPP is considered abnormal when either the CTG/ NST is nonreactive, or the maximum amniotic fluid pocket is below 2 cm. The most common reason for an abnormal modified BPP is a nonreactive CTG/NST, requiring additional ultrasound observation to complete a five-component BPP and determine fetal acid-base balance. The BPP is scored over a 30-minute ultrasound observation period of the fetus. Fetal breathing movements are considered present if one or more episodes of 30 seconds of breathing or hiccups are observed. Fetal body movement is present when three or more discrete body or limb movements are observed. Fetal tone is present when one or more episodes of extension and flexion of the fetal extremities are observed. Each component of the BPP receives a score of 2 for its presence and 0 for its absence. Scores of 8–10, 6, and 4 or less are considered normal, equivocal, and abnormal, respectively.

In unselected pregnancies, the rate of stillbirth in the week following a normal modified or five-component BPP is 0.8/1000 (negative predictive rate >99.9%). FGR fetuses show a sequential loss of heart rate reactivity, breathing movements, gross body movement, and tone with decrease in pH. 287,288,301,302 In FGR pregnancies, an abnormal BPP (score of 4 or less) is associated with an umbilical artery pH of less than 7.20, with sensitivity increasing to 100% at a score of 0/10.^{301,302,319}

The BPP is a more accurate predictor of fetal acid-base status at the time of testing than CTG/NST, with a similar accuracy as cCTG. Therefore, a five-component BPP can be used to clarify fetal acidbase status when a nonreactive CTG/NST is obtained. The frequency of BPP testing is guided by the same principles as timing of fetal heart rate testing.

8.1.6 | Umbilical artery Doppler

Umbilical artery Doppler is universally recommended for monitoring of FGR because it assesses the hemodynamic aspect of placental dysfunction.134,143,308,310 It is estimated that approximately one-third of the villous circulation needs to be damaged before a decrease in umbilical artery end-diastolic velocity occurs. Absent or reversed umbilical artery end-diastolic velocity corresponds to malperfusion of 50%-70% of the villous vascular tree.³²⁸ Because elevated villous blood flow resistance is predominantly associated with the placental pathology found in early-onset FGR, umbilical artery Doppler does not reliably predict outcome in late-onset FGR.³²⁹⁻³³¹

The umbilical artery Doppler waveform can be quantified using the pulsatility index, or by visual classification of end-diastolic velocity as absent (AEDV) or reversed (REDV). With increasing degrees of placental blood flow resistance, an abnormal umbilical artery waveform is defined as either having an elevated pulsatility index, AEDV, or REDV. The degree of placental blood flow resistance elevation is the primary factor determining the rate of clinical progression and the associated risk for fetal deterioration and stillbirth in early-onset-FGR.^{286,289,291,292} When the umbilical artery pulsatility index is elevated but end-diastolic forward flow is still present, the median time interval to additional surveillance abnormalities is 2 weeks. Once AEDV occurs, cardiovascular deterioration advances after a median of 5 days and the weighted odds ratio for stillbirth is 3.6 (2.3–5.6).286,291,332 When REDV occurs, the median interval for further fetal deterioration is 2 days and the weighted odds ratio for stillbirth is 7.3 (4.6-11.4).^{291,331}

In patients with normal umbilical artery Doppler, the recommended frequency to repeat Doppler monitoring ranges from weekly to every other week. However, when AEDV develops, Doppler surveillance is recommended at minimum twice weekly, and for REDV at least three times weekly unless delivery criteria have been met.

8.1.7 | Cerebral artery Doppler

The majority of professional societies now recommend middle cerebral artery Doppler for monitoring in late-onset FGR. Concurrent measurement of the umbilical artery and middle cerebral artery pulsatility index allows calculation of the cerebroplacental Doppler ratio. Both the cerebroplacental ratio and middle cerebral artery pulsatility index decrease as a hemodynamic response to fetal hypoxemia and therefore reflect placental dysfunction, even in those pregnancies where the villous blood flow resistance is not **34 WII FV-CANECOLOGY** (W)

elevated enough to produce an abnormal umbilical artery pulsatility index. Approximately 20% of term SGA fetuses with normal umbilical artery Doppler have a decreased middle cerebral artery pulsatility index, which is associated with a higher rate of cesarean section for intrapartum distress, poor neonatal transition, and adverse developmental outcome.333-335 The cerebroplacental Doppler ratio is more closely related to fetal hypoxia than its individual components,³³⁶ but has a similar predictive accuracy for perinatal death, fetal distress, or poor neonatal transition as the umbilical artery pulsatility index.³³⁷

Cardiovascular deterioration in late-onset FGR is characterized by abnormal cerebral artery Doppler. Therefore, an important role of middle cerebral artery Doppler is to provide an estimate of perinatal risk in patients with normal umbilical artery Doppler.^{292,331} Because of the higher risk for adverse outcome within 1 week of a decrease in middle cerebral artery pulsatility index, it is recommended to utilize at least twice weekly surveillance in this setting.

8.1.8 | Ductus venosus Doppler

The few professional societies that recommend ductus venosus Doppler evaluation specify that it should be performed in specialized centers that have expertise in the comprehensive perinatal management of early-onset FGR.³¹² The relative forward flow in atrial systole in the ductus venosus decreases with worsening placental function or reduced fetal cardiac function, leading to an increase in the pulsatility index for veins, absent, or reversal of the a-wave.286,288,291,292,338

Abnormal ductus venosus Doppler is primarily observed in early-onset FGR and can provide an estimate of fetal acid-base balance and the risk of stillbirth. The odds ratio of absent or reversed atrial systolic velocity for an umbilical artery pH less than 7.20 at birth is 4.4 $(1.2-17.2)$ ^{339,340} The weighted odds ratio of absent or reversed ductus venosus atrial systolic velocity for fetal death is 11.6 $(6.3 - 19.7).$ ³³¹

Abnormal ductus venosus Doppler also predicts fetal decompensation to an abnormal BPP, reduced variability on cCTG, or stillbirth. In fetuses with elevated ductus venosus pulsatility index for veins but forward flow during atrial systole, the median interval to progressive venous Doppler deterioration can be as short as 2 days. 291 In patients that do not yet meet delivery criteria, ductus venosus Doppler is recommended at minimum twice weekly in patients with AEDV and three times weekly when REDV is observed.286,291,292,341 When ductus venosus Doppler indices increase as a new finding, the frequency of monitoring needs to be increased further.

8.1.9 | Surveillance strategy

Monitoring in FGR pregnancies is intended to prevent fetal compromise or stillbirth, and the choice of tests and their timing is

heavily influenced by gestational age. A robust plan is essential, since expectant management with ongoing monitoring, particularly in the setting of early-onset FGR, can result in a three to five-fold increased stillbirth rate when compared with immediate delivery, depending on the degree of cardiovascular compromise that is tolerated before triggering delivery.^{294,342,343} The optimal monitoring frequency in FGR has not been determined due to the varying circumstances of gestational age and severity of FGR. A combination of surveillance modalities is needed to accurately determine fetal acidbase status at the time of testing, as well as allowing anticipation of future deterioration.289-292,298,344 The accurate prediction of fetal acid-base status is required to prevent unnecessary intervention and nonindicated delivery. The anticipation of deterioration informs subsequent monitoring intervals that provide a safety net against unanticipated fetal acidosis and asphyxia. The combination of biophysical (CTG/NST, cCTG, BPP) and cardiovascular parameters (umbilical artery, middle cerebral artery, and ductus venosus Doppler) is considered a robust approach for FGR surveillance. Among these modalities, the combination of CTG/NST and umbilical artery Doppler is universally recommended.

There is good evidence that umbilical artery Doppler offers sufficient information to determine monitoring frequency in earlyonset FGR. Although middle cerebral artery Doppler could provide additional information in those late-onset FGR pregnancies with normal umbilical artery Doppler, this practice has not been evaluated.292,328,329 Based on observational data in term FGR with normal umbilical artery Doppler, the approximate interval to stillbirth in patients with abnormal middle cerebral artery Doppler is 4 days, suggesting the need for twice weekly CTG/NST monitoring. In the absence of further evidence on the clinical benefit of middle cerebral artery Doppler, twice weekly CTG/NST monitoring in FGR after 32 weeks of gestation in patients with normal umbilical artery Doppler provides the same safety net (Table 1).

When umbilical artery pulsatility index is elevated, weekly Doppler is suggested, and when there is AEDV or REDV, more frequent assessment is recommended (Table 1). In early-onset FGR with AEDV or REDV, the risk of stillbirth increases when the ductus venosus Doppler or the CTG/NST patterns become abnormal.^{292,319,340} However, there is currently no evidence that adjusting the timing of monitoring based on ductus venosus Doppler improves outcome. In patients with AEDV the stillbirth rate is 0%–1% when at least once daily CTG/NST, cCTG, or BPP is performed with predefined delivery criteria.^{341,342} When monitoring is continued to allow for an increase in the ductus venosus pulsatility index for veins, the stillbirth rate is 2%, and 11% of deliveries occur for abnormal STV, 19% for an abnormal BPP, and 22% for FHR decelerations.^{304,342} When monitoring is continued in anticipation of reversal of the ductus venosus a-wave velocity, the stillbirth rate is 4%, and 20% of deliveries occur for abnormal STV, 29% for an abnormal BPP, and 31% for FHR decelerations (Table 1).^{342,345} This indicates that with ongoing monitoring the risk of FHR abnormalities or an abnormal BPP requiring delivery cannot be predicted by the ductus venosus Doppler.^{319,346} Based on the regional pattern of practice, this indicates that in patients who **ARELAMED ET AL. AND RELAMED ET AL. CONDUCT AND RELAMED ET AL. CONDUCT AND RELAY AND**

are admitted for AEDV, the minimum frequency of CTG/NST or BPP should be daily and more frequent with REDV (Table 1).³¹⁹

8.2 | **Timing of delivery**

The timing of delivery in FGR is determined by gestational age, severity of FGR, findings of fetal monitoring tests, and maternal factors such as pre-eclampsia (Table 1 and Figure 5). Delivery indications can be considered as absolute if they are independent of gestational age, and relative if the threshold to deliver based on the surveillance findings varies across gestational age.

8.2.1 | Gestational age-related risks in fetal growth restriction

With advancing gestational age there are several important changes in the relative risks of delivery versus ongoing surveillance that define the delivery thresholds.

From 24–28 weeks of gestation each day of pregnancy prolongation results in an estimated 2% decrease in neonatal death, as well as major neonatal complications including bronchopulmonary dysplasia, high-grade intraventricular hemorrhage, and surgical necrotizing enterocolitis. The impact of prematurity, neonatal weight below 500 g, challenging resuscitation, and decreased tolerance for low Apgar scores results in average neonatal survival rates below 50% and intact survival below 50% until 26 weeks. 68,298,345,346

Between 28 and 30 weeks of gestation the daily increment in survival is approximately 0.7%. After 30 weeks, neonatal survival rates exceed 90%, $68,296,297$ and there is a significant decrease in major neonatal complications from approximately 35% at 30 weeks to less than 10% at 34 weeks, as well as a decrease in the risk of neurodevelopmental delay for neonates delivered after this time. FGR infants delivered prior to 30 weeks have a three-fold higher rate of developmental abnormalities and an up to eight-fold increased rate of cerebral palsy.10,295

From 34–38 weeks of gestation neonates are more likely to require admission to the intensive care nursery but have reduced risks of major neonatal complications.^{347,348} In SGA fetuses that remain undelivered after 38 weeks, the risk of stillbirth doubles every week and reaches 60/10 000 for pregnancies that continue beyond the due date. 349,350

8.2.2 | Gestational age-related management strategy

The balance between fetal and neonatal risks defines the predominant management strategy at different gestational epochs. Accordingly, the goal of management shifts from gaining fetal viability at 26 weeks to a graded improvement in survival, neonatal morbidity,

and neurodevelopment by delaying delivery until 34–36 weeks. The increase in stillbirth rate in undelivered fetuses increasingly favors delivery from 36 weeks onward.

Timing of delivery in FGR has been evaluated in three randomized trials. The growth restriction intervention trial (GRIT) randomized pregnancies that had abnormal fetal biometry and umbilical artery Doppler studies performed as part of clinical management into immediate delivery after completion of a course of steroids versus delivery when the managing physician was no longer comfortable with conservative management.^{294,295} The monitoring protocol and delivery criteria were not specified. The trial demonstrated that, in the absence of specific criteria, either management approach resulted in the same perinatal outcome. Delaying delivery increased the risk of stillbirth, while earlier delivery resulted in a higher degree of prematurity-related complications that either led to neonatal death or an increased risk of developmental delay.

The disproportionate intrauterine growth intervention trial at term (DIGITAT) randomized SGA fetuses by several biometry criteria, independent of the umbilical artery Doppler pattern, to induction or expectant monitoring between 36 and 41 weeks of gestation.³⁴⁷ The study demonstrated that, while elective induction did not affect neonatal or obstetric outcomes, deliveries prior to 38 weeks resulted in a higher rate of admissions to the neonatal intensive care unit.

These trials demonstrate that the relative risk for neonatal complications requires definitive delivery indications until 38 weeks of gestation. After that time, delivery for indication of FGR is likely to prevent stillbirth in ongoing pregnancies. The continuous decrease in neonatal risks requires that delivery indications at early gestational ages occur at a higher threshold for fetal risks than after 30–32 weeks.

8.2.3 | Absolute delivery criteria for fetal growth restriction (independent of gestational age)

Absolute delivery criteria are findings associated with important health risks to the mother or fetus, and therefore require delivery without consideration of gestational age (Figure 5).

The fetal biophysical variables are strongly influenced by oxygen tension in the regulatory centers. A 30-minute BBP score of 0 or 2, or a 60-minute score of 4 indicates a prelabor fetal pH of less than 7.20 and requires delivery to prevent fetal demise.

Repetitive FHR decelerations, a sinusoidal heart rate, absent variability with recurrent late decelerations, or bradycardia predict fetal acidemia and poor perinatal outcome and require delivery if the causative stimulus cannot be removed. When cCTG is used, a shortterm variation below 2.6 ms is below the 5th percentile irrespective of gestational age and requires delivery for its strong association with fetal acidemia.

Maternal pre-eclampsia with severe features complicates up to 30% of FGR pregnancies, with a higher proportion in early-onset FGR. In the absence of effective treatment other than delivery, pre-eclampsia with uncontrolled severe hypertension, HELLP **36 198 • 36 • CENTER** COLOGY **36 • CENTER COLOGY CONTROL SCIENCE CONTROL SCIENCE CONTROL SCIENCE CONTROL CONTR**

syndrome (hemolysis, elevated liver enzyme levels, and low platelets), or other evidence of end-organ damage (e.g. oliguria or acute renal injury other than proteinuria, pulmonary edema, or eclampsia) requires delivery (Figure 5).

8.2.4 | Relative delivery criteria for fetal growth restriction (adjusted for gestational age)

The trial of umbilical and fetal flow in Europe (TRUFFLE) evaluated two monitoring strategies and specific delivery criteria in earlyonset FGR, with survival without neurodevelopmental impairment at 2 years of age as the primary outcome.³⁴³ Monitoring with cCTG and umbilical artery Doppler was universal in all patients, while ductus venosus Doppler was only added in two study arms. Patients were randomized to one of three specific delivery criteria: (1) abnormal cCTG STV; (2) mild ductus venosus abnormalities; and (3) severe ductus venosus abnormalities with absence or reversal of the a-wave. Because patients with ductus venosus Doppler monitoring also had FHR monitoring, safety net delivery criteria based on cCTG were also applied in these groups. These included STV below 2.6 ms irrespective of gestational age and below 3.0 ms from 29 weeks onward. In addition, umbilical artery Doppler findings were utilized as relative delivery criteria from 30 weeks onward for REDV and 32 weeks onward for AEDV. The choice of these thresholds is supported by a recent meta-analysis that found that in undelivered FGR pregnancies, umbilical artery REDV has a 19% stillbirth rate, which exceeds mortality for neonates delivered from 30 weeks onward, while AEDV carries a 6.8% stillbirth risk, which favors delivery due to lower neonatal mortality from 32 weeks onward (Table 1 and Figure 1).³³²

The TRUFFLE study demonstrated that a predefined management strategy produces better outcomes than expected in all FGR pregnancies.³⁴³ The primary endpoint was less frequently observed in patients randomized to deliver for late ductus venosus abnormality. Overall, cCTG was the most frequent trigger for delivery. In the three arms, delivery was based on abnormal STV in 11%–51% of participants, and visually apparent FHR decelerations led to delivery in 22%–31% of participants. While the strategy of awaiting absent or reversed ductus venosus a-wave to determine delivery produced the better study outcome, it is noteworthy that the stillbirth rate increased four-fold compared with patients who were monitored with cCTG and umbilical artery Doppler. In addition, an absent ductus venosus a-wave only triggered delivery in 10% of participants in this study arm. The frequency of delivery decisions based on FHR abnormalities emphasizes the importance of concurrently monitoring growth-restricted fetuses with more than one modality.

Because cCTG is not universally available, most healthcare providers need to rely on traditional CTG/NST monitoring. While BPP has not been studied in randomized intervention trials in FGR, it is an established monitoring tool to verify fetal status in patients with a nonreactive tracing. In FGR, an abnormal BPP predicts abnormal arterial pH with a similar accuracy to cCTG and is an independent delivery trigger at a comparable frequency to cCTG.^{304,319}

Therefore, it is recommended that in FGR fetuses with nonreassuring CTG/NST not yet meeting the criteria for delivery, a BPP is completed to establish fetal status. If the expertise is not available to perform a BPP, prolongation of the CTG/NST or increase in testing frequency may be required to determine if delivery is necessary.

Optimal delivery criteria for FGR presenting after 32 weeks of gestation have not been evaluated in a randomized trial and are based on expert consensus. Table 1 and Figure 5 summarize the management approaches and recommendations for delivery. When local neonatal outcomes are consistently more favorable for FGR neonates, relative delivery indications may be applied at earlier gestational ages than indicated. For example, improved neonatal survival may justify delivery for REDV from 30 weeks onward.

8.3 | **Mode of delivery and intrapartum considerations**

FGR in itself is not an indication for cesarean section. However, primary cesarean section may be considered in selected cases of severe FGR where the likelihood of successful vaginal delivery is low.

Fetuses with placenta-mediated FGR are less likely to tolerate the stress associated with labor and are at increased risk of requiring intrapartum urgent cesarean section for nonreassuring FHR tracing. Therefore, in certain cases of FGR, a trial of labor is highly unlikely to be successful and might be associated with fetal risks to the extent that a primary cesarean section should be preferred. This depends on multiple factors including gestational age, severity of FGR, Doppler changes, associated pre-eclampsia, parity, cervical Bishop score, and patient preference (Table 1).

In cases of early-onset FGR the main goal is to prolong pregnancy and maximize fetal maturation by means of expectant management under close monitoring until there is evidence of late Doppler changes in the umbilical artery (AEDV or REDV), ductus venosus alterations, or FHR abnormalities. Therefore, at the point when delivery is indicated in cases of severe early-onset FGR, the fetus might already be experiencing some degree of hypoxia or acidosis,291 in which case the likelihood of the fetus tolerating labor is low and the rate of cesarean section has been reported to be greater than 80%.³⁵¹ In addition, labor induction in general is less likely to be successful during the preterm period.^{352,353} For these reasons, primary cesarean section is usually the preferred option when delivery is indicated in cases of severe early-onset FGR. 354

In contrast, late-onset FGR is usually less severe and fetal hypoxia or acidosis is less likely to be present at the time when delivery is indicated. Indeed, in the DIGITAT trial the rate of vaginal delivery was greater than 80% in pregnancies induced for SGA with normal umbilical artery Doppler after 36 weeks of gestation.³⁴⁷ This observation suggests that most term SGA fetuses with normal umbilical artery Doppler can tolerate labor and that the presence of late-onset FGR in the absence of additional factors does not preclude induction of labor. Several studies have tried to individualize the decision regarding mode of delivery through the development of models for the prediction of urgent cesarean section in women with late-onset SGA undergoing labor induction. The factors that were most predictive of urgent cesarean section were gestational age, severity of SGA (EFW <3rd percentile), cerebral Doppler (middle cerebral artery and cerebroplacental ratio), and Bishop score.^{355,356} For example. in a large cohort study of 509 women undergoing labor induction for late-onset SGA, the predictive model had a positive predictive value of 36% and a negative predictive value of 89% for urgent cesarean section for nonreassuring fetal state.³⁵⁵ Thus, although this information can be helpful for patient counselling regarding mode of delivery and may reassure women with none of these risk factors of the high likelihood of a successful trial of labor (nearly 90%), the positive predictive value of these models (i.e. a risk of cesarean section in the range of 30%–40%) is not high enough to preclude a trial of labor even when these risk factors are present.

The optimal approach for cervical ripening in women undergoing induction for FGR remains unclear. In a recent meta-analysis of 12 trials on cervical ripening in pregnancies complicated by SGA or FGR, the authors concluded that mechanical methods (such as balloon catheters) seem to be associated with a lower risk of cesarean section and intrapartum complications compared with alternatives such as dinoprostone.³⁵⁷ Given these data, it seems reasonable to prefer balloon catheter over prostaglandin preparations, when possible, for cervical ripening in pregnancies with suspected FGR. If prostaglandin agents are used, a reversible method (e.g. dinoprostone vaginal insert) should be preferred.

During labor, continuous FHR monitoring is recommended. Delivery should take place at an institution with the appropriate level of neonatal care for the gestational age and the anticipated management needs of the neonate.

It is recommended that the placenta is sent for histopathological evaluation after delivery. Ideally this should be done in accordance with the Amsterdam workshop consensus statement. ³⁵⁸ Highquality evaluation of the placental pathology is not only likely to increase the precision of the diagnosis but also provides information on the risks of recurrence.18,359,360

8.4 | **Medical interventions**

8.4.1 | Antenatal corticosteroids

The efficacy of antenatal corticosteroids in cases of FGR has been questioned, based on reports of elevated endogenous cortisol levels in this population when compared with normally grown fetuses. $361-364$ In addition, the unique cardiovascular, hormonal, and metabolic changes characteristic of growth-restricted fetuses^{276,365-369} have raised concerns that exposure to exogenous steroids may produce potentially harmful cardiovascular and metabolic effects in these already compromised fetuses. Indeed, exposure to corticosteroids has been shown to result in Doppler changes in growth-restricted fetuses such as transient increase in diastolic flow in the umbilical artery $370-373$ and the middle cerebral artery, $374-376$ which have been

attributed to peripheral vasodilatation or an increase in cardiac output and circulatory stress.^{376,377} Despite this, recent data support the efficacy and safety of antenatal corticosteroids in the subgroup of SGA fetuses, 378,379 which should be administered when delivery is anticipated, ideally within 1-7 days before birth.³⁸⁰ When administered in cases of severe FGR with late Doppler changes, an inpatient setting is advised where the fetus can be closely monitored. Finally, it is important to recognize that the "improvement" in umbilical artery Doppler that is often seen following administration of antenatal corticosteroids is transient, and is thought to be the result of vasodilation of the fetoplacental arterial tree and increased fetal cardiac output rather than a true decrease in placental resistance.³⁸¹ Therefore, these transient changes should not be interpreted as an improvement in fetal status and should not affect the management plan. Of note, the absence of any change in end-diastolic flow in response to antenatal corticosteroids is a concern and predicts subsequent fetal deterioration.³⁷²

8.4.2 | Magnesium sulfate for neuroprotection

Administration of magnesium sulfate to women at risk of preterm birth has been shown to have a neuroprotective role, with a decrease in the risk of perinatal mortality, cerebral palsy, and gross motor dysfunction.382,383 Possible mechanisms thought to be involved in the beneficial effects of magnesium sulfate include reducing intracellular calcium levels, stabilizing blood pressure, normalizing cerebral blood flow, blocking the effects of excitatory neurotransmitters such as glutamate, and antioxidant and anti-inflammatory effects.384,385 However, the optimal protocol for the administration of magnesium sulfate for the purpose of neuroprotection remains unclear and available protocols vary with regard to the timing of administration, upper gestational age limit, dose, duration, and need for repeat doses. 386-389

The observation that term FGR infants have higher cord blood magnesium levels compared with normally grown infants raises the theoretical concern that maternal administration of magnesium sulfate in cases of FGR might result in toxic magnesium levels in the fetus.390,391 However, there are currently no data on the efficacy and safety of magnesium sulfate in FGR fetuses that can support or refute these theoretical concerns. Therefore, there is currently no evidence in favor or against recommending administration of magnesium sulfate for neuroprotection in women at risk of preterm birth with suspected FGR.379 We believe that, at the current time, it is reasonable to extrapolate the efficacy of magnesium sulfate to specific subgroups of pregnancies, including those complicated by FGR, especially given that FGR is an independent risk factor for cerebral palsy.

8.4.3 | Treatments under investigation

Several novel therapies aiming to improve poor placentation and uterine blood flow are being explored, some of which are described **38 | WILEY-** CINECOLOGY (W) **AND RELATED ET AL. CONSTETNIES**

below. However, there are currently no proven treatments for FGR, and any of the therapies currently under investigation should be evaluated only in an appropriately regulated research setting.³⁹²

Phosphodiesterase type-5 inhibitors, such as sildenafil citrate, potentiate nitric oxide availability, lead to vasodilatation, 393,394 and can improve umbilical artery and middle cerebral artery Doppler.³⁹⁵ However, in the recently published STRIDER trial, which randomized 135 women with early-onset FGR to 25 mg sildenafil three times daily or placebo, sildenafil did not prolong pregnancy or improve pregnancy outcomes.³⁹⁶ More recently, a similar randomized trial was halted prematurely due to lack of benefit along with concerns that sildenafil may cause neonatal pulmonary hypertension.³⁹⁷

Another approach is to target the uteroplacental circulation with maternal vascular endothelial growth factor gene therapy, thereby improving local vasodilatation and angiogenesis. 392 Clinically, vector

delivery into the uterine arteries can be achieved with intervention radiology. This approach is currently being investigated in the ongoing EVERREST trial.398 Protein pump inhibitors have been shown in vitro to decrease sFlt-1 and soluble endoglin and improve markers of endothelial dysfunction. However, in a recent randomized trial involving 120 women with preterm pre-eclampsia, esomeprazole did not improve pregnancy outcomes.³⁹⁹ Pravastatin has been shown to have anti-inflammatory, antioxidant, and proangiogenic properties.^{400,401} However, in a recently published randomized trial of 94 women with early-onset pre-eclampsia, the administration of 40 mg pravastatin daily did not lower maternal sFlt-1 levels or prolong pregnancy when compared with placebo.⁴⁰² Other novel potential therapies include nanoparticles and microRNAs that deliver drugs locally to the uterine arterial endothelium or trophoblasts, to improve uterine blood flow and placental function.

8.5 | **Recommendations**

FIGO recommends the following for the management of fetal growth restriction (FGR)

16. There are currently no proven treatments for FGR. ● example and provide the control of example and example of the strong

9 | **POSTPARTUM A SSESSMENT AND COUNSELLING FOR FUTURE PREGNANCIES**

9.1 | **Infant follow-up**

Growth-restricted infants are at increased risk of both immediate and long-term complications, and therefore require closer follow-up than normally grown infants in the first years of life.

Growth-restricted infants have lower survival rates compared with those appropriate for gestational age.⁴⁰³ Although this may be attributed in part to prematurity that is often associated with FGR, birth weight has been shown to be an independent prognostic factor for neonatal mortality, irrespective of gestational age.⁴⁰⁴ In a population-based cohort study, the mortality rate of term FGR neonates was approximately five-fold higher compared with appropriate for gestational age neonates (0.3% vs 0.06%).⁴⁰⁵

FGR can affect postnatal growth. In cases of mild FGR, infants tend to achieve normal height during the first year of life.406 In cases affected by severe FGR, however, height in the late teens is lower than those born appropriate for gestational age (169.9 ± 1.5 vs 175.4 ± 0.8 cm; *P* < 0.0001 for boys; and 159.4 \pm 1.3 vs 163.1 \pm 0.8 cm; *P* < 0.0005 for girls).⁴⁰⁷

FGR infants are also at increased risk of adverse long-term neurodevelopmental outcomes. A systematic review of this topic found that FGR infants are at higher risk of poor neurodevelopmental outcomes measured up to 3 years of age; however, high levels of heterogeneity in primary outcomes were reported in the studies included in the review. 12 Of note, adverse neurodevelopmental outcomes may at least partly be related to coexisting increased prematurity rates.⁴⁰⁸

In line with the developmental origins of health and disease hypothesis, FGR has been associated, in both animal and human studies, with an increased risk of future noncommunicable diseases including obesity, diabetes, hypertension, and cardiovascular disease.⁴⁰⁹⁻⁴¹¹ The risk is especially high in those infants who experience rapid catch-up growth in the first few years of life.^{412,413} The mechanisms underlying these associations are not entirely clear. However, fetal programming by means of epigenetic changes as well as direct organ damage are thought to play a role.⁴¹⁴ Ongoing studies are investigating the optimal follow-up and prevention strategies to decrease the risk of these complications. 415,416

9.2 | **Maternal follow-up**

It is well established that women with a history of pregnancy complicated by FGR or other placenta-mediated complications such as pre-eclampsia are at an increased risk of future cardiovascular disease, especially in the presence of early-onset disease. In a population-based study that included more than 100 000 pregnancies and provided maternal follow-up for 15–19 years, delivery of a low birth weight infant was associated with increased maternal risk of ischemic

heart disease or death (aHR 1.9; 95% CI, 1.5-2.4).⁴¹⁷ Moreover, a combination of FGR, pre-eclampsia, and preterm delivery amplified the risk of disease seven-fold. For a detailed review of the evidence supporting these associations, their underlying mechanisms, and recommendations on maternal follow-up and prevention strategies, please refer to the recently published FIGO postpregnancy initiative on long-term maternal implications of pregnancy complications and follow-up considerations.⁴

9.3 | **Counselling regarding future pregnancies**

The most frequent and relevant question that care providers are being asked by couples whose prior pregnancy was complicated by FGR relates to the likelihood of a similar complication in subsequent pregnancies. The answer to this question is often difficult and depends on several factors, namely the underlying etiology, severity and timing of onset, and the presence or absence of modifiable risk factors (e.g. maternal medical conditions or smoking). In cases of placenta-mediated FGR, the results of the placental histopathological examination may provide valuable information that can assist care providers in counselling patients regarding the risk of recurrence, role of further investigation, and potential preventive interventions in subsequent pregnancies.

9.3.1 | Risk of recurrence based on severity and onset

Most of the data on the risk of recurrence of placenta-mediated complications come from studies evaluating hypertensive complications of pregnancy. In a recent individual patient data meta-analysis of 22 studies, the overall risk of recurrence of hypertensive complications was 21%, and was higher in women who experienced early-onset hypertensive complications.⁴¹⁸ Data on the recurrence of FGR are limited.⁴¹⁹⁻⁴²² In a population-based study, the overall recurrence rate of FGR in women who gave birth to an infant with a birth weight below the 10th percentile was 24%, compared with 6% in women without a history of FGR (OR 3.9; 95% CI, 3.7–4.0). The risk of recurrence was related to the severity of FGR, and was nearly six-fold when the infant birth weight was below the 5th percentile (OR 5.7; 95% CI, $5.4-6.0$).⁴²³ Thus, couples with FGR in the first pregnancy can be reassured that the overall chance of recurrence in subsequent pregnancies is less than 25%. However, interpretation of the data is limited by the lack of distinction between constitutionally SGA infants and infants who were truly growth restricted, as much of the association described in that study may be driven by the recurrence of constitutional SGA. Therefore, counselling regarding the risk of recurrence should be further refined based on the risk factors of the individual patient, severity of FGR as reflected by timing of onset and Doppler findings, the co-presence of pre-eclampsia, and placental histopathological findings.

9.3.2 | Risk of recurrence based on placental histopathology

The results of the placental histopathological examination are important for two main reasons. First, they may assist care providers in counselling couples regarding the most likely etiology of FGR, especially when the clinical presentation and Doppler findings were inconclusive. Second, placental findings may provide valuable information regarding the risk of recurrence, as certain types of placental pathologies are associated with a relatively high recurrence rate. The main types of placental pathologies, the clinical phenotypes associated with these pathologies, and their estimated risks of recurrence are summarized in Table 2.⁴²⁴⁻⁴²⁶

9.3.3 | Role of thrombophilia screening

Whether women who experienced placenta-mediated pregnancy complications should be screened for antiphospholipid syndrome is a matter of debate. Although the consensus criteria for antiphospholipid syndrome include premature birth before 34 weeks for severe pre-eclampsia or features consistent with placental insufficiency including birth weight below the 10th percentile, 427 the association of antiphospholipid (aPL) antibodies with these conditions is relatively weak and conflicting, especially for FGR.⁴²⁸⁻⁴³⁰ In addition, although some care providers recommend treatment with LMWH during pregnancy to women with aPL syndrome and previous preterm birth for placenta-mediated complications, this practice is mostly extrapolated from women with aPL syndrome and recurrent pregnancy loss, where there is some evidence in favor of LMWH.⁴³¹⁻⁴³³ However, the only trial on LMWH in women with aPL syndrome and prior placenta-related complications (FRUIT trial) found no evidence that LMWH improves outcomes in these cases.⁴³⁴ Given the above, there is insufficient evidence to justify routine screening for aPL antibodies in women with prior FGR.⁴³⁵ However, screening for aPL antibodies is recommended in women with a history of thromboembolism or recurrent pregnancy loss (or ≥1 late fetal loss), and may be considered in selected cases of women with a history of severe FGR associated with severe earlyonset pre-eclampsia, when placental examination shows features of severe maternal vascular malperfusion, especially central or multiple areas of villous infarction that are due to multiple spiral artery thromboses.

Management of women already diagnosed with antiphospholipid syndrome based on a history of placenta-mediated complications is also under debate. Based on the evidence from the FRUIT trial described above, some only recommend treatment with aspirin in this setting,⁴³⁶ while others recommend either surveillance or LMWH during the antepartum and postpartum periods.⁴³⁷ Based on available evidence we only recommend treatment with aspirin, and suggest that LMWH be considered only in selected cases, such as for women who have experienced recurrent complications despite aspirin treatment (aspirin failure).

The findings are clearer for inherited thrombophilias. Most prospective studies found no significant association between inherited thrombophilia and placenta-mediated complications.⁴³⁸⁻⁴⁴³ Furthermore, the TIPPS and FRUIT trials found no benefit of LMWH in women with thrombophilia and a history of placenta-mediated pregnancy complications.444,445 These findings were confirmed by a recent individual patient data meta-analysis that found no benefit of LMWH in decreasing the risk of recurrence of placenta-mediated complications, including in women with thrombophilia.¹⁴⁰ Therefore, there is no indication for routine screening for inherited thrombophilia in women with prior FGR.^{446,447}

9.3.4 | Preconception counselling and management of future pregnancies

Given the considerable risk of recurrence of FGR, efforts should be made to decrease this risk in future pregnancies.⁴⁴⁸ Modifiable risk factors for FGR such as smoking or poor nutritional status should be identified as early as possible and managed accordingly, as discussed in section 5.5.1.

There is some evidence that administration of aspirin can reduce the risk of FGR. However, as described in section 5.5.2 most available data focused on the prevention of pre-eclampsia as the primary outcome in women at high risk of pre-eclampsia, with the prevention of FGR being considered a secondary outcome. Data on the prevention of recurrence of FGR in women with a history of FGR are limited.⁴⁴⁹ Therefore, some recommend that aspirin should be considered in women with past FGR only if they have risk factors for pre-eclampsia at the time of the next pregnancy.⁴⁵⁰ However, we believe that given the safety of aspirin and the overlap in pathogenesis of pre-eclampsia and FGR, it is reasonable to recommend aspirin to women with a history of placenta-mediated FGR in the previous pregnancy, using the same regimen of aspirin used for the prevention of pre-eclampsia. This recommendation is shared by most professional societies.134

Data on the role of LMWH to prevent recurrence of placentamediated complications including FGR are conflicting, and this topic is reviewed in section 5.5.2. Based on available data, LMWH therapy should not be used in women with a past history of FGR except in a research setting.

Given the association of insufficient gestational weight gain with FGR, we recommend monitoring of weight gain and informing women about their target weight gain range, as described in section 5.5.1. Other interventions, such as bed rest or nutritional supplements are of unproven benefit and should not be routinely offered.451,452 The risk of recurrence can be further stratified in early pregnancy by means of prenatal screening with biochemical markers (PAPP-A, beta hCG, alpha-fetoprotein, and PlGF) as well as by uterine artery Doppler, as described in section 5. Due to the increased risk of recurrence, pregnant women with a history of FGR in a previous pregnancy should be managed in a high-risk pregnancy clinic and should receive closer antenatal surveillance, including close monitoring of fetal growth and maternal blood pressure.⁴⁵³

Abbreviations: CNS, central nervous system; FGR, fetal growth restriction; IVIG, intravenous immunoglobulins.

TABLE 2 Phenotypes and risk of recurrence associated with specific types of placental pathologies. **TABLE 2** Phenotypes and risk of recurrence associated with specific types of placental pathologies. $\frac{\binom{2}{3}}{\text{FIGO}}$

9.4 | **Recommendations**

FIGO recommends the following for postpartum assessment and counselling for future pregnancies in women with a history of fetal growth restriction (FGR)

10 | SUMMARY AND FUTURE RESEARCH **DIRECTIONS**

FGR is an important cause of stillbirth, neonatal mortality, and shortand long-term neonatal morbidity. Early prediction and preventive strategies, timely diagnosis, and management using a standardized protocol to determine the proper monitoring and timing of delivery can decrease the risk of stillbirth and improve perinatal outcomes in pregnancies complicated by FGR.

Future research should focus on the development of new fetal assessment tools that may improve the accuracy of the prediction of fetal deterioration and thus further optimize timing of delivery of FGR fetuses, as well as on novel treatments that may improve placental function in cases of placenta-mediated FGR and thereby deferring delivery in cases of early-onset FGR.

REFERENCES

- 1. Gordijn SJ, Beune IM, Thilaganathan B, et al. Consensus definition of fetal growth restriction: a Delphi procedure. *Ultrasound Obstet Gynecol*. 2016;48(3):333-339.
- 2. Hadlock FP, Harrist RB, Sharman RS, Deter RL, Park SK. Estimation of fetal weight with the use of head, body, and femur measurements–a prospective study. *Am J Obstet Gynecol*. 1985;151(3):333-337.
- 3. Visser GHA, Nicholson WK, Barnea ER, et al. FIGO position paper on reference charts for fetal growth and size at birth: Which one to use? *Int J Gynecol Obstet*. 2020 Nov 28. [Epub ahead of print].
- 4. Sheiner E, Kapur A, Retnakaran R, et al. FIGO (International Federation of Gynecology and Obstetrics) postpregnancy initiative: long-term maternal implications of pregnancy complications-follow-up considerations. *Int J Gynecol Obstet*. 2019;147(Suppl 1):1-31.
- 5. Guyatt GH, Oxman AD, Vist GE, et al. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ*. 2008;336(7650):924-926.
- 6. Schunemann HJ, Jaeschke R, Cook DJ, et al. An official ATS statement: grading the quality of evidence and strength of recommendations in ATS guidelines and recommendations. *Am J Respir Crit Care Med*. 2006;174:605-614.
- 7. Balshem H, Helfand M, Schunemann HJ, et al. GRADE guidelines: 3. Rating the quality of evidence. *J Clin Epidemiol*. 2011;64(4): 401-406.
- 8. Pels A, Beune IM, van Wassenaer-Leemhuis AG, Limpens J, Ganzevoort W. Early-onset fetal growth restriction: a systematic review on mortality and morbidity. *Acta Obstet Gynecol Scand*. 2020;99(2):153-166.
- 9. Baschat AA, Viscardi RM, Hussey-Gardner B, Hashmi N, Harman C. Infant neurodevelopment following fetal growth restriction: relationship with antepartum surveillance parameters. *Ultrasound Obstet Gynecol*. 2009;33(1):44-50.
- 10. Baschat AA. Neurodevelopment following fetal growth restriction and its relationship with antepartum parameters of placental dysfunction. *Ultrasound Obstet Gynecol*. 2011;37(5):501-514.
- 11. Baschat AA. Neurodevelopment after fetal growth restriction. *Fetal Diagn Ther*. 2014;36(2):136-142.
- 12. Levine TA, Grunau RE, McAuliffe FM, Pinnamaneni R, Foran A, Alderdice FA. Early childhood neurodevelopment after intrauterine growth restriction: a systematic review. *Pediatrics*. 2015;135(1):126-141.
- 13. Murray E, Fernandes M, Fazel M, Kennedy SH, Villar J, Stein A. Differential effect of intrauterine growth restriction on childhood neurodevelopment: a systematic review. *BJOG*. 2015;122(8):1062-1072.
- 14. Melamed N, Asztalos E, Murphy K, et al. Neurodevelopmental disorders among term infants exposed to antenatal corticosteroids during pregnancy: a population-based study. *BMJ Open*. 2019;9(9):e031197.
- 15. Crispi F, Miranda J, Gratacos E. Long-term cardiovascular consequences of fetal growth restriction: biology, clinical implications, and opportunities for prevention of adult disease. *Am J Obstet Gynecol*. 2018;218(2S):S869-S879.
- 16. Figueras F, Gratacos E. Update on the diagnosis and classification of fetal growth restriction and proposal of a stage-based management protocol. *Fetal Diagn Ther*. 2014;36(2):86-98.
- 17. Mifsud W, Sebire NJ. Placental pathology in early-onset and late-onset fetal growth restriction. *Fetal Diagn Ther*. 2014;36(2):117-128.
- 18. Aviram A, Sherman C, Kingdom J, Zaltz A, Barrett J, Melamed N. Defining early vs late fetal growth restriction by placental pathology. *Acta Obstet Gynecol Scand*. 2019;98(3):365-373.
- 19. Crovetto F, Crispi F, Scazzocchio E, et al. First-trimester screening for early and late small-for-gestational-age neonates using maternal serum biochemistry, blood pressure and uterine artery Doppler. *Ultrasound Obstet Gynecol*. 2014;43(1):34-40.
- 20. Maulik D. Fetal growth restriction: the etiology. *Clin Obstet Gynecol*. 2006;49(2):228-235.
- 21. Nardozza LM, Caetano AC, Zamarian AC, et al. Fetal growth restriction: current knowledge. *Arch Gynecol Obstet*. 2017;295(5):1061-1077.
- 22. Hiersch L, Yogev Y. Pregnancy: impact of maternal nutrition on intrauterine fetal growth. *World Rev Nutr Diet*. 2018;117:151-164.
- 23. Yogev Y, Hiersch L. Pregnancy: impact of maternal nutrition on intrauterine fetal growth. *World Rev Nutr Diet*. 2014;109:101-108.
- 24. Ghaly A, Maki Y, Nygard K, Hammond R, Hardy DB, Richardson BS. Maternal nutrient restriction in guinea pigs leads to fetal growth restriction with increased brain apoptosis. *Pediatric Res*. 2019;85(1):105-112.
- 25. Liu Y, Li H, Sha Q, et al. Effects of maternal undernutrition on the growth, development and antioxidant status of ovine placentome subtypes during late pregnancy. *Theriogenology*. 2018;110:96-102.
- 26. Papathakis PC, Singh LN, Manary MJ. How maternal malnutrition affects linear growth and development in the offspring. *Mol Cell Endocrinol*. 2016;435:40-47.
- 27. Rahman MM, Abe SK, Rahman MS, et al. Maternal anemia and risk of adverse birth and health outcomes in low- and middle-income countries: systematic review and meta-analysis. *Am J Clin Nutr*. 2016;103(2):495-504.
- 28. Stangret A, Wnuk A, Szewczyk G, Pyzlak M, Szukiewicz D. Maternal hemoglobin concentration and hematocrit values may affect fetus development by influencing placental angiogenesis. *J Matern Fetal Neonatal Med*. 2017;30(2):199-204.
- 29. Burton GJ, Jauniaux E. Pathophysiology of placental-derived fetal growth restriction. *Am J Obstet Gynecol*. 2018;218(2S):S745-S761.
- 30. Kingdom JC, Audette MC, Hobson SR, Windrim RC, Morgen E. A placenta clinic approach to the diagnosis and management of fetal growth restriction. *Am J Obstet Gynecol*. 2018;218(2S):S80 3-S817.
- 31. Zur RL, Kingdom JC, Parks WT, Hobson SR. The Placental Basis of Fetal Growth Restriction. *Obstet Gynecol Clin North Am*. 2020;47(1):81-98.
- 32. Gluck O, Schreiber L, Marciano A, Mizrachi Y, Bar J, Kovo M. Pregnancy outcome and placental pathology in small for gestational age neonates in relation to the severity of their growth restriction. *J Matern Fetal Neonatal Med*. 2019;32(9):1468-1473.
- 33. Bos M, Harris-Mostert E, van der Meeren LE, et al. Clinical outcomes in chronic intervillositis of unknown etiology. *Placenta*. 2020;91:19-23.
- 34. Snijders RJ, Sherrod C, Gosden CM, Nicolaides KH. Fetal growth retardation: associated malformations and chromosomal abnormalities. *Am J Obstet Gynecol*. 1993;168(2):547-555.
- 35. Borrell A, Grande M, Pauta M, Rodriguez-Revenga L, Figueras F. Chromosomal microarray analysis in fetuses with growth restriction and normal karyotype: a systematic review and meta-analysis. *Fetal Diagn Ther*. 2018;44(1):1-9.
- 36. Sagi-Dain L, Peleg A, Sagi S. Risk for chromosomal aberrations in apparently isolated intrauterine growth restriction: a systematic review. *Prenatal Diagn*. 2017;37(11):1061-1066.
- 37. Borrell A, Grande M, Meler E, et al. Genomic microarray in fetuses with early growth restriction: a multicenter study. *Fetal Diagn Ther*. 2017;42(3):174-180.
- 38. Khoury MJ, Erickson JD, Cordero JF, McCarthy BJ. Congenital malformations and intrauterine growth retardation: a population study. *Pediatrics*. 1988;82(1):83-90.
- 39. Longo S, Borghesi A, Tzialla C, Stronati M. IUGR and infections. *Early Hum Dev*. 2014;90(Suppl 1):S42-44.

46 | ALLER CANCOLOGY (WE) $\langle \hat{X} \rangle$

- 40. Crino JP, Driggers RW. Ultrasound findings associated with antepartum viral infection. *Clin Obstet Gynecol*. 2018;61(1):106-121.
- 41. Leruez-Ville M, Ville Y. Fetal cytomegalovirus infection. *Best Pract Res Clin Obstet Gynaecol*. 2017;38:97-107.
- 42. Accrombessi M, Zeitlin J, Massougbodji A, Cot M, Briand V. What do we know about risk factors for fetal growth restriction in Africa at the time of sustainable development goals? A Scoping Review. *Paediatr Perinat Epidemiol*. 2018;32(2):184-196.
- 43. Platt DJ, Miner JJ. Consequences of congenital Zika virus infection. *Curr Opin Virol*. 2017;27:1-7.
- 44. Mandelbrot L. Fetal varicella diagnosis, management, and outcome. *Prenatal Diagn*. 2012;32(6):511-518.
- 45. Seitz J, Morales-Prieto DM, Favaro RR, Schneider H, Markert UR. Molecular principles of intrauterine growth restriction in plasmodium falciparum infection. *Front Endocrinol*. 2019;10:98.
- 46. Umbers AJ, Aitken EH, Rogerson SJ. Malaria in pregnancy: small babies, big problem. *Trends Parasitol*. 2011;27(4):168-175.
- 47. Yoon I, Slesinger TL. Radiation exposure in pregnancy. In: StatPearls [Internet]. Treasure Island, FL: StatPearls Publishing; 2020.
- 48. Ganapathy V. Drugs of abuse and human placenta. *Life Sci*. 2011;88(21–22):926-930.
- 49. Holbrook BD, Rayburn WF. Teratogenic risks from exposure to illicit drugs. *Obstet Gynecol Clin North Am*. 2014;41(2):229-239.
- 50. Carter RC, Jacobson JL, Molteno CD, Dodge NC, Meintjes EM, Jacobson SW. Fetal alcohol growth restriction and cognitive impairment. *Pediatrics*. 2016;138(2):e20160775.
- 51. Basso O, Wilcox AJ, Weinberg CR. Birth weight and mortality: causality or confounding? *Am J Epidemiol*. 2006;164(4):303-311.
- 52. Flenady V, Koopmans L, Middleton P, et al. Major risk factors for stillbirth in high-income countries: a systematic review and meta-analysis. *Lancet*. 2011;377(9774):1331-1340.
- 53. Gardosi J, Madurasinghe V, Williams M, Malik A, Francis A. Maternal and fetal risk factors for stillbirth: population based study. *BMJ*. 2013;346:f108.
- 54. Pilliod RA, Cheng YW, Snowden JM, Doss AE, Caughey AB. The risk of intrauterine fetal death in the small-for-gestational-age fetus. *Am J Obstet Gynecol*. 2012;207(4):318.e311-318e316.
- 55. Kady SM, Gardosi J. Perinatal mortality and fetal growth restriction. *Best Pract Res Clin Obstet Gynaecol*. 2004;18(3):397-410.
- 56. Bukowski R, Hansen NI, Willinger M, et al. Fetal growth and risk of stillbirth: a population-based case-control study. *PLoS Med*. 2014;11(4):e1001633.
- 57. McIntire DD, Bloom SL, Casey BM, Leveno KJ. Birth weight in relation to morbidity and mortality among newborn infants. *N Engl J Med*. 1999;340(16):1234-1238.
- 58. Hiersch L, Lipworth H, Kingdom J, Barrett J, Melamed N. Identification of the optimal growth chart and threshold for the prediction of antepartum stillbirth. *Arch Gynecol Obstet*. 2020 Aug 14. [Epub ahead of print].
- 59. Temming LA, Dicke JM, Stout MJ, et al. Early second-trimester fetal growth restriction and adverse perinatal outcomes. *Obstet Gynecol*. 2017;130(4):865-869.
- 60. Figueras F, Gratacos E. An integrated approach to fetal growth restriction. *Best Pract Res Clin Obstet Gynaecol*. 2017;38:48-58.
- 61. Lackman F, Capewell V, Richardson B, daSilva O, Gagnon R. The risks of spontaneous preterm delivery and perinatal mortality in relation to size at birth according to fetal versus neonatal growth standards. *Am J Obstet Gynecol*. 2001;184(5):946-953.
- 62. Maulik D, Frances Evans J, Ragolia L. Fetal growth restriction: pathogenic mechanisms. *Clin Obstet Gynecol*. 2006;49(2):219-227.
- 63. Proctor LK, Kfouri J, Hiersch L, et al. Association between hypertensive disorders and fetal growth restriction in twin compared with singleton gestations. *Am J Obstet Gynecol*. 2019;221(3):251. e1-251.e8.
- 64. Ananth CV, Oyelese Y, Prasad V, Getahun D, Smulian JC. Evidence of placental abruption as a chronic process: associations with

vaginal bleeding early in pregnancy and placental lesions. *Eur J Obstet Gynecol Reprod Biol*. 2006;128(1–2):15-21.

- 65. Hiersch L, Shinar S, Melamed N, et al. Recurrent placenta-mediated complications in women with three consecutive deliveries. *Obstet Gynecol*. 2017;129(3):416-421.
- 66. Gardosi J, Francis A. Adverse pregnancy outcome and association with small for gestational age birthweight by customized and population-based percentiles. *Am J Obstet Gynecol*. 2009;201(1):28. e1-8.
- 67. Pallotto EK, Kilbride HW. Perinatal outcome and later implications of intrauterine growth restriction. *Clin Obstet Gynecol*. 2006;49(2):257-269.
- 68. Baschat AA, Cosmi E, Bilardo CM, et al. Predictors of neonatal outcome in early-onset placental dysfunction. *Obstet Gynecol*. 2007;109(2 Pt 1):253-261.
- 69. Cavallaro A, Veglia M, Svirko E, Vannuccini S, Volpe G, Impey L. Using fetal abdominal circumference growth velocity in the prediction of adverse outcome in near-term small-for-gestational-age fetuses. *Ultrasound Obstet Gynecol*. 2018;52(4):494-500.
- 70. Sharma D, Farahbakhsh N, Shastri S, Sharma P. Intrauterine growth restriction - part 2. *J Matern Fetal Neonatal Med*. 2016;29(24):4037-4048.
- 71. Sharma D, Shastri S, Sharma P. Intrauterine growth restriction: antenatal and postnatal aspects. *Clin Med Insights Pediatr*. 2016;10:67-83.
- 72. Flamant C, Gascoin G. Short-term outcome and small for gestational age newborn management [in French]. *J Gynecol Obstet Biol Reprod (Paris)*. 2013;42(8):985-995.
- 73. Figueras F, Oros D, Cruz-Martinez R, et al. Neurobehavior in term, small-for-gestational age infants with normal placental function. *Pediatrics*. 2009;124(5):e934-941.
- 74. Eixarch E, Meler E, Iraola A, et al. Neurodevelopmental outcome in 2-year-old infants who were small-for-gestational age term fetuses with cerebral blood flow redistribution. *Ultrasound Obstet Gynecol*. 2008;32(7):894-899.
- 75. Savchev S, Sanz-Cortes M, Cruz-Martinez R, et al. Neurodevelopmental outcome of full-term small-for-gestational-age infants with normal placental function. *Ultrasound Obstet Gynecol*. 2013;42(2):201-206.
- 76. Figueras F, Cruz-Martinez R, Sanz-Cortes M, et al. Neurobehavioral outcomes in preterm, growth-restricted infants with and without prenatal advanced signs of brain-sparing. *Ultrasound Obstet Gynecol*. 2011;38(3):288-294.
- 77. Sanz-Cortes M, Figueras F, Bargallo N, Padilla N, Amat-Roldan I, Gratacos E. Abnormal brain microstructure and metabolism in small-for-gestational-age term fetuses with normal umbilical artery Doppler. *Ultrasound Obstet Gynecol*. 2010;36(2):159-165.
- 78. Padilla N, Falcon C, Sanz-Cortes M, et al. Differential effects of intrauterine growth restriction on brain structure and development in preterm infants: a magnetic resonance imaging study. *Brain Res*. 2011;1382:98-108.
- 79. Barker DJ. Adult consequences of fetal growth restriction. *Clin Obstet Gynecol*. 2006;49(2):270-283.
- 80. Barker DJ, Gluckman PD, Godfrey KM, Harding JE, Owens JA, Robinson JS. Fetal nutrition and cardiovascular disease in adult life. *Lancet*. 1993;341(8850):938-941.
- 81. Crispi F, Figueras F, Cruz-Lemini M, Bartrons J, Bijnens B, Gratacos E. Cardiovascular programming in children born small for gestational age and relationship with prenatal signs of severity. *Am J Obstet Gynecol*. 2012;207(2):121.e1-9.
- 82. Valsamakis G, Kanaka-Gantenbein C, Malamitsi-Puchner A, Mastorakos G. Causes of intrauterine growth restriction and the postnatal development of the metabolic syndrome. *Ann N Y Acad Sci*. 2006;1092:138-147.
- 83. Poon LC, Karagiannis G, Staboulidou I, Shafiei A, Nicolaides KH. Reference range of birth weight with gestation and first-trimester

prediction of small-for-gestation neonates. *Prenatal Diagn*. 2011;31(1):58-65.

- 84. Odibo AO, Nelson D, Stamilio DM, Sehdev HM, Macones GA. Advanced maternal age is an independent risk factor for intrauterine growth restriction. *Am J Perinatol*. 2006;23(5):325-328.
- 85. Han Z, Mulla S, Beyene J, Liao G, McDonald SD; Knowledge Synthesis Group. Maternal underweight and the risk of preterm birth and low birth weight: a systematic review and meta-analyses. *Int J Epidemiol*. 2011;40(1):65-101.
- 86. Jaddoe VW, Bakker R, Hofman A, et al. Moderate alcohol consumption during pregnancy and the risk of low birth weight and preterm birth. The Generation R Study. *Ann Epidemiol*. 2007;17(10):834-840.
- 87. McCowan LM, Dekker GA, Chan E, et al. Spontaneous preterm birth and small for gestational age infants in women who stop smoking early in pregnancy: prospective cohort study. *BMJ*. 2009;338:b1081.
- 88. Gouin K, Murphy K, Shah PS. Knowledge Synthesis Group on Determinants of Low Birth W, Preterm B: Effects of cocaine use during pregnancy on low birthweight and preterm birth: systematic review and metaanalyses. *Am J Obstet Gynecol*. 2011;204(4):340. e1-12.
- 89. Jackson RA, Gibson KA, Wu YW, Croughan MS. Perinatal outcomes in singletons following in vitro fertilization: a meta-analysis. *Obstet Gynecol*. 2004;103(3):551-563.
- 90. Andres RL, Day MC. Perinatal complications associated with maternal tobacco use. *Semin Neonatol*. 2000;5(3):231-241.
- 91. Di Renzo GC, Conry JA, Blake J, et al. International Federation of Gynecology and Obstetrics opinion on reproductive health impacts of exposure to toxic environmental chemicals. *Int J Gynecol Obstet*. 2015;131(3):219-225.
- 92. Smith GC. First trimester origins of fetal growth impairment. *Semin Perinatol*. 2004;28(1):41-50.
- 93. Smith GCS. Universal screening for foetal growth restriction. *Best Pract Res Clin Obstet Gynaecol*. 2018;49:16-28.
- 94. Morris RK, Bilagi A, Devani P, Kilby MD. Association of serum PAPP-A levels in first trimester with small for gestational age and adverse pregnancy outcomes: systematic review and meta-analysis. *Prenatal Diagn*. 2017;37(3):253-265.
- 95. Gaccioli F, Aye I, Sovio U, Charnock-Jones DS, Smith GCS. Screening for fetal growth restriction using fetal biometry combined with maternal biomarkers. *Am J Obstet Gynecol*. 2018;218(2S):S725-S737.
- 96. Lesmes C, Gallo DM, Gonzalez R, Poon LC, Nicolaides KH. Prediction of small-for-gestational-age neonates: screening by maternal serum biochemical markers at 19–24 weeks. *Ultrasound Obstet Gynecol*. 2015;46(3):341-349.
- 97. Smith GC, Shah I, White IR, Pell JP, Crossley JA, Dobbie R. Maternal and biochemical predictors of antepartum stillbirth among nulliparous women in relation to gestational age of fetal death. *BJOG*. 2007;114(6):705-714.
- 98. Proctor LK, Toal M, Keating S, et al. Placental size and the prediction of severe early-onset intrauterine growth restriction in women with low pregnancy-associated plasma protein-A. *Ultrasound Obstet Gynecol*. 2009;34(3):274-282.
- 99. Benn PA, Horne D, Briganti S, Rodis JF, Clive JM. Elevated second-trimester maternal serum hCG alone or in combination with elevated alpha-fetoprotein. *Obstet Gynecol*. 1996;87(2):217-222.
- 100. Wang A, Rana S, Karumanchi SA. Preeclampsia: the role of angiogenic factors in its pathogenesis. *Physiology (Bethesda)*. 2009;24:147-158.
- 101. Smith GC, Crossley JA, Aitken DA, et al. Circulating angiogenic factors in early pregnancy and the risk of preeclampsia, intrauterine growth restriction, spontaneous preterm birth, and stillbirth. *Obstet Gynecol*. 2007;109(6):1316-1324.
- 102. Erez O, Romero R, Espinoza J, et al. The change in concentrations of angiogenic and anti-angiogenic factors in maternal plasma between the first and second trimesters in risk assessment for the subsequent development of preeclampsia and small-for-gestational age. *J Matern Fetal Neonatal Med*. 2008;21(5):279-287.
- 103. Poon LC, Zaragoza E, Akolekar R, Anagnostopoulos E, Nicolaides KH. Maternal serum placental growth factor (PlGF) in small for gestational age pregnancy at 11(+0) to 13(+6) weeks of gestation. *Prenatal Diagn*. 2008;28(12):1110-1115.
- 104. Cowans NJ, Stamatopoulou A, Matwejew E, von Kaisenberg CS, Spencer K. First-trimester placental growth factor as a marker for hypertensive disorders and SGA. *Prenatal Diagn*. 2010;30(6):565-570.
- 105. Maynard SE, Min JY, Merchan J, et al. Excess placental soluble fms-like tyrosine kinase 1 (sFlt1) may contribute to endothelial dysfunction, hypertension, and proteinuria in preeclampsia. *J Clin Invest*. 2003;111(5):649-658.
- 106. Zeisler H, Llurba E, Chantraine F, et al. Predictive value of the sFlt-1:PlGF ratio in women with suspected preeclampsia. *New Engl J Med*. 2016;374(1):13-22.
- 107. Levytska K, Higgins M, Keating S, et al. Placental pathology in relation to uterine artery doppler findings in pregnancies with severe intrauterine growth restriction and abnormal umbilical artery doppler changes. *Am J Perinatol*. 2017;34(5):451-457.
- 108. Toal M, Keating S, Machin G, et al. Determinants of adverse perinatal outcome in high-risk women with abnormal uterine artery Doppler images. *Am J Obstet Gynecol*. 2008;198(3):330.e1-7.
- 109. Lesmes C, Gallo DM, Saiid Y, Poon LC, Nicolaides KH. Prediction of small-for-gestational-age neonates: screening by uterine artery Doppler and mean arterial pressure at 19–24 weeks. *Ultrasound Obstet Gynecol*. 2015;46(3):332-340.
- 110. Cnossen JS, Morris RK, ter Riet G, et al. Use of uterine artery Doppler ultrasonography to predict pre-eclampsia and intrauterine growth restriction: a systematic review and bivariable meta-analysis. *CMAJ*. 2008;178(6):701-711.
- 111. Drouin O, Boutin A, Paquette K, et al. First-trimester uterine artery doppler for the prediction of SGA at birth: the great obstetrical syndromes study. *J Obstet Gynaecol Can*. 2018;40(12):1592-1599.
- 112. Rodriguez A, Tuuli MG, Odibo AO. First-, second-, and third-trimester screening for preeclampsia and intrauterine growth restriction. *Clin Lab Med*. 2016;36(2):331-351.
- 113. Moloney A, Hladunewich M, Manly E, et al. The predictive value of sonographic placental markers for adverse pregnancy outcome in women with chronic kidney disease. *Pregnancy Hypertens*. 2020;20:27-35.
- 114. Viero S, Chaddha V, Alkazaleh F, et al. Prognostic value of placental ultrasound in pregnancies complicated by absent end-diastolic flow velocity in the umbilical arteries. *Placenta*. 2004;25(8–9):735-741.
- 115. Proctor LK, Whittle WL, Keating S, Viero S, Kingdom JC. Pathologic basis of echogenic cystic lesions in the human placenta: role of ultrasound-guided wire localization. *Placenta*. 2010;31(12):1111-1115.
- 116. Porat S, Fitzgerald B, Wright E, Keating S, Kingdom JC. Placental hyperinflation and the risk of adverse perinatal outcome. *Ultrasound Obstet Gynecol*. 2013;42(3):315-321.
- 117. Florido J, Ocon O, del Castillo L, et al. Analysis of measurement process of placental volume in early pregnancy: an interobserver reliability study. *J Perinat Med*. 2014;42(5):559-564.
- 118. Burstein E, Sheiner E, Hershkovitz R. Three-dimensional placental volume measurements between 11 and 13 weeks' gestation. *Am J Perinatol*. 2009;26(2):169-171.
- 119. Farina A. Systematic review on first trimester three-dimensional placental volumetry predicting small for gestational age infants. *Prenatal Diagn*. 2016;36(2):135-141.

48 IMALLE IN CONFIDENTAL CONFIDENTIAL

- 120. Hafner E, Metzenbauer M, Hofinger D, et al. Comparison between three-dimensional placental volume at 12 weeks and uterine artery impedance/notching at 22 weeks in screening for pregnancy-induced hypertension, pre-eclampsia and fetal growth restriction in a low-risk population. *Ultrasound Obstet Gynecol*. 2006;27(6):652-657.
- 121. Crovetto F, Triunfo S, Crispi F, et al. Differential performance of first-trimester screening in predicting small-for-gestational-age neonate or fetal growth restriction. *Ultrasound Obstet Gynecol*. 2017;49(3):349-356.
- 122. McCowan LM, Thompson JM, Taylor RS, et al. Prediction of small for gestational age infants in healthy nulliparous women using clinical and ultrasound risk factors combined with early pregnancy biomarkers. *PLoS One*. 2017;12(1):e0169311.
- 123. Grivell R, Dodd J, Robinson J. The prevention and treatment of intrauterine growth restriction. *Best Pract Res Clin Obstet Gynaecol*. 2009;23(6):795-807.
- 124. Cedergren M. Effects of gestational weight gain and body mass index on obstetric outcome in Sweden. *Int J Gynecol Obstet*. 2006;93(3):269-274.
- 125. Rasmussen KM, Yaktine AL, eds. *Institute of Medicine (US) and National Research Council (US) Committee to Reexamine IOM Pregnancy Weight Guidelines. Weight Gain During Pregnancy: Reexamining the Guidelines*. Washington, DC: National Academies Press (US); 2009.
- 126. Cedergren MI. Optimal gestational weight gain for body mass index categories. *Obstet Gynecol*. 2007;110(4):759-764.
- 127. Lumley J, Chamberlain C, Dowswell T, Oliver S, Oakley L, Watson L. Interventions for promoting smoking cessation during pregnancy. *Cochrane Database Syst Rev*. 2009(3):CD001055.
- 128. Tobacco and nicotine cessation during pregnancy: ACOG Committee opinion, number 807. *Obstet Gynecol*. 2020;135(5):e22 1-e229.
- 129. Mills JL, Graubard BI, Harley EE, Rhoads GG, Berendes HW. Maternal alcohol consumption and birth weight. How much drinking during pregnancy is safe? *JAMA*. 1984;252(14):1875-1879.
- 130. Rolnik DL, Wright D, Poon LC, et al. Aspirin versus placebo in pregnancies at high risk for preterm preeclampsia. *New Engl J Med*. 2017;377(7):613-622.
- 131. Meher S, Duley L, Hunter K, Askie L. Antiplatelet therapy before or after 16 weeks' gestation for preventing preeclampsia: an individual participant data meta-analysis. *Am J Obstet Gynecol*. 2017;216(2):121-128.e2.
- 132. Roberge S, Nicolaides K, Demers S, Hyett J, Chaillet N, Bujold E. The role of aspirin dose on the prevention of preeclampsia and fetal growth restriction: systematic review and meta-analysis. *Am J Obstet Gynecol*. 2017;216(2):110-120.e6.
- 133. Ayala DE, Ucieda R, Hermida RC. Chronotherapy with low-dose aspirin for prevention of complications in pregnancy. *Chronobiol Int*. 2013;30(1–2):260-279.
- 134. McCowan LM, Figueras F, Anderson NH. Evidence-based national guidelines for the management of suspected fetal growth restriction: comparison, consensus, and controversy. *Am J Obstet Gynecol*. 2018;218(2S):S855-S868.
- 135. Sobel ML, Kingdom J, Drewlo S. Angiogenic response of placental villi to heparin. *Obstet Gynecol*. 2011;117(6):1375-1383.
- 136. Mousavi S, Moradi M, Khorshidahmad T, Motamedi M. Antiinflammatory effects of heparin and its derivatives: a systematic review. *Adv Pharmacol Sci*. 2015;2015:507151.
- 137. Yinon Y, Ben Meir E, Margolis L, et al. Low molecular weight heparin therapy during pregnancy is associated with elevated circulatory levels of placental growth factor. *Placenta*. 2015;36(2):121-124.
- 138. Oberkersch R, Attorresi AI, Calabrese GC. Low-molecular-weight heparin inhibition in classical complement activation pathway during pregnancy. *Thromb Res*. 2010;125(5):e240-245.
- 139. Rodger MA, Carrier M, Le Gal G, et al. Meta-analysis of lowmolecular-weight heparin to prevent recurrent placentamediated pregnancy complications. *Blood*. 2014;123(6): 822-828.
- 140. Rodger MA, Gris J-C, de Vries JIP, et al. Low-molecular-weight heparin and recurrent placenta-mediated pregnancy complications: a meta-analysis of individual patient data from randomised controlled trials. *Lancet*. 2016;388(10060):2629-2641.
- 141. Groom KM, McCowan LM, Mackay LK, et al. Enoxaparin for the prevention of preeclampsia and intrauterine growth restriction in women with a history: a randomized trial. *Am J Obstet Gynecol*. 2017;216(3):296.e1-296.e14.
- 142. McLaughlin K, Scholten RR, Parker JD, Ferrazzi E, Kingdom JCP. Low molecular weight heparin for the prevention of severe preeclampsia: where next? *Br J Clin Pharmacol*. 2018;84(4): 673-678.
- 143. American College of Obstetricians and Gynecologists' Committee on Practice Bulletins—Obstetrics and the Society for Maternal-Fetal Medicine. ACOG practice bulletin no. 204: fetal growth restrictions *Obstet Gynecol*. 2019;133(2):e97-e109.
- 144. Morse K, Williams A, Gardosi J. Fetal growth screening by fundal height measurement. *Best Pract Res Clin Obstet Gynaecol*. 2009;23(6):809-818.
- 145. Papageorghiou AT, Ohuma EO, Gravett MG, et al. International standards for symphysis-fundal height based on serial measurements from the Fetal Growth Longitudinal Study of the INTERGROWTH-21st Project: prospective cohort study in eight countries. *BMJ*. 2016;355:i5662.
- 146. Sparks TN, Cheng YW, McLaughlin B, Esakoff TF, Caughey AB. Fundal height: a useful screening tool for fetal growth? *J Matern Fetal Neonatal Med*. 2011;24(5):708-712.
- 147. Knox AJ, Sadler L, Pattison NS, Mantell CD, Mullins P. An obstetric scoring system: its development and application in obstetric management. *Obstet Gynecol*. 1993;81(2):195-199.
- 148. Azziz R, Smith S, Fabro S. The development and use of a standard symphysial-fundal height growth curve in the prediction of small for gestational age neonates. *Int J Gynecol Obstet*. 1988;26(1):81-87.
- 149. Belizan JM, Villar J, Nardin JC, Malamud J, De Vicurna LS. Diagnosis of intrauterine growth retardation by a simple clinical method: measurement of uterine height. *Am J Obstet Gynecol*. 1978;131(6):643-646.
- 150. Calvert JP, Crean EE, Newcombe RG, Pearson JF. Antenatal screening by measurement of symphysis-fundus height. *Br Med J (Clin Res Ed)*. 1982;285(6345):846-849.
- 151. Challis K, Osman NB, Nystrom L, Nordahl G, Bergstrom S. Symphysis-fundal height growth chart of an obstetric cohort of 817 Mozambican women with ultrasound-dated singleton pregnancies. *Trop Med Int Health*. 2002;7(8):678-684.
- 152. Ebite LE, Ebeigbe PN, Igbigbi P, Akpuaka FC. Symphysiofundal height growth curve and growth velocity in pregnant women in a Nigerian community. *J Obstet Gynaecol*. 2009;29(7):605-608.
- 153. Hakansson A, Aberg A, Nyberg P, Schersten B. A new symphysis-fundus height growth chart based on a well defined female population with ultrasound-dated singleton pregnancies. *Acta Obstet Gynecol Scand*. 1995;74(9):682-686.
- 154. Jensen OH, Larsen S. Evaluation of symphysis-fundus measurements and weighing during pregnancy. *Acta Obstet Gynecol Scand*. 1991;70(1):13-16.
- 155. Pay AS, Wiik J, Backe B, Jacobsson B, Strandell A, Klovning A. Symphysis-fundus height measurement to predict small-for-gestational-age status at birth: a systematic review. *BMC Pregnancy Childbirth*. 2015;15:22.
- 156. Pay ASD, Froen JF, Staff AC, Jacobsson B, Gjessing HK. Symphysisfundus measurement – the predictive value of a new reference curve. *Tidsskr Nor Laegeforen*. 2017;137(10):717-720.
- 157. Robert Peter J, Ho JJ, Valliapan J, Sivasangari S. Symphysial fundal height (SFH) measurement in pregnancy for detecting abnormal fetal growth. *Cochrane Database Syst Rev*. 2015(9):CD008136.
- 158. Goto E. Prediction of low birthweight and small for gestational age from symphysis-fundal height mainly in developing countries: a meta-analysis. *J Epidemiol Community Health*. 2013;67(12):999-1005.
- 159. Griffiths A, Pinto A, Margarit L. A survey of methods used to measure symphysis fundal height. *J Obstet Gynaecol*. 2008;28(7):692-694.
- 160. Salomon LJ, Alfirevic Z, Da Silva CF, et al. ISUOG Practice Guidelines: ultrasound assessment of fetal biometry and growth. *Ultrasound Obstet Gynecol*. 2019;53(6):715-723.
- 161. Melamed N, Yogev Y, Meizner I, Mashiach R, Bardin R, Ben-Haroush A. Sonographic fetal weight estimation: which model should be used? *J Ultrasound Med*. 2009;28(5):617-629.
- 162. Nahum GG, Stanislaw H. Ultrasonographic prediction of term birth weight: how accurate is it? *Am J Obstet Gynecol*. 2003;188(2):566-574.
- 163. Stanislaw H, Nahum GG. Accuracy of birth weight prediction methods. *J Reprod Med*. 2008;53(3):238-239; author reply 239.
- 164. Dudley NJ. A systematic review of the ultrasound estimation of fetal weight. *Ultrasound Obstet Gynecol*. 2005;25(1):80-89.
- 165. Milner J, Arezina J. The accuracy of ultrasound estimation of fetal weight in comparison to birth weight: a systematic review. *Ultrasound*. 2018;26(1):32-41.
- 166. Melamed N, Ben-Haroush A, Meizner I, Mashiach R, Glezerman M, Yogev Y. Accuracy of sonographic weight estimation as a function of fetal sex. *Ultrasound Obstet Gynecol*. 2011;38(1):67-73.
- 167. Melamed N, Ben-Haroush A, Meizner I, Mashiach R, Yogev Y, Pardo J. Accuracy of sonographic fetal weight estimation: a matter of presentation. *Ultrasound Obstet Gynecol*. 2011;38(4): 418-424.
- 168. Melamed N, Yogev Y, Meizner I, Mashiach R, Pardo J, Ben-Haroush A. Prediction of fetal macrosomia: effect of sonographic fetal weight-estimation model and threshold used. *Ultrasound Obstet Gynecol*. 2011;38(1):74-81.
- 169. Melamed N, Yogev Y, Ben-Haroush A, Meizner I, Mashiach R, Glezerman M. Does use of a sex-specific model improve the accuracy of sonographic weight estimation? *Ultrasound Obstet Gynecol*. 2012;39(5):549-557.
- 170. Melamed N, Yogev Y, Linder N, et al. Role of fetal length in the prediction of fetal weight. *J Ultrasound Med*. 2012;31(5):687-694.
- 171. Melamed N, Ryan G, Windrim R, Toi A, Kingdom J. Choice of formula and accuracy of fetal weight estimation in small-for-gestational-age fetuses. *J Ultrasound Med*. 2016;35(1):71-82.
- 172. Expert Panel on Women's Imaging, Shipp TD, Zelop CM, et al. ACR Appropriateness Criteria© growth disturbances-risk of fetal growth restriction. *J Am Coll Radiol*. 2019;16(5S):S116 -S125.
- 173. Lausman A, Kingdom J; Maternal Fetal Medicine C. Intrauterine growth restriction: screening, diagnosis, and management. *J Obstet Gynaecol Can*. 2013;35(8):741-748.
- 174. Bakalis S, Cao K, Johal N, Cuckow P, Pandya P. The value of the routine third trimester ultrasound scan in antenatal care: Problems with guidance and outdated data in a highly technological field. *Eur J Obstet Gynecol Reprod Biol*. 2020;245:51-55.
- 175. Lindqvist PG, Molin J. Does antenatal identification of small-for-gestational age fetuses significantly improve their outcome? *Ultrasound Obstet Gynecol*. 2005;25(3):258-264.
- 176. Ray CL, Grange G. Routine third trimester ultrasound in low risk pregnancy confers no benefit!: AGAINST: Arguments for a routine third trimester ultrasound: what the meta-analysis does not show!. *BJOG*. 2016;123(7):1122.
- 177. Thornton J. Routine third trimester ultrasound in low risk pregnancy confers no benefit!: FOR: The benefits of routine third-trimester scanning are less clear cut. *BJOG*. 2016;123(7):1121.
- 178. Bricker L, Medley N, Pratt JJ. Routine ultrasound in late pregnancy (after 24 weeks' gestation). *Cochrane Database Syst Rev*. 2015(6):CD001451.
- 179. Ashimi Balogun O, Sibai BM, Pedroza C, Blackwell SC, Barrett TL, Chauhan SP. Serial third-trimester ultrasonography compared with routine care in uncomplicated pregnancies: a randomized controlled trial. *Obstet Gynecol*. 2018;132(6):1358-1367.
- 180. Skrastad RB, Eik-Nes SH, Sviggum O, et al. A randomized controlled trial of third-trimester routine ultrasound in a non-selected population. *Acta Obstet Gynecol Scand*. 2013;92(12):1353-1360.
- 181. Sovio U, White IR, Dacey A, Pasupathy D, Smith GCS. Screening for fetal growth restriction with universal third trimester ultrasonography in nulliparous women in the Pregnancy Outcome Prediction (POP) study: a prospective cohort study. *Lancet*. 2015;386(10008):2089-2097.
- 182. Hiersch L, Melamed N. Fetal growth velocity and body proportion in the assessment of growth. *Am J Obstet Gynecol*. 2018;218(2S):pp. S700–S711.e1.
- 183. Miranda J, Rodriguez-Lopez M, Triunfo S, et al. Prediction of fetal growth restriction using estimated fetal weight vs a combined screening model in the third trimester. *Ultrasound Obstet Gynecol*. 2017;50(5):603-611.
- 184. De Castro H, Ciobanu A, Formuso C, Akolekar R, Nicolaides KH. Value of routine ultrasound examination at 35–37 weeks' gestation in diagnosis of non-cephalic presentation. *Ultrasound Obstet Gynecol*. 2020;55(2):248-256.
- 185. Ewigman BG, Crane JP, Frigoletto FD, LeFevre ML, Bain RP, McNellis D. Effect of prenatal ultrasound screening on perinatal outcome. *RADIUS Study Group. New Engl J Med*. 1993;329(12):821-827.
- 186. Ficara A, Syngelaki A, Hammami A, Akolekar R, Nicolaides KH. Value of routine ultrasound examination at 35–37 weeks' gestation in diagnosis of fetal abnormalities. *Ultrasound Obstet Gynecol*. 2020;55(1):75-80.
- 187. Ciobanu A, Rouvali A, Syngelaki A, Akolekar R, Nicolaides KH. Prediction of small for gestational age neonates: screening by maternal factors, fetal biometry, and biomarkers at 35–37 weeks' gestation. *Am J Obstet Gynecol*. 2019;220(5):486.e1-486.e11.
- 188. Martinez-Portilla RJ, Caradeux J, Meler E, Lip-Sosa DL, Sotiriadis A, Figueras F. Third-trimester uterine-artery Doppler for prediction of adverse outcome in late small-for-gestational-age fetuses: systematic review and meta-analysis. *Ultrasound Obstet Gynecol*. 2019;55(5):575-585.
- 189. Conde-Agudelo A, Villar J, Kennedy SH, Papageorghiou AT. Predictive accuracy of cerebroplacental ratio for adverse perinatal and neurodevelopmental outcomes in suspected fetal growth restriction: systematic review and meta-analysis. *Ultrasound Obstet Gynecol*. 2018;52(4):430-441.
- 190. Katz J, Wu LA, Mullany LC, et al. Prevalence of small-forgestational-age and its mortality risk varies by choice of birth-weightfor-gestation reference population. *PLoS One*. 2014;9(3):e92074.
- 191. Salomon LJ, Bernard JP, Duyme M, Buvat I, Ville Y. The impact of choice of reference charts and equations on the assessment of fetal biometry. *Ultrasound Obstet Gynecol*. 2005;25(6):559-565.
- 192. Alexander GR, Himes JH, Kaufman RB, Mor J, Kogan M. A United States national reference for fetal growth. *Obstet Gynecol*. 1996;87(2):163-168.
- 193. Hadlock FP, Harrist RB, Martinez-Poyer J. In utero analysis of fetal growth: a sonographic weight standard. *Radiology*. 1991;181(1):129-133.
- 194. Papageorghiou AT, Ohuma EO, Altman DG, et al. International standards for fetal growth based on serial ultrasound measurements: the Fetal Growth Longitudinal Study of the INTERGROWTH-21st Project. *Lancet*. 2014;384(9946):869-879.
- 195. Villar J, Papageorghiou AT, Pang R, et al. The likeness of fetal growth and newborn size across non-isolated populations in the INTERGROWTH-21st Project: the Fetal Growth Longitudinal

Study and Newborn Cross-Sectional Study. *Lancet Diabetes Endocrinol*. 2014;2(10):781-792.

- 196. Stirnemann J, Villar J, Salomon LJ, et al. International estimated fetal weight standards of the INTERGROWTH-21(st) Project. *Ultrasound Obstet Gynecol*. 2017;49(4):478-486.
- 197. Kiserud T, Piaggio G, Carroli G, et al. The World Health Organization fetal growth charts: a multinational longitudinal study of ultrasound biometric measurements and estimated fetal weight. *PLoS Med*. 2017;14(1):e1002220.
- 198. Buck Louis GM, Grewal J, Albert PS, et al. Racial/ethnic standards for fetal growth: the NICHD Fetal Growth Studies. *Am J Obstet Gynecol*. 2015;213(4):449.e41-449.e41.
- 199. Tarca AL, Romero R, Gudicha DW, et al. A new customized fetal growth standard for African American women: the PRB/NICHD Detroit study. *Am J Obstet Gynecol*. 2018;218(2S):pp. S679–S691. e4.
- 200. Gardosi J, Chang A, Kalyan B, Sahota D, Symonds EM. Customised antenatal growth charts. *Lancet*. 1992;339(8788):283-287.
- 201. Gardosi J, Francis A, Turner S, Williams M. Customized growth charts: rationale, validation and clinical benefits. *Am J Obstet Gynecol*. 2018;218(2S):S609-S618.
- 202. Deter RL. Individualized growth assessment predicts birth weight accurately. *Ultrasound Obstet Gynecol*. 1996;7(2):156-157.
- 203. Deter RL, Levytska K, Melamed N, Lee W, Kingdom JC. Classifying neonatal growth outcomes: use of birth weight, placental evaluation and individualized growth assessment. *J Matern Fetal Neonatal Med*. 2016;29(24):3939-3949.
- 204. Deter RL, Lee W, Yeo L, et al. Individualized growth assessment: conceptual framework and practical implementation for the evaluation of fetal growth and neonatal growth outcome. *Am J Obstet Gynecol*. 2018;218(2S):S656-S678.
- 205. Gardosi J. Fetal growth and ethnic variation. *Lancet Diabetes Endocrinol*. 2014;2(10):773-774.
- 206. Albert PS, Grantz KL. Fetal growth and ethnic variation. *Lancet Diabetes Endocrinol*. 2014;2(10):773.
- 207. WHO Multicentre Growth Reference Study Group. WHO Child Growth Standards based on length/height, weight and age. *Acta Paediatr Suppl*. 2006;450:76-85.
- 208. Kiserud T, Benachi A, Hecher K, et al. The World Health Organization fetal growth charts: concept, findings, interpretation, and application. *Am J Obstet Gynecol*. 2018;218(2S):S619 -S629.
- 209. Cheng Y, Leung TY, Lao T, Chan YM, Sahota DS. Impact of replacing Chinese ethnicity-specific fetal biometry charts with the INTERGROWTH-21(st) standard. *BJOG*. 2016;123(Suppl 3):48-55.
- 210. Poon LC, Tan MY, Yerlikaya G, Syngelaki A, Nicolaides KH. Birth weight in live births and stillbirths. *Ultrasound Obstet Gynecol*. 2016;48(5):602-606.
- 211. Anderson NH, Sadler LC, McKinlay CJD, McCowan LME. INTERGROWTH-21st vs customized birthweight standards for identification of perinatal mortality and morbidity. *Am J Obstet Gynecol*. 2016;214(4):509.e1-509.e7.
- 212. Melamed N, Ray JG, Shah PS, Berger H, Kingdom JC. Should we use customized fetal growth percentiles in urban Canada? *J Obstet Gynaecol Can*. 2014;36(2):164-170.
- 213. Gardosi J. Customized charts and their role in identifying pregnancies at risk because of fetal growth restriction. *J Obstet Gynaecol Can*. 2014;36(5):408-415.
- 214. Francis A, Hugh O, Gardosi J. Customized vs INTERGROWTH-21(st) standards for the assessment of birthweight and stillbirth risk at term. *Am J Obstet Gynecol*. 2018;218(2S):S692-S699.
- 215. Pritchard N, Lindquist A, Siqueira IDA, Walker SP, Permezel M. INTERGROWTH-21st compared with GROW customized centiles in the detection of adverse perinatal outcomes at term. *J Matern Fetal Neonatal Med*. 2020;33(6):961-966.
- 216. Ego A, Subtil D, Grange G, et al. Customized versus population-based birth weight standards for identifying growth restricted infants: a French multicenter study. *Am J Obstet Gynecol*. 2006;194(4):1042-1049.
- 217. Sovio U, Smith GCS. The effect of customization and use of a fetal growth standard on the association between birthweight percentile and adverse perinatal outcome. *Am J Obstet Gynecol*. 2018;218(2S):S738-S744.
- 218. Hutcheon JA, Walker M, Platt RW. Assessing the value of customized birth weight percentiles. *Am J Epidemiol*. 2011;173(4):459-467.
- 219. Hutcheon J. Do customized birth weight charts add anything but complexity to the assessment of fetal growth? *J Obstet Gynaecol Can*. 2014;36(2):107-109.
- 220. Mikolajczyk RT, Zhang J, Betran AP, et al. A global reference for fetal-weight and birthweight percentiles. *Lancet*. 2011;377(9780):1855-1861.
- 221. Pritchard NL, Hiscock RJ, Lockie E, et al. Identification of the optimal growth charts for use in a preterm population: An Australian state-wide retrospective cohort study. *PLoS Med*. 2019;16(10):e1002923.
- 222. Hiersch L, Okby R, Freeman H, et al. Differences in fetal growth patterns between twins and singletons. *J Matern Fetal Neonatal Med*. 2020;33(15):2546-2555.
- 223. Grantz KL, Grewal J, Albert PS, et al. Dichorionic twin trajectories: the NICHD Fetal Growth Studies. *Am J Obstet Gynecol*. 2016;215(2):221.e1-221.e16.
- 224. Blickstein I. Is it normal for multiples to be smaller than singletons? *Best Pract Res Clin Obstet Gynaecol*. 2004;18(4):613-623.
- 225. Stirrup OT, Khalil A, D'Antonio F, Thilaganathan B; Southwest Thames Obstetric Research Collaborative (STORK). Fetal growth reference ranges in twin pregnancy: analysis of the Southwest Thames Obstetric Research Collaborative (STORK) multiple pregnancy cohort. *Ultrasound Obstet Gynecol*. 2015;45(3):301-307.
- 226. Gielen M, Lindsey PJ, Derom C, et al. Twin-specific intrauterine 'growth' charts based on cross-sectional birthweight data. *Twin Res Hum Genet*. 2008;11(2):224-235.
- 227. Mendez-Figueroa H, Truong VTT, Pedroza C, Chauhan SP. Growth among twins: use of singleton versus twin-specific growth nomograms. *Am J Perinatol*. 2018;35(2):184-191.
- 228. Bleker OP, Wolf H, Oosting J. The placental cause of fetal growth retardation in twin gestations. *Acta Genet Med Gemellol*. 1995;44(2):103-106.
- 229. Vatnick I, Schoknecht PA, Darrigrand R, Bell AW. Growth and metabolism of the placenta after unilateral fetectomy in twin pregnant ewes. *J Dev Physiol*. 1991;15(6):351-356.
- 230. Liao AW, Brizot Mde L, Kang HJ, Assuncao RA, Zugaib M. Longitudinal reference ranges for fetal ultrasound biometry in twin pregnancies. *Clinics*. 2012;67(5):451-455.
- 231. Muhlhausler BS, Hancock SN, Bloomfield FH, Harding R. Are twins growth restricted? *Pediatr Res*. 2011;70(2):117-122.
- 232. Doom EC, Delbaere I, Martens G, Temmerman M. Birth weight for gestational age among Flemish twin population. *Facts Views Vis Obgyn*. 2012;4(1):42-49.
- 233. Dollberg S, Haklai Z, Mimouni FB, Gorfein I, Gordon ES. Birth weight standards in the live-born population in Israel. *Isr Med Assoc J*. 2005;7(5):311-314.
- 234. Min S-J, Luke B, Gillespie B, et al. Birth weight references for twins. *Am J Obstet Gynecol*. 2000;182(5):1250-1257.
- 235. Ong S, Lim MN, Fitzmaurice A, Campbell D, Smith AP, Smith N. The creation of twin centile curves for size. *BJOG*. 2002;109(7):753-758.
- 236. FIGO Working Group on Good Clinical Practice in Maternal-Fetal Medicine. Good clinical practice advice: Role of ultrasound in the management of twin pregnancy. *Int J Gynecol Obstet*. 2019;144(3):338-339.
- 237. Committee on Practice Bulletins—Obstetrics; Society for Maternal-Fetal Medicine. Practice bulletin no. 169: multifetal

gestations: twin, triplet, and higher-order multifetal pregnancies. *Obstet Gynecol*. 2016;128(4):e131-e146.

- 238. Morin L, Lim KN. 260-Ultrasound in twin pregnancies. *J Obstet Gynaecol Can*. 2017;39(10):e398-e411.
- 239. Khalil A, Rodgers M, Baschat A, et al. ISUOG practice guidelines: role of ultrasound in twin pregnancy. *Ultrasound Obstet Gynecol*. 2016;47(2):247-263.
- 240. Alexander JM, Hammond KR, Steinkampf MP. Multifetal reduction of high-order multiple pregnancy: comparison of obstetrical outcome with nonreduced twin gestations. *Fertil Steril*. 1995;64(6):1201-1203.
- 241. Begum G, Stevens A, Smith EB, et al. Epigenetic changes in fetal hypothalamic energy regulating pathways are associated with maternal undernutrition and twinning. *FASEB J*. 2012;26(4):1694-1703.
- 242. Tsai PC, Van Dongen J, Tan Q, et al. DNA methylation changes in the IGF1R gene in birth weight discordant adult monozygotic twins. *Twin Res Hum Genet*. 2015;18(6):635-646.
- 243. Williams-Wyss O, Zhang S, MacLaughlin SM, et al. Embryo number and periconceptional undernutrition in the sheep have differential effects on adrenal epigenotype, growth, and development. *Am J Physiol Endocrinol Metab*. 2014;307(2):E141-150.
- 244. Kalafat E, Sebghati M, Thilaganathan B, Khalil A; Southwest Thames Obstetric Research Collaborative. Predictive accuracy of Southwest Thames Obstetric Research Collaborative (STORK) chorionicity-specific twin growth charts for stillbirth: a validation study. *Ultrasound Obstet Gynecol*. 2019;53(2):193-199.
- 245. Odibo AO, McDonald RE, Stamilio DM, Ural SH, Macones GA. Perinatal outcomes in growth-restricted twins compared with age-matched growth-restricted singletons. *Am J Perinatol*. 2005;22(5):269-273.
- 246. Joseph KS, Fahey J, Platt RW, et al. An outcome-based approach for the creation of fetal growth standards: do singletons and twins need separate standards? *Am J Epidemiol*. 2009;169(5):616-624.
- 247. Herlihy N, Odom E, Cohen N, Stroustrup A, Rebarber A, Fox NS. Long-term outcomes of small for gestational age twins born at 34 weeks or later. *Am J Perinatol*. 2018;35(3):254-261.
- 248. Kibel M, Kahn M, Sherman C, et al. Placental abnormalities differ between small for gestational age fetuses in dichorionic twin and singleton pregnancies. *Placenta*. 2017;60:28-35.
- 249. Barber E, Weiner E, Feldstein O, et al. The differences in placental pathology and neonatal outcome in singleton vs. twin gestation complicated by small for gestational age. *Arch Gynecol Obstet*. 2018;298(6):1107-1114.
- 250. D'Antonio F, Odibo AO, Prefumo F, et al. Weight discordance and perinatal mortality in twin pregnancy: systematic review and meta-analysis. *Ultrasound Obstet Gynecol*. 2018;52(1):11-23.
- 251. Lean SC, Derricott H, Jones RL, Heazell AEP. Advanced maternal age and adverse pregnancy outcomes: a systematic review and meta-analysis. *PLoS One*. 2017;12(10):e0186287.
- 252. Yogev Y, Melamed N, Bardin R, Tenenbaum-Gavish K, Ben-Shitrit G, Ben-Haroush A. Pregnancy outcome at extremely advanced maternal age. *Am J Obstet Gynecol*. 2010;203(6):558.e1-7.
- 253. Shamu S, Abrahams N, Temmerman M, Musekiwa A, Zarowsky C. A systematic review of African studies on intimate partner violence against pregnant women: prevalence and risk factors. *PLoS One*. 2011;6(3):e17591.
- 254. Felker-Kantor E, Wallace M, Theall K. Living in violence: Neighborhood domestic violence and small for gestational age births. *Health Place*. 2017;46:130-136.
- 255. Saccone G, Berghella V, Sarno L, et al. Celiac disease and obstetric complications: a systematic review and metaanalysis. *Am J Obstet Gynecol*. 2016;214(2):225-234.
- 256. Yee LM, Silver RM, Haas DM, et al. Quality of periconceptional dietary intake and maternal and neonatal outcomes. *Am J Obstet Gynecol*. 2020;223(1):121.e1-121.e8.
- 257. Black RE, Victora CG, Walker SP, et al. Maternal and child undernutrition and overweight in low-income and middle-income countries. *Lancet*. 2013;382(9890):427-451.
- 258. Luke S, Kirby RS. Timing of maternal tobacco exposure, hypertension, and risk of singleton small-for-gestational age infants. *Am J Perinatol*. 2018;35(3):215-219.
- 259. Tong VT, England LJ, Rockhill KM, D'Angelo DV. Risks of preterm delivery and small for gestational age infants: effects of nondaily and low-intensity daily smoking during pregnancy. *Paediatr Perinat Epidemiol*. 2017;31(2):144-148.
- 260. Mahendru AA, Daemen A, Everett TR, et al. Impact of ovulation and implantation timing on first-trimester crown-rump length and gestational age. *Ultrasound Obstet Gynecol*. 2012;40(6): 630-635.
- 261. Saito M, Yazawa K, Hashiguchi A, Kumasaka T, Nishi N, Kato K. Time of ovulation and prolonged pregnancy. *Am J Obstet Gynecol*. 1972;112(1):31-38.
- 262. Hadlock FP, Shah YP, Kanon DJ, Lindsey JV. Fetal crown-rump length: reevaluation of relation to menstrual age (5–18 weeks) with high-resolution real-time US. *Radiology*. 1992;182(2): 501-505.
- 263. Daya S. Accuracy of gestational age estimation by means of fetal crown-rump length measurement. *Am J Obstet Gynecol*. 1993;168(3 Pt 1):903-908.
- 264. Altman DG, Chitty LS. New charts for ultrasound dating of pregnancy. *Ultrasound Obstet Gynecol*. 1997;10(3):174-191.
- 265. Savitz DA, Terry JW Jr, Dole N, Thorp JM Jr, Siega-Riz AM, Herring AH. Comparison of pregnancy dating by last menstrual period, ultrasound scanning, and their combination. *Am J Obstet Gynecol*. 2002;187(6):1660-1666.
- 266. Bottomley C, Bourne T. Dating and growth in the first trimester. *Best Pract Res Clin Obstet Gynaecol*. 2009;23(4):439-452.
- 267. Piantelli G, Sacchini C, Coltri A, Ludovici G, Paita Y, Gramellini D. Ultrasound dating-curve analysis in the assessment of gestational age. *Clin Exp Obstet Gynecol*. 1994;21(2):108-118.
- 268. Butt K, Lim KI. Guideline no. 388-determination of gestational age by ultrasound. *J Obstet Gynaecol Can*. 2019;41(10):1497-1507.
- 269. Agathokleous M, Chaveeva P, Poon LC, Kosinski P, Nicolaides KH. Meta-analysis of second-trimester markers for trisomy 21. *Ultrasound Obstet Gynecol*. 2013;41(3):247-261.
- 270. Sagi-Dain L, Maya I, Reches A, et al. Chromosomal microarray analysis results from pregnancies with various ultrasonographic anomalies. *Obstet Gynecol*. 2018;132(6):1368-1375.
- 271. Papageorghiou AT, Fratelli N, Leslie K, Bhide A, Thilaganathan B. Outcome of fetuses with antenatally diagnosed short femur. *Ultrasound Obstet Gynecol*. 2008;31(5):507-511.
- 272. Krakow D, Rimoin DL. The skeletal dysplasias. *Genet Med*. 2010;12(6):327-341.
- 273. D'Ambrosio V, Vena F, Marchetti C, et al. Midtrimester isolated short femur and perinatal outcomes: a systematic review and meta-analysis. *Acta Obstet Gynecol Scand*. 2019;98(1):11-17.
- 274. Khalil A, Sotiriadis A, Chaoui R, et al. ISUOG Practice Guidelines: role of ultrasound in congenital infection. *Ultrasound Obstet Gynecol*. 2020;56(1):128-151.
- 275. Baschat AA, Weiner CP. Umbilical artery doppler screening for detection of the small fetus in need of antepartum surveillance. *Am J Obstet Gynecol*. 2000;182(1 Pt 1):154-158.
- 276. Baschat AA. Pathophysiology of fetal growth restriction: implications for diagnosis and surveillance. *Obstet Gynecol Surv*. 2004;59(8):617-627.
- 277. Trudinger BJ, Cook CM. Umbilical and uterine artery flow velocity waveforms in pregnancy associated with major fetal abnormality. *Br J Obstet Gynaecol*. 1985;92(7):666-670.
- 278. Rochelson B, Schulman H, Farmakides G, et al. The significance of absent end-diastolic velocity in umbilical artery velocity waveforms. *Am J Obstet Gynecol*. 1987;156(5):1213-1218.

52 ALLER GUNEOLOGY 6. ALLER CONFIDENTIAL

- 279. Hata K, Hata T, Senoh D, Aoki S, Takamiya O, Kitao M. Umbilical artery blood flow velocity waveforms and associations with fetal abnormality. *Gynecol Obstet Invest*. 1989;27(4):179-182.
- 280. Griffin M, Seed PT, Webster L, et al. Diagnostic accuracy of placental growth factor and ultrasound parameters to predict the small-for-gestational-age infant in women presenting with reduced symphysis-fundus height. *Ultrasound Obstet Gynecol*. 2015;46(2):182-190.
- 281. Yinon Y, Farine D, No YMH. 240-Cytomegalovirus infection in pregnancy. *J Obstet Gynaecol Can*. 2018;40(2):e134-e141.
- 282. Enders M, Daiminger A, Exler S, Ertan K, Enders G, Bald R. Prenatal diagnosis of congenital cytomegalovirus infection in 115 cases: a 5 years' single center experience. *Prenat Diagn*. 2017;37(4):389-398.
- 283. Peng R, Yang J, Xie HN, Lin MF, Zheng J. Chromosomal and subchromosomal anomalies associated to small for gestational age fetuses with no additional structural anomalies. *Prenat Diagn*. 2017;37(12):1219-1224.
- 284. Brun S, Pennamen P, Mattuizzi A, et al. Interest of chromosomal microarray analysis in the prenatal diagnosis of fetal intrauterine growth restriction. *Prenat Diagn*. 2018;38(13):1111-1119.
- 285. Ma Y, Pei Y, Yin C, et al. Subchromosomal anomalies in small for gestational-age fetuses and newborns. *Arch Gynecol Obstet*. 2019;300(3):633-639.
- 286. Ferrazzi E, Bozzo M, Rigano S, et al. Temporal sequence of abnormal Doppler changes in the peripheral and central circulatory systems of the severely growth-restricted fetus. *Ultrasound Obstet Gynecol*. 2002;19(2):140-146.
- 287. Cosmi E, Ambrosini G, D'Antona D, Saccardi C, Mari G. Doppler, cardiotocography, and biophysical profile changes in growth-restricted fetuses. *Obstet Gynecol*. 2005;106(6):1240-1245.
- 288. Baschat AA, Gembruch U, Harman CR. The sequence of changes in Doppler and biophysical parameters as severe fetal growth restriction worsens. *Ultrasound Obstet Gynecol*. 2001;18(6):571-577.
- 289. Hecher K, Bilardo CM, Stigter RH, et al. Monitoring of fetuses with intrauterine growth restriction: a longitudinal study. *Ultrasound Obstet Gynecol*. 2001;18(6):564-570.
- 290. Unterscheider J, Daly S, Geary MP, et al. Predictable progressive Doppler deterioration in IUGR: does it really exist? *Am J Obstet Gynecol*. 2013;209(6):539.e1-7.
- 291. Turan OM, Turan S, Gungor S, et al. Progression of Doppler abnormalities in intrauterine growth restriction. *Ultrasound Obstet Gynecol*. 2008;32(2):160-167.
- 292. Crimmins S, Desai A, Block-Abraham D, Berg C, Gembruch U, Baschat AA. A comparison of Doppler and biophysical findings between liveborn and stillborn growth-restricted fetuses. *Am J Obstet Gynecol*. 2014;211(6):669.e1-10.
- 293. Bilardo CM, Wolf H, Stigter RH, et al. Relationship between monitoring parameters and perinatal outcome in severe, early intrauterine growth restriction. *Ultrasound Obstet Gynecol*. 2004;23(2):119-125.
- 294. Group GS. A randomised trial of timed delivery for the compromised preterm fetus: short term outcomes and Bayesian interpretation. *BJOG*. 2003;110(1):27-32.
- 295. Thornton JG, Hornbuckle J, Vail A, Spiegelhalter DJ, Levene M. GRIT study group. Infant wellbeing at 2 years of age in the Growth Restriction Intervention Trial (GRIT): multicentred randomised controlled trial. *Lancet*. 2004;364(9433):513-520.
- 296. Lees C, Marlow N, Arabin B, et al. Perinatal morbidity and mortality in early-onset fetal growth restriction: cohort outcomes of the trial of randomized umbilical and fetal flow in Europe (TRUFFLE). *Ultrasound Obstet Gynecol*. 2013;42(4):400-408.
- 297. Sharp A, Jackson R, Cornforth C, et al. A prediction model for short-term neonatal outcomes in severe early-onset fetal growth restriction. *Eur J Obstet Gynecol Reprod Biol*. 2019;241:109-118.
- 298. Oros D, Figueras F, Cruz-Martinez R, Meler E, Munmany M, Gratacos E. Longitudinal changes in uterine, umbilical and fetal cerebral Doppler indices in late-onset small-for-gestational age fetuses. *Ultrasound Obstet Gynecol*. 2011;37(2):191-195.
- 299. Figueras F, Gardosi J. Intrauterine growth restriction: new concepts in antenatal surveillance, diagnosis, and management. *Am J Obstet Gynecol*. 2011;204(4):288-300.
- 300. Chauhan SP, Rice MM, Grobman WA, et al. Neonatal morbidity of small- and large-for-gestational-age neonates born at term in uncomplicated pregnancies. *Obstet Gynecol*. 2017;130(3):511-519.
- 301. Frusca T, Todros T, Lees C, Bilardo CM, Investigators T. Outcome in early-onset fetal growth restriction is best combining computerized fetal heart rate analysis with ductus venosus Doppler: insights from the Trial of Umbilical and Fetal Flow in Europe. *Am J Obstet Gynecol*. 2018;218(2S):S783-S789.
- 302. Manning FA, Snijders R, Harman CR, Nicolaides K, Menticoglou S, Morrison I. Fetal biophysical profile score. VI. Correlation with antepartum umbilical venous fetal pH. *Am J Obstet Gynecol*. 1993;169(4):755-763.
- 303. Vintzileos AM, Fleming AD, Scorza WE, et al. Relationship between fetal biophysical activities and umbilical cord blood gas values. *Am J Obstet Gynecol*. 1991;165(3):707-713.
- 304. Baschat AA, Galan HL, Bhide A, et al. Doppler and biophysical assessment in growth restricted fetuses: distribution of test results. *Ultrasound Obstet Gynecol*. 2006;27(1):41-47.
- 305. Tuffnell DJ, Cartmill RS, Lilford RJ. Fetal movements; factors affecting their perception. *Eur J Obstet Gynecol Reprod Biol*. 1991;39(3):165-167.
- 306. Moore TR, Piacquadio K. A prospective evaluation of fetal movement screening to reduce the incidence of antepartum fetal death. *Am J Obstet Gynecol*. 1989;160(5 Pt 1):1075-1080.
- 307. Norman JE, Heazell AEP, Rodriguez A, et al. Awareness of fetal movements and care package to reduce fetal mortality (AFFIRM): a stepped wedge, cluster-randomised trial. *Lancet*. 2018;392(10158):1629-1638.
- 308. Tveit JV, Saastad E, Stray-Pedersen B, et al. Reduction of late stillbirth with the introduction of fetal movement information and guidelines - a clinical quality improvement. *BMC Pregnancy Childbirth*. 2009;9:32.
- 309. Saastad E, Winje BA, Stray Pedersen B, Froen JF. Fetal movement counting improved identification of fetal growth restriction and perinatal outcomes–a multi-centre, randomized, controlled trial. *PLoS One*. 2011;6(12):e28482.
- 310. Unterscheider J, O'Donoghue K, Malone FD. Guidelines on fetal growth restriction: a comparison of recent national publications. *Am J Perinatol*. 2015;32(4):307-316.
- 311. ACOG practice bulletin no. 204 summary: fetal growth restriction. *Obstet Gynecol*. 2019;133(2):390-392.
- 312. Vayssiere C, Sentilhes L, Ego A, et al. Fetal growth restriction and intra-uterine growth restriction: guidelines for clinical practice from the French College of Gynaecologists and Obstetricians. *Eur J Obstet Gynecol Reprod Biol*. 2015;193:10-18.
- 313. Lavin JP Jr, Miodovnik M, Barden TP. Relationship of nonstress test reactivity and gestational age. *Obstet Gynecol*. 1984;63(3):338-344.
- 314. Cousins LM, Poeltler DM, Faron S, Catanzarite V, Daneshmand S, Casele H. Nonstress testing at ≤32.0 weeks' gestation: a randomized trial comparing different assessment criteria. *Am J Obstet Gynecol*. 2012;207(4):pp. 311, e1–7.
- 315. Practice bulletin no. 145: antepartum fetal surveillance. *Obstet Gynecol*. 2014;124(1):182-192.
- 316. Macones GA, Hankins GD, Spong CY, Hauth J, Moore T. The 2008 National Institute of Child Health and Human Development workshop report on electronic fetal monitoring: update on definitions, interpretation, and research guidelines. *Obstet Gynecol*. 2008;112(3):661-666.
- 317. Freeman RK, Anderson G, Dorchester W. A prospective multiinstitutional study of antepartum fetal heart rate monitoring. I. Risk of perinatal mortality and morbidity according to antepartum fetal heart rate test results. *Am J Obstet Gynecol*. 1982;143(7):771-777.
- 318. Grivell RM, Alfirevic Z, Gyte GM, Devane D. Antenatal cardiotocography for fetal assessment. *Cochrane Database Syst Rev*. 2012;(12):CD007863.
- 319. Turan S, Turan OM, Berg C, et al. Computerized fetal heart rate analysis, Doppler ultrasound and biophysical profile score in the prediction of acid-base status of growth-restricted fetuses. *Ultrasound Obstet Gynecol*. 2007;30(5):750-756.
- 320. Pardey J, Moulden M, Redman CW. A computer system for the numerical analysis of nonstress tests. *Am J Obstet Gynecol*. 2002;186(5):1095-1103.
- 321. Wolf H, Arabin B, Lees CC, et al. Longitudinal study of computerized cardiotocography in early fetal growth restriction. *Ultrasound Obstet Gynecol*. 2017;50(1):71-78.
- 322. Serra V, Moulden M, Bellver J, Redman CW. The value of the shortterm fetal heart rate variation for timing the delivery of growthretarded fetuses. *BJOG*. 2008;115(9):1101-1107.
- 323. Lobmaier SM, Huhn EA, Pildner von Steinburg S, et al. Phaserectified signal averaging as a new method for surveillance of growth restricted fetuses. *J Matern Fetal Neonatal Med*. 2012;25(12):2523-2528.
- 324. Manning FA, Harman CR, Morrison I, Menticoglou SM, Lange IR, Johnson JM. Fetal assessment based on fetal biophysical profile scoring. IV. An analysis of perinatal morbidity and mortality. *Am J Obstet Gynecol*. 1990;162(3):703-709.
- 325. Nabhan AF, Abdelmoula YA. Amniotic fluid index versus single deepest vertical pocket as a screening test for preventing adverse pregnancy outcome. *Cochrane Database Syst Rev*. 2008(3):CD006593.
- 326. Chauhan SP, Sanderson M, Hendrix NW, Magann EF, Devoe LD. Perinatal outcome and amniotic fluid index in the antepartum and intrapartum periods: a meta-analysis. *Am J Obstet Gynecol*. 1999;181(6):1473-1478.
- 327. Clark SL, Sabey P, Jolley K. Nonstress testing with acoustic stimulation and amniotic fluid volume assessment: 5973 tests without unexpected fetal death. *Am J Obstet Gynecol*. 1989;160(3):694-697.
- 328. Morrow RJ, Adamson SL, Bull SB, Ritchie JW. Effect of placental embolization on the umbilical arterial velocity waveform in fetal sheep. *Am J Obstet Gynecol*. 1989;161(4):1055-1060.
- 329. Chang TC, Robson SC, Spencer JA, Gallivan S. Prediction of perinatal morbidity at term in small fetuses: comparison of fetal growth and Doppler ultrasound. *Br J Obstet Gynaecol*. 1994;101(5):422-427.
- 330. Vergani P, Andreotti C, Roncaglia N, et al. Doppler predictors of adverse neonatal outcome in the growth restricted fetus at 34 weeks' gestation or beyond. *Am J Obstet Gynecol*. 2003;189(4):1007-1011.
- 331. Bahado-Singh RO, Kovanci E, Jeffres A, et al. The Doppler cerebroplacental ratio and perinatal outcome in intrauterine growth restriction. *Am J Obstet Gynecol*. 1999;180(3 Pt 1):750-756.
- 332. Caradeux J, Martinez-Portilla RJ, Basuki TR, Kiserud T, Figueras F. Risk of fetal death in growth-restricted fetuses with umbilical and/ or ductus venosus absent or reversed end-diastolic velocities before 34 weeks of gestation: a systematic review and meta-analysis. *Am J Obstet Gynecol*. 2018;218(2S):pp. S774–S782.e21.
- 333. Hershkovitz R, Kingdom JC, Geary M, Rodeck CH. Fetal cerebral blood flow redistribution in late gestation: identification of compromise in small fetuses with normal umbilical artery Doppler. *Ultrasound Obstet Gynecol*. 2000;15(3):209-212.
- 334. Severi FM, Bocchi C, Visentin A, et al. Uterine and fetal cerebral Doppler predict the outcome of third-trimester small-for-gestational age fetuses with normal umbilical artery Doppler. *Ultrasound Obstet Gynecol*. 2002;19(3):225-228.
- 335. Cruz-Martinez R, Figueras F, Oros D, et al. Cerebral blood perfusion and neurobehavioral performance in full-term small-for-gestational-age fetuses. *Am J Obstet Gynecol*. 2009;201(5):474.e1-7.
- 336. Arbeille P, Maulik D, Fignon A, et al. Assessment of the fetal PO2 changes by cerebral and umbilical Doppler on lamb fetuses during acute hypoxia. *Ultrasound Med Biol*. 1995;21(7):861-870.
- 337. Vollgraff Heidweiller-Schreurs CA, van Osch IR, Heymans MW, et al. Cerebroplacental ratio in predicting adverse perinatal outcome: a meta-analysis of individual participant data. *BJOG*. 2020 May 3. [Epub ahead of print].
- 338. Bellotti M, Pennati G, De Gasperi C, Bozzo M, Battaglia FC, Ferrazzi E. Simultaneous measurements of umbilical venous, fetal hepatic, and ductus venosus blood flow in growth-restricted human fetuses. *Am J Obstet Gynecol*. 2004;190(5):1347-1358.
- 339. Baschat AA, Guclu S, Kush ML, Gembruch U, Weiner CP, Harman CR. Venous Doppler in the prediction of acid-base status of growth-restricted fetuses with elevated placental blood flow resistance. *Am J Obstet Gynecol*. 2004;191(1):277-284.
- 340. Rizzo G, Capponi A, Talone PE, Arduini D, Romanini C. Doppler indices from inferior vena cava and ductus venosus in predicting pH and oxygen tension in umbilical blood at cordocentesis in growth-retarded fetuses. *Ultrasound Obstet Gynecol*. 1996;7(6):401-410.
- 341. Turan OM, Turan S, Berg C, et al. Duration of persistent abnormal ductus venosus flow and its impact on perinatal outcome in fetal growth restriction. *Ultrasound Obstet Gynecol*. 2011;38(3):295-302.
- 342. Divon MY, Girz BA, Lieblich R, Langer O. Clinical management of the fetus with markedly diminished umbilical artery end-diastolic flow. *Am J Obstet Gynecol*. 1989;161(6 Pt 1):1523-1527.
- 343. Lees CC, Marlow N, van Wassenaer-Leemhuis A, et al. 2 year neurodevelopmental and intermediate perinatal outcomes in infants with very preterm fetal growth restriction (TRUFFLE): a randomised trial. *Lancet*. 2015;385(9983):2162-2172.
- 344. Baschat AA. Integrated fetal testing in growth restriction: combining multivessel Doppler and biophysical parameters. *Ultrasound Obstet Gynecol*. 2003;21(1):1-8.
- 345. Wallenstein MB, Birnie KL, Arain YH, et al. Failed endotracheal intubation and adverse outcomes among extremely low birth weight infants. *J Perinatol*. 2016;36(2):112-115.
- 346. Genzel-Boroviczeny O, Hempelman J, Zoppelli L, Martinez A. Predictive value of the 1-min Apgar score for survival at 23–26 weeks gestational age. *Acta Paediatr*. 2010;99(12):1790-1794.
- 347. Boers KE, Vijgen SM, Bijlenga D, et al. Induction versus expectant monitoring for intrauterine growth restriction at term: randomised equivalence trial (DIGITAT). *BMJ*. 2010;341:c7087.
- 348. Boers KE, van Wyk L, van der Post JAM, et al. Neonatal morbidity after induction vs expectant monitoring in intrauterine growth restriction at term: a subanalysis of the DIGITAT RCT. *Am J Obstet Gynecol*. 2012;206(4):344.e1-7.
- 349. Trudell AS, Cahill AG, Tuuli MG, Macones GA, Odibo AO. Risk of stillbirth after 37 weeks in pregnancies complicated by small-for-gestational-age fetuses. *Am J Obstet Gynecol*. 2013;208(5):376.e1-7.
- 350. van Wyk L, Boers KE, van der Post JA, et al. Effects on (neuro) developmental and behavioral outcome at 2 years of age of induced labor compared with expectant management in intrauterine growth-restricted infants: long-term outcomes of the DIGITAT trial. *Am J Obstet Gynecol*. 2012;206(5):406.e1-7.
- 351. Poulain P, Palaric JC, Milon J, et al. Absent end diastolic flow of umbilical artery Doppler: pregnancy outcome in 62 cases. *Eur J Obstet Gynecol Reprod Biol*. 1994;53(2):115-119.
- 352. Melamed N, Yogev Y, Hadar E, Hod M, Ben-Haroush A. Preinduction cervical ripening with prostaglandin E2 at preterm. *Acta Obstet Gynecol Scand*. 2008;87(1):63-67.
- 353. Sievert RA, Kuper SG, Jauk VC, Parrish M, Biggio JR, Harper LM. Predictors of vaginal delivery in medically indicated early preterm

induction of labor. *Am J Obstet Gynecol*. 2017;217(3):375.e1-375. e7.

- 354. Cruz-Lemini M, Crispi F, Van Mieghem T, et al. Risk of perinatal death in early-onset intrauterine growth restriction according to gestational age and cardiovascular Doppler indices: a multicenter study. *Fetal Diagn Ther*. 2012;32(1–2):116-122.
- 355. Figueras F, Savchev S, Triunfo S, Crovetto F, Gratacos E. An integrated model with classification criteria to predict small-for-gestational-age fetuses at risk of adverse perinatal outcome. *Ultrasound Obstet Gynecol*. 2015;45(3):279-285.
- 356. Garcia-Simon R, Figueras F, Savchev S, Fabre E, Gratacos E, Oros D. Cervical condition and fetal cerebral Doppler as determinants of adverse perinatal outcome after labor induction for late-onset small-for-gestational-age fetuses. *Ultrasound Obstet Gynecol*. 2015;46(6):713-717.
- 357. Familiari A, Khalil A, Rizzo G, et al. Adverse intrapartum outcome in pregnancies complicated by small for gestational age and late fetal growth restriction undergoing induction of labor with Dinoprostone, Misoprostol or mechanical methods: a systematic review and meta-analysis. *Eur J Obstet Gynecol Reprod Biol*. 2020;252:455-467.
- 358. Khong TY, Mooney EE, Ariel I, et al. Sampling and definitions of placental lesions: Amsterdam Placental Workshop Group Consensus Statement. *Arch Pathol Lab Med*. 2016;140(7):698-713.
- 359. Khong TY, Ting M, Gordijn SJ. Placental pathology and clinical trials: Histopathology data from prior and study pregnancies may improve analysis. *Placenta*. 2017;52:58-61.
- 360. Levy M, Alberti D, Kovo M, et al. Placental pathology in pregnancies complicated by fetal growth restriction: recurrence vs. new onset. *Arch Gynecol Obstet*. 2020;301(6):1397-1404.
- 361. Economides DL, Nicolaides KH, Linton EA, Perry LA, Chard T. Plasma cortisol and adrenocorticotropin in appropriate and small for gestational age fetuses. *Fetal Ther*. 1988;3(3):158-164.
- 362. Shams M, Kilby MD, Somerset DA, et al. 11Beta-hydroxysteroid dehydrogenase type 2 in human pregnancy and reduced expression in intrauterine growth restriction. *Hum Reprod*. 1998;13(4):799-804.
- 363. McTernan CL, Draper N, Nicholson H, et al. Reduced placental 11beta-hydroxysteroid dehydrogenase type 2 mRNA levels in human pregnancies complicated by intrauterine growth restriction: an analysis of possible mechanisms. *J Clin Endocrinol Metab*. 2001;86(10):4979-4983.
- 364. Morrison JL. Sheep models of intrauterine growth restriction: fetal adaptations and consequences. *Clin Exp Pharmacol Physiol*. 2008;35(7):730-743.
- 365. Beltrand J, Verkauskiene R, Nicolescu R, et al. Adaptive changes in neonatal hormonal and metabolic profiles induced by fetal growth restriction. *J Clin Endocrinol Metab*. 2008;93(10):4027-4032.
- 366. Rizzo G, Capponi A, Cavicchioni O, Vendola M, Arduini D. Low cardiac output to the placenta: an early hemodynamic adaptive mechanism in intrauterine growth restriction. *Ultrasound Obstet Gynecol*. 2008;32(2):155-159.
- 367. Turan S, Miller J, Baschat AA. Integrated testing and management in fetal growth restriction. *Semin Perinatol*. 2008;32(3):194-200.
- 368. Hubinont C, Nicolini U, Fisk NM, Tannirandorn Y, Rodeck CH. Endocrine pancreatic function in growth-retarded fetuses. *Obstet Gynecol*. 1991;77(4):541-544.
- 369. Ville Y, Proudler A, Kuhn P, Nicolaides KH. Aldosterone concentration in normal, growth-retarded, anemic, and hydropic fetuses. *Obstet Gynecol*. 1994;84(4):511-514.
- 370. Wallace EM, Baker LS. Effect of antenatal betamethasone administration on placental vascular resistance. *Lancet*. 1999;353(9162):1404-1407.
- 371. Senat MV, Ville Y. Effect of steroids on arterial Doppler in intrauterine growth retardation fetuses. *Fetal Diagn Ther*. 2000;15(1): 36-40.
- 372. Simchen MJ, Alkazaleh F, Adamson SL, et al. The fetal cardiovascular response to antenatal steroids in severe early-onset intrauterine growth restriction. *Am J Obstet Gynecol*. 2004;190(2):296-304.
- 373. Robertson MC, Murila F, Tong S, Baker LS, Yu VY, Wallace EM. Predicting perinatal outcome through changes in umbilical artery Doppler studies after antenatal corticosteroids in the growthrestricted fetus. *Obstet Gynecol*. 2009;113(3):636-640.
- 374. Wijnberger LD, Bilardo CM, Hecher K, Stigter RH, Visser GH. Effect of antenatal glucocorticoid therapy on arterial and venous blood flow velocity waveforms in severely growth-restricted fetuses. *Ultrasound Obstet Gynecol*. 2004;23(6):584-589.
- 375. Mulder EJ, de Heus R, Visser GH. Antenatal corticosteroid therapy: short-term effects on fetal behaviour and haemodynamics. *Semin Fetal Neonatal Med*. 2009;14(3):151-156.
- 376. Piazze J, Dillon KC, Cerekja A. Betamethasone effects on umbilical arteries and ductus venosus Doppler velocity waveforms in growth-restricted fetuses. *J Matern Fetal Neonatal Med*. 2012;25(7):1179-1182.
- 377. McMillen IC, Adams MB, Ross JT, et al. Fetal growth restriction: adaptations and consequences. *Reproduction*. 2001;122(2):195-204.
- 378. Melamed N, Pittini A, Barrett J, et al. Antenatal corticosteroids and outcomes of small-for-gestational-age neonates. *Obstet Gynecol*. 2016;128(5):1001-1008.
- 379. Ting JY, Kingdom JC, Shah PS. Antenatal glucocorticoids, magnesium sulfate, and mode of birth in preterm fetal small for gestational age. *Am J Obstet Gynecol*. 2018;218(2S):S818-S828.
- 380. Melamed N, Shah J, Soraisham A, et al. Association between antenatal corticosteroid administration-to-birth interval and outcomes of preterm neonates. *Obstet Gynecol*. 2015;125(6):1377-1384.
- 381. Cahill LS, Whitehead CL, Hobson SR, et al. Effect of maternal betamethasone administration on feto-placental vascular resistance in the mousedagger. *Biol Reprod*. 2019;101(4):823-831.
- 382. Rouse DJ, Hirtz DG, Thom E, et al. A randomized, controlled trial of magnesium sulfate for the prevention of cerebral palsy. *New Engl J Med*. 2008;359(9):895-905.
- 383. Doyle LW, Crowther CA, Middleton P, Marret S, Rouse D. Magnesium sulphate for women at risk of preterm birth for neuroprotection of the fetus. *Cochrane Database Syst Rev*. 2009(1):CD004661.
- 384. Marret S, Doyle LW, Crowther CA, Middleton P. Antenatal magnesium sulphate neuroprotection in the preterm infant. *Semin Fetal Neonatal Med*. 2007;12(4):311-317.
- 385. Costantine MM, Drever N. Antenatal exposure to magnesium sulfate and neuroprotection in preterm infants. *Obstet Gynecol Clin North Am*. 2011;38(2):351-366, xi.
- 386. American College of Obstetricians and Gynecologists Committee on Obstetric Practice; Society for Maternal-Fetal Medicine. Committee Opinion No. 455: Magnesium sulfate before anticipated preterm birth for neuroprotection. *Obstet Gynecol*. 2010;115(3):669-671.
- 387. Reeves SA, Gibbs RS, Clark SL. Magnesium for fetal neuroprotection. *Am J Obstet Gynecol*. 2011;204(3):202.e1-4.
- 388. Raju TN, Mercer BM, Burchfield DJ, Joseph GF Jr. Periviable birth: executive summary of a joint workshop by the Eunice Kennedy Shriver National Institute of Child Health and Human Development, Society for Maternal-Fetal Medicine, American Academy of Pediatrics, and American College of Obstetricians and Gynecologists. *Am J Obstet Gynecol*. 2014;210(5):406-417.
- 389. Magee L, Sawchuck D, Synnes A, von Dadelszen P; Magnesium Sulphate For Fetal Neuroprotection Consensus Committee; Maternal Fetal Medicine Committee. SOGC Clinical Practice Guideline. Magnesium sulphate for fetal neuroprotection. *J Obstet Gynaecol Can*. 2011;33(5):516-529.
- 390. Mallard C, Loeliger M, Copolov D, Rees S. Reduced number of neurons in the hippocampus and the cerebellum in the postnatal

 • CONFIGURATION CONFIDENTIAL 55 14/11 FY $\left|\frac{G_{\text{A}}}{G_{\text{B}}}\right|$ **14/11 FY** $\left|\frac{55}{G_{\text{B}}}\right|$

guinea-pig following intrauterine growth-restriction. *Neuroscience*. 2000;100(2):327-333.

- 391. Sasaki J, Fukami E, Mimura S, Hayakawa M, Kitoh J, Watanabe K. Abnormal cerebral neuronal migration in a rat model of intrauterine growth retardation induced by synthetic thromboxane A(2). *Early Hum Dev*. 2000;58(2):91-99.
- 392. Groom KM, David AL. The role of aspirin, heparin, and other interventions in the prevention and treatment of fetal growth restriction. *Am J Obstet Gynecol*. 2018;218(2S):S829-S840.
- 393. Oyston C, Stanley JL, Oliver MH, Bloomfield FH, Baker PN. Maternal administration of sildenafil citrate alters fetal and placental growth and fetal-placental vascular resistance in the growth-restricted ovine fetus. *Hypertension*. 2016;68(3):760-767.
- 394. Dilworth MR, Andersson I, Renshall LJ, et al. Sildenafil citrate increases fetal weight in a mouse model of fetal growth restriction with a normal vascular phenotype. *PLoS One*. 2013;8(10):e77748.
- 395. Dastjerdi MV, Hosseini S, Bayani L. Sildenafil citrate and uteroplacental perfusion in fetal growth restriction. *J Res Med Sci*. 2012;17(7):632-636.
- 396. Sharp A, Cornforth C, Jackson R, et al. Maternal sildenafil for severe fetal growth restriction (STRIDER): a multicentre, randomised, placebo-controlled, double-blind trial. *Lancet Child Adolesc Health*. 2018;2(2):93-102.
- 397. Pels A, Derks J, Elvan-Taspinar A, et al. Maternal sildenafil vs placebo in pregnant women with severe early-onset fetal growth restriction: a randomized clinical trial. *JAMA Netw Open*. 2020;3(6):e205323.
- 398. Spencer R, Ambler G, Brodszki J, et al. EVERREST prospective study: a 6-year prospective study to define the clinical and biological characteristics of pregnancies affected by severe early onset fetal growth restriction. *BMC Pregnancy Childbirth*. 2017;17(1):43.
- 399. Cluver CA, Hannan NJ, van Papendorp E, et al. Esomeprazole to treat women with preterm preeclampsia: a randomized placebo controlled trial. *Am J Obstet Gynecol*. 2018;219(4):388.e1-388.e17.
- 400. Bauer AJ, Banek CT, Needham K, et al. Pravastatin attenuates hypertension, oxidative stress, and angiogenic imbalance in rat model of placental ischemia-induced hypertension. *Hypertension*. 2013;61(5):1103-1110.
- 401. Costantine MM, Tamayo E, Lu F, et al. Using pravastatin to improve the vascular reactivity in a mouse model of soluble fms-like tyrosine kinase-1-induced preeclampsia. *Obstet Gynecol*. 2010;116(1):114-120.
- 402. Ahmed A, Williams DJ, Cheed V, et al. Pravastatin for early-onset pre-eclampsia: a randomised, blinded, placebo-controlled trial. *BJOG*. 2020;127(4):478-488.
- 403. Griffin IJ, Lee HC, Profit J, Tancedi DJ. The smallest of the small: short-term outcomes of profoundly growth restricted and profoundly low birth weight preterm infants. *J Perinatol*. 2015;35(7):503-510.
- 404. Malin GL, Morris RK, Riley R, Teune MJ, Khan KS. When is birthweight at term abnormally low? A systematic review and meta-analysis of the association and predictive ability of current birthweight standards for neonatal outcomes. *BJOG*. 2014;121(5):515-526.
- 405. Ray JG, Park AL, Fell DB. Mortality in infants affected by preterm birth and severe small-for-gestational age birth weight. *Pediatrics*. 2017;140(6):e20171881.
- 406. Karlberg J, Albertsson-Wikland K. Growth in full-term small-for-gestational-age infants: from birth to final height. *Pediatr Res*. 1995;38(5):733-739.
- 407. Paz I, Seidman DS, Danon YL, Laor A, Stevenson DK, Gale R. Are children born small for gestational age at increased risk of short stature? *Am J Dis Child*. 1993;147(3):337-339.
- 408. De Jesus LC, Pappas A, Shankaran S, et al. Outcomes of small for gestational age infants born at <27 weeks' gestation. *J Pediatr*. 2013;163(1):55-60.e1-3.
- 409. Longo S, Bollani L, Decembrino L, Di Comite A, Angelini M, Stronati M. Short-term and long-term sequelae in intrauterine growth retardation (IUGR). *J Matern Fetal Neonatal Med*. 2013;26(3):222-225.
- 410. Carmody JB, Charlton JR. Short-term gestation, long-term risk: prematurity and chronic kidney disease. *Pediatrics*. 2013;131(6):1168-1179.
- 411. Gluckman PD, Hanson MA, Cooper C, Thornburg KL. Effect of in utero and early-life conditions on adult health and disease. *New Engl J Med*. 2008;359(1):61-73.
- 412. Bhargava SK, Sachdev HS, Fall CH, et al. Relation of serial changes in childhood body-mass index to impaired glucose tolerance in young adulthood. *New Eng J Med*. 2004;350(9):865-875.
- 413. Barker DJ, Osmond C, Forsen TJ, Kajantie E, Eriksson JG. Trajectories of growth among children who have coronary events as adults. *New Eng J Med*. 2005;353(17):1802-1809.
- 414. Eriksson JG. Developmental Origins of Health and Disease - from a small body size at birth to epigenetics. *Ann Med*. 2016;48(6):456-467.
- 415. Heindel JJ, Vandenberg LN. Developmental origins of health and disease: a paradigm for understanding disease cause and prevention. *Curr Opin Pediatr*. 2015;27(2):248-253.
- 416. Jaddoe VWV. Translational challenges for the developmental origins of health and disease: time to fulfill the promises for innovative prevention strategies. *J Dev Orig Health Dis*. 2019;10(3): 260-262.
- 417. Smith GC, Pell JP, Walsh D. Pregnancy complications and maternal risk of ischaemic heart disease: a retrospective cohort study of 129,290 births. *Lancet*. 2001;357(9273):2002-2006.
- 418. van Oostwaard MF, Langenveld J, Schuit E, et al. Recurrence of hypertensive disorders of pregnancy: an individual patient data metaanalysis. *Am J Obstet Gynecol*. 2015;212(5):624.e1-17.
- 419. Hoffman HJ, Bakketeig LS. Heterogeneity of intrauterine growth retardation and recurrence risks. *Semin Perinatol*. 1984;8(1):15-24.
- 420. Patterson RM, Gibbs CE, Wood RC. Birth weight percentile and perinatal outcome: recurrence of intrauterine growth retardation. *Obstet Gynecol*. 1986;68(4):464-468.
- 421. Bratton SL, Shoultz DA, Williams MA. Recurrence risk of low birthweight deliveries among women with a prior very low birthweight delivery. *Am J Perinatol*. 1996;13(3):147-150.
- 422. Bakketeig LS, Hoffman HJ, Jacobsen G, Hagen JA, Storvik BE. Intrauterine growth pattern by the tendency to repeat small-for-gestational-age births in successive pregnancies. *Acta Obstet Gynecol Scand Suppl*. 1997;165:3-7.
- 423. Ananth CV, Kaminsky L, Getahun D, Kirby RS, Vintzileos AM. Recurrence of fetal growth restriction in singleton and twin gestations. *J Matern Fetal Neonatal Med*. 2009;22(8):654-661.
- 424. Redline RW. Placental pathology: a systematic approach with clinical correlations. *Placenta*. 2008;29(Suppl A):S86-S91.
- 425. Redline RW. Classification of placental lesions. *Am J Obstet Gynecol*. 2015;213(4 Suppl):S21-28.
- 426. Redline RW. The clinical implications of placental diagnoses. *Semin Perinatol*. 2015;39(1):2-8.
- 427. Miyakis S, Lockshin MD, Atsumi T, et al. International consensus statement on an update of the classification criteria for definite antiphospholipid syndrome (APS). *J Thromb Haemost*. 2006;4(2):295-306.
- 428. Gleicher N, Pratt D, Dudkiewicz A. The association of antiphospholipid antibodies with pregnancies complicated by fetal growth restriction. *Obstet Gynecol*. 1992;79(4):637-638.
- 429. Lynch A, Marlar R, Murphy J, et al. Antiphospholipid antibodies in predicting adverse pregnancy outcome. *A prospective study. Ann Intern Med*. 1994;120(6):470-475.
- 430. Pattison NS, Chamley LW, McKay EJ, Liggins GC, Butler WS. Antiphospholipid antibodies in pregnancy: prevalence and clinical associations. *Br J Obstet Gynaecol*. 1993;100(10):909-913.

56 | A/II FA/ GXNECOLOGY (We) $\langle \hat{X} \rangle$

- 431. Empson M, Lassere M, Craig JC, Scott JR. Recurrent pregnancy loss with antiphospholipid antibody: a systematic review of therapeutic trials. *Obstet Gynecol*. 2002;99(1):135-144.
- 432. Ziakas PD, Pavlou M, Voulgarelis M. Heparin treatment in antiphospholipid syndrome with recurrent pregnancy loss: a systematic review and meta-analysis. *Obstet Gynecol*. 2010;115(6):1256-1262.
- 433. Mak A, Cheung MW, Cheak AA, Ho RC. Combination of heparin and aspirin is superior to aspirin alone in enhancing live births in patients with recurrent pregnancy loss and positive anti-phospholipid antibodies: a meta-analysis of randomized controlled trials and meta-regression. *Rheumatology (Oxford)*. 2010;49(2):281-288.
- 434. van Hoorn ME, Hague WM, van Pampus MG, Bezemer D, de Vries JI, Investigators F. Low-molecular-weight heparin and aspirin in the prevention of recurrent early-onset pre-eclampsia in women with antiphospholipid antibodies: the FRUIT-RCT. *Eur J Obstet Gynecol Reprod Biol*. 2016;197:168-173.
- 435. Committee on Practice Bulletins—Obstetrics, American College of Obstetricians and Gynecologists. Practice bulletin no. 132: antiphospholipid syndrome. *Obstet Gynecol*. 2012;120(6):1514-1521.
- 436. Tektonidou MG, Andreoli L, Limper M, et al. EULAR recommendations for the management of antiphospholipid syndrome in adults. *Ann Rheum Dis*. 2019;78(10):1296-1304.
- 437. Bates SM, Greer IA, Pabinger I, Sofaer S, Hirsh J. Venous thromboembolism, thrombophilia, antithrombotic therapy, and pregnancy: American College of Chest Physicians Evidence-Based Clinical Practice Guidelines (8th Edition). *Chest*. 2008;133(6 Suppl):844S-886S.
- 438. Howley HE, Walker M, Rodger MA. A systematic review of the association between factor V Leiden or prothrombin gene variant and intrauterine growth restriction. *Am J Obstet Gynecol*. 2005;192(3):694-708.
- 439. Franchi F, Cetin I, Todros T, et al. Intrauterine growth restriction and genetic predisposition to thrombophilia. *Haematologica*. 2004;89(4):444-449.
- 440. Dizon-Townson D, Miller C, Sibai B, et al. The relationship of the factor V Leiden mutation and pregnancy outcomes for mother and fetus. *Obstet Gynecol*. 2005;106(3):517-524.
- 441. Said JM, Higgins JR, Moses EK, et al. Inherited thrombophilia polymorphisms and pregnancy outcomes in nulliparous women. *Obstet Gynecol*. 2010;115(1):5-13.
- 442. Murphy RP, Donoghue C, Nallen RJ, et al. Prospective evaluation of the risk conferred by factor V Leiden and thermolabile methylenetetrahydrofolate reductase polymorphisms in pregnancy. *Arterioscler Thromb Vasc Biol*. 2000;20(1):266-270.
- 443. Silver RM, Zhao Y, Spong CY, et al. Prothrombin gene G20210A mutation and obstetric complications. *Obstet Gynecol*. 2010;115(1):14-20.
- 444. Rodger MA, Hague WM, Kingdom J, et al. Antepartum dalteparin versus no antepartum dalteparin for the prevention of pregnancy complications in pregnant women with thrombophilia (TIPPS): a multinational open-label randomised trial. *Lancet*. 2014;384(9955):1673-1683.
- 445. de Vries JI, van Pampus MG, Hague WM, Bezemer PD, Joosten JH, Investigators F. Low-molecular-weight heparin added to aspirin in the prevention of recurrent early-onset pre-eclampsia in women with inheritable thrombophilia: the FRUIT-RCT. *J Thromb Haemost*. 2012;10(1):64-72.
- 446. Bates SM, Greer IA, Middeldorp S, Veenstra DL, Prabulos AM, Vandvik PO. VTE, thrombophilia, antithrombotic therapy, and pregnancy: antithrombotic therapy and prevention of thrombosis, 9th ed: American College of Chest Physicians Evidence-Based Clinical Practice Guidelines. *Chest*. 2012;141(2 Suppl):e691S-e736S.
- 447. American College of Obstetricians and Gynecologists' Committee on Practice Bulletins-Obstetrics. ACOG practice bulletin no.

197: inherited thrombophilias in pregnancy. *Obstet Gynecol*. 2018;132(1):e18-e34.

- 448. Berghella V. Prevention of recurrent fetal growth restriction. *Obstet Gynecol*. 2007;110(4):904-912.
- 449. Wallenburg HC, Rotmans N. Prevention of recurrent idiopathic fetal growth retardation by low-dose aspirin and dipyridamole. *Am J Obstet Gynecol*. 1987;157(5):1230-1235.
- 450. ACOG Committee Opinion No. 743: low-dose aspirin use during pregnancy. *Obstet Gynecol*. 2018;132(1):e44-e52.
- 451. Horvath A, Koletzko B, Szajewska H. Effect of supplementation of women in high-risk pregnancies with long-chain polyunsaturated fatty acids on pregnancy outcomes and growth measures at birth: a meta-analysis of randomized controlled trials. *Br J Nutr*. 2007;98(2):253-259.
- 452. Gulmezoglu AM, Hofmeyr GJ. Bed rest in hospital for suspected impaired fetal growth. *Cochrane Database Syst Rev*. 2000(2):CD000034.
- 453. Morris RK, Malin G, Robson SC, Kleijnen J, Zamora J, Khan KS. Fetal umbilical artery Doppler to predict compromise of fetal/neonatal wellbeing in a high-risk population: systematic review and bivariate meta-analysis. *Ultrasound Obstet Gynecol*. 2011;37(2):135-142.
- 454. de Laat MW, van der Meij JJ, Visser GH, Franx A, Nikkels PG. Hypercoiling of the umbilical cord and placental maturation defect: associated pathology? *Pediatr Dev Pathol*. 2007;10(4):293-299.
- 455. Redline RW, Ravishankar S. Fetal vascular malperfusion, an update. *APMIS*. 2018;126(7):561-569.
- 456. Saleemuddin A, Tantbirojn P, Sirois K, et al. Obstetric and perinatal complications in placentas with fetal thrombotic vasculopathy. *Pediatr Dev Pathol*. 2010;13(6):459-464.
- 457. Redline RW, Abramowsky CR. Clinical and pathologic aspects of recurrent placental villitis. *Hum Pathol*. 1985;16(7):727-731.
- 458. Redline RW. Villitis of unknown etiology: noninfectious chronic villitis in the placenta. *Hum Pathol*. 2007;38(10):1439-1446.
- 459. Kim MJ, Romero R, Kim CJ, et al. Villitis of unknown etiology is associated with a distinct pattern of chemokine up-regulation in the feto-maternal and placental compartments: implications for conjoint maternal allograft rejection and maternal anti-fetal graftversus-host disease. *J Immunol*. 2009;182(6):3919-3927.
- 460. Kim CJ, Romero R, Chaemsaithong P, Kim JS. Chronic inflammation of the placenta: definition, classification, pathogenesis, and clinical significance. *Am J Obstet Gynecol*. 2015;213(4 Suppl):S53-69.
- 461. Nowak C, Joubert M, Jossic F, et al. Perinatal prognosis of pregnancies complicated by placental chronic villitis or intervillositis of unknown etiology and combined lesions: about a series of 178 cases. *Placenta*. 2016;44:104-108.
- 462. Derricott H, Jones RL, Heazell AE. Investigating the association of villitis of unknown etiology with stillbirth and fetal growth restriction – a systematic review. *Placenta*. 2013;34(10):856-862.
- 463. Chen A, Roberts DJ. Placental pathologic lesions with a significant recurrence risk – what not to miss!. *APMIS*. 2018;126(7): 589-601.
- 464. Boog G. Chronic villitis of unknown etiology. *Eur J Obstet Gynecol Reprod Biol*. 2008;136(1):9-15.
- 465. Boyd TK, Redline RW. Chronic histiocytic intervillositis: a placental lesion associated with recurrent reproductive loss. *Hum Pathol*. 2000;31(11):1389-1396.
- 466. Mekinian A, Costedoat-Chalumeau N, Masseau A, et al. Chronic histiocytic intervillositis: outcome, associated diseases and treatment in a multicenter prospective study. *Autoimmunity*. 2015;48(1):40-45.
- 467. Hannaford P, Mittal N, Sethna F, Dahlstrom JE. Recurrent chronic intervillositis: the diagnostic challenge – a case report and review of the literature. *J Obstet Gynaecol Can*. 2019;41(3):344-347.

- 468. Bos M, Nikkels P, Cohen D, et al. Towards standardized criteria for diagnosing chronic intervillositis of unknown etiology: a systematic review. *Placenta*. 2018;61:80-88.
- 469. Contro E, deSouza R, Bhide A. Chronic intervillositis of the placenta: a systematic review. *Placenta*. 2010;31(12):1106-1110.
- 470. Andres RL, Kuyper W, Resnik R, Piacquadio KM, Benirschke K. The association of maternal floor infarction of the

placenta with adverse perinatal outcome. *Am J Obstet Gynecol*. 1990;163(3):935-938.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.