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Synergetic effect of recoverin and calmodulin on regulation of rhodopsin kinase

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Phosphorylation of photoactivated rhodopsin by rhodopsin kinase (RK or GRK1), a first step of the phototransduction cascade turnoff, is under the control of Ca²⁺/recoverin. Here, we demonstrate that calmodulin, a ubiquitous Ca²⁺-sensor, can inhibit RK, though less effectively than recoverin does. We have utilized the surface plasmon resonance technology to map the calmodulin binding site in the RK molecule. Calmodulin does not interact with the recoverin-binding site within amino acid residues M1-S25 of the enzyme. Instead, the high affinity calmodulin binding site is localized within a stretch of amino acid residues V150-K175 in the N-terminal regulatory region of RK. Moreover, the inhibitory effect of calmodulin and recoverin on RK activity is synergetic, which is in agreement with the existence of separate binding sites for each Ca²⁺-sensing protein. The synergetic inhibition of RK by both Ca²⁺-sensors occurs over a broader range of Ca²⁺-concentration than by recoverin alone, indicating increased Ca²⁺-sensitivity of RK regulation in the presence of both Ca²⁺-sensors. Taken together, our data suggest that RK regulation by calmodulin in photoreceptor cells could complement the well-known inhibitory effect of recoverin on RK.

Keywords: rhodopsin kinase, calmodulin, recoverin, phosphorylation, surface plasmon resonance

INTRODUCTION

Key aspects of the neuronal activity from neurotransmission to gene expression are regulated by changes in the intracellular free calcium concentration. Effects of calcium on various processes in the cell are mediated by Ca²⁺ sensor proteins. The important property of these proteins is the ability to change their conformation upon Ca²⁺-binding, thereby transmitting the signal to the effector enzymes. The effects of calcium in different cells of the most tissues are mediated primarily by ubiquitous calcium sensor calmodulin (Hoeftlich and Ikura, 2002), belonging to the large group of EF-hand calcium-binding proteins. In addition to calmodulin, the other EF-hand calcium-sensitive proteins of the neuronal calcium sensor (NCS) and caldendrin/calneuron families are expressed in neurons (McCue et al., 2010). These proteins regulate different aspects of neuronal function including neurotransmitter release, channel and receptor regulation, control of gene transcription, neuronal growth, and survival. NCS proteins provide a fine-tuning of regulatory mechanisms triggered by changes in calcium level in neurons while calmodulin operates more as a universal sensor of calcium signals. Some of the protein targets are common for both calmodulin and NCS proteins and in a number of cases they were shown to compete for the same binding sites in target molecules (Schaad et al., 1996; Haynes et al., 2006; Few et al., 2011).

The concentration of calmodulin in neurons (including rod outer segments, ROS) is normally 10–25 μM (Liebman, 1972; Kohnken et al., 1981; Kakiuchi et al., 1982) and is comparable to the level of NCS family proteins (maximum intracellular concentration of 30–40 μM was observed in the case of hippocalcin in hippocampal neurons and recoverin in photoreceptors; Klenchin et al., 1995; McCue et al., 2010; Mikhaylova et al., 2011). In this case, one of the possible mechanisms underlying the effective functioning of the NCSs is based on the different affinities of NCSs and calmodulin to calcium. This is for example realized when calmodulin and NCS proteins regulate a specific target protein in different ranges of cytoplasmic calcium (for review, see Burgoyne, 2007; McCue et al., 2010).

In addition to competitive regulation based on the variable Ca²⁺-sensitivity, the effective functioning of NCSs, and calmodulin can be implemented through their interaction with target proteins using separate binding sites. The examples of simultaneous regulation of the effector enzymes by several Ca²⁺-binding proteins are quite common. In photoreceptor cells, retinal guanylyl cyclase 1 contains independent binding sites for four Ca²⁺-binding proteins: guanylyl cyclase activating protein 1 (GCAP1; Lange et al., 1999), GCAP2 (Duda et al., 2005), S100B (Duda et al., 2002), and neurocalcin (Venkataraman et al., 2008).

Four Ca^{2+} binding EF-hand proteins are involved in regulation of sodium/proton exchanger 1 (NHE) via different sites on either juxtamembrane region of NHE1 (calcineurin homologous proteins 1, 2, and tescalcin) or C-terminal regulatory domain (calmodulin; Köster et al., 2011). Another example includes Ca^{2+} -dependent regulation of chimeric calmodulin-dependent protein kinase from plants (Sathyanarayanan et al., 2000). This enzyme contains visinin-like domain, which serves as an intramolecular NCS-like regulator. Such a structure allows the enzyme to complement the external regulation of its activity by calmodulin with intramolecular regulation utilizing the visinin-like domain.

Recoverin is an NCS protein that is primarily expressed in the photoreceptor cells, where it plays a key role in the recovery phase of phototransduction by regulating the activity of rhodopsin kinase (RK; Gorodovikova and Philippov, 1993; Kawamura, 1993; Gorodovikova et al., 1994a,b; Chen et al., 1995; Klenchin et al., 1995; Senin et al., 1995; Makino et al., 2004). Recoverin consists of two domains each containing a pair (functional and non-functional) of EF-hand-type Ca^{2+} -binding sites (Flaherty et al., 1993). The N-terminus of the protein is acylated with myristic acid (Dizhoor et al., 1992). Upon binding of two calcium ions, recoverin undergoes a series of conformational changes allowing the exposure of the amino acid residues of its hydrophobic pocket intended for interaction with RK (Tanaka et al., 1995; Ames et al., 1997, 2006; Komolov et al., 2005). In parallel, the myristoyl group of recoverin becomes exposed imparting the interaction with photoreceptor membranes (so-called Ca^{2+} /myristoyl switch mechanism; Zozulya and Stryer, 1992; Dizhoor et al., 1993). In the photoreceptor cells, recoverin inhibits RK in the dark while this inhibition is relieved upon illumination and the subsequent drop of the intracellular calcium level (Senin et al., 2002b). It allows RK to phosphorylate bleached receptor, rhodopsin, which is the first step to its desensitization (Wilden et al., 1986).

RK (or GRK1) is a member of the G protein-coupled receptor-kinase (GRK) family that specifically phosphorylate agonist-occupied G protein-coupled receptors (GPCRs) and initiate the recruitment of arrestins that induces further receptor desensitization and internalization (Pronin et al., 2002). The common feature of GRKs is the central catalytic domain flanked by two sections of the regulatory RH-domain [from regulator of G protein signaling (RGS) homology] (Singh et al., 2008; Boguth et al., 2010). A C-terminal cysteine residue in RK is modified by a farnesyl group which is involved both in the binding of the enzyme to membranes and in the interaction with its substrate rhodopsin (Inglese et al., 1992; McCarthy and Akhtar, 2000). The recoverin-binding site in RK is localized in the N-terminal portion of the molecule upstream of the RH-domain and consists of the first 25 amino acid residues of the enzyme (Ames et al., 2006; Higgins et al., 2006). Binding of recoverin to this site does not prevent receptor-kinase interaction and thus formation of a transient ternary complex between the kinase, recoverin, and the receptor. Instead, recoverin affects a conformational transition in the RK molecule that is necessary for the phosphorylation of the receptor (Komolov et al., 2009). Previous work has provided some quantitative characteristics of RK regulation by recoverin. The equilibrium dissociation constant

(K_D) of recoverin–RK complex is about $6 \mu\text{M}$ (Satpaev et al., 1998). The half-maximal inhibition of rhodopsin phosphorylation occurs at $3\text{--}6 \mu\text{M}$ recoverin and at $1.5\text{--}3 \mu\text{M}$ $[\text{Ca}^{2+}]_{\text{free}}$ (Klenchin et al., 1995; Senin et al., 1997; Weiergräber et al., 2006; Zernii et al., 2011).

While retina-specific GRK1 (RK) and GRK7 are regulated by Ca^{2+} /recoverin, other GRKs (GRK2-6) are found to be inhibited by the universal mediator of Ca^{2+} signaling calmodulin (Chuang et al., 1996; Pronin et al., 1997). It was shown that calmodulin binding sites in GRK2 and GRK5 are located at both the N- and C-terminal regions of their RH domains and seem to be independent (Levay et al., 1998). Another report suggests that the activity of RK can be regulated not only by recoverin, but also by some additional NCSs as well as by the other Ca^{2+} -binding proteins presented in the photoreceptor cell (De Castro et al., 1995). Among the latter is calmodulin (Kohnken et al., 1981) which in surface plasmon resonance (SPR) studies binds to RK in a Ca^{2+} -dependent manner. The apparent K_D of that complex was $0.1 \mu\text{M}$ and the half-maximal binding was observed at $3 \mu\text{M}$ Ca^{2+} (Levay et al., 1998). Since calmodulin and recoverin are colocalized with RK in photoreceptor cells (Kohnken et al., 1981; Dizhoor et al., 1991; Palczewski et al., 1993; Eckmiller, 2002), the actual process of the Ca^{2+} -dependent regulation of rhodopsin phosphorylation by the enzyme could involve both Ca^{2+} -sensors. In the present study, we demonstrate that a Ca^{2+} -dependent regulation of the RK activity is indeed jointly mediated by recoverin and calmodulin. In particular, we show the synergetic inhibition of the enzyme activity by these Ca^{2+} -sensitive regulators. Additionally, we study the structural basis of this process by mapping the calmodulin binding site in the RK molecule.

MATERIALS AND METHODS

PURIFICATION OF RECOVERIN AND CALMODULIN

Heterologous expression and purification of myristoylated wild-type recoverin has been described in detail in a previous study (Senin et al., 2002a). Calmodulin was purified from bovine brain according to Gopalakrishna and Anderson (1982) with some modifications. Briefly, 100 g of brain tissue was extracted with 200 ml of 50 mM Tris–HCl pH 7.5, 1 mM EDTA, 1 mM 2-mercaptoethanol, 0.5 mM phenylmethylsulfonyl fluoride (PMSF). The extract was subjected to isoelectric precipitation at pH 4.3 followed by heat denaturation of the dissolved pellet at 100°C . Further, renatured calmodulin was loaded onto a phenyl-Sepharose 6B column (GE Healthcare) equilibrated with 50 mM Tris–HCl pH 7.5, 1 mM 2-mercaptoethanol, 0.1 mM CaCl_2 . The column was washed with the same buffer containing 0.5 M NaCl, and calmodulin fractions were eluted with 1 mM EGTA. The resulted fractions were pooled and loaded onto a 5-ml HiTrap DEAE FF anion exchange chromatography column (GE Healthcare) pre-equilibrated with 50 mM Tris–HCl pH 7.5, 1 mM 2-mercaptoethanol at 1 ml/min. After washing the column with the same buffer (~ 10 column volumes), calmodulin was eluted by 30 ml of a linear ionic strength gradient (0–500 mM NaCl) at 0.5 ml/min. Calmodulin containing fractions as revealed by SDS-PAGE were pooled, dialyzed overnight versus 20 mM Tris–HCl pH 8.0, concentrated, and stored at -80°C .

PREPARATION OF ROD OUTER SEGMENTS AND UREA-WASHED ROS MEMBRANES

Bovine ROS were prepared from frozen retinæ according to a well-established procedure (Papermaster and Dreyer, 1974) with some modifications. All procedures were performed under dim red light. 200 frozen retinæ were thawed in 200 ml of buffer A [20 mM Tris-HCl pH 8.0, 0.1 mM EDTA, 1 mM dithiothreitol (DTT)] containing 45% (w/v) sucrose and the resulted mixture was vigorously shaken for 10 min. The suspension was then centrifuged (3,000 g, 4°C) for 20 min and the pellet was discarded. The ROS containing supernatant was diluted twice with buffer A and centrifuged (8,600 g, 4°C) for 30 min. The supernatant was discarded and the pellet containing crude ROS was resuspended in minimal volume of buffer A with addition of 34% (w/v) sucrose, layered over a freshly prepared 34–36% sucrose density step gradient and centrifuged (72,000 g, 4°C) for 2 h. The centrifugation resulted in formation of the compact band containing ROS fraction, which was carefully removed, diluted twice with buffer A and centrifuged for 15 min at 27,200 g, 4°C. The resulting intact ROS pellet was stored at –80°C. Urea-washed ROS membranes were prepared by homogenizing ROS in 5 M urea followed by five-times washing with buffer A as previously described (Senin et al., 2002a).

PREPARATION OF RK CONSTRUCTS AND NATIVE RK

Standard cloning techniques were applied to generate seven RK constructs from a full-length RK cDNA template in a pT7-7 plasmid. DNA fragments corresponding to the RK amino terminal region (N-RK: M1-G183), RK carboxy terminal region (C-RK: I455-S561) as well as to RK amino terminal region stretches M1-S25, M1-L102, L102-G183, D100-V150, and F125-K175 were PCR-amplified and inserted into the pGEX-5x-1 vector to achieve expression as glutathione S-transferase (GST) fusion proteins. Procedures for heterologous expression and affinity chromatography purification of GST-RK constructs were the same as previously described (Komolov et al., 2009). Native RK was purified from ROS as described in detail elsewhere (Senin et al., 2011).

PHOSPHORYLATION ASSAY

RK activity was measured using an *in vitro* rhodopsin phosphorylation assay essentially as described (Weiergräber et al., 2006). Briefly, the assay was performed in 50 μ l reaction mixture containing 10 μ M rhodopsin (urea-washed ROS membranes), 20 mM Tris-HCl pH 7.5, 2 mM MgCl₂, 1 mM γ -³²P-ATP (30–100 dpm/pmol), 1 mM DTT, 1 mM PMSF, and 0.3–0.5 units of rhodopsin kinase with addition of either 200 μ M CaCl₂ or 1 mM EGTA. Recoverin and/or calmodulin at concentrations indicated in the figure legends was added, as appropriate. 1,2-bis(*o*-amino-5-bromophenoxy) ethane-*N,N,N',N'*-tetraacetic acid tetrapotassium salt (5,5'-dibromo-BAPTA) was used as a Ca²⁺-buffer in Ca²⁺-titration experiments according to our previous work (Senin et al., 2002a). The reaction was initiated by addition of ATP and samples were incubated under continuous light for 30 min at 37°C. Incubation was terminated by adding 1 ml of 10% (w/v) trichloroacetic acid. The resulting precipitate was collected by centrifugation and washed three times with 1 ml of 10% trichloroacetic acid; the final pellet was used for Cherenkov counting. The data were analyzed in SigmaPlot 11 (Systat Software)

by plotting relative RK activity [assuming 100% activity at no inhibitor(-s) added] versus Ca²⁺-sensing protein(-s) or free Ca²⁺ concentration and by fitting the plot to four parameter Hill sigmoidal (Eq. 1):

$$y = y_0 + \frac{ax^b}{c^b + x^b} \quad (1)$$

where y is the relative RK activity and x is the recoverin and/or calmodulin or free calcium concentration. The resulting values b (Hill coefficient, n) and c (half-maximal inhibition, IC₅₀) were expressed as best-fit value \pm SE of the fit.

SURFACE PLASMON RESONANCE

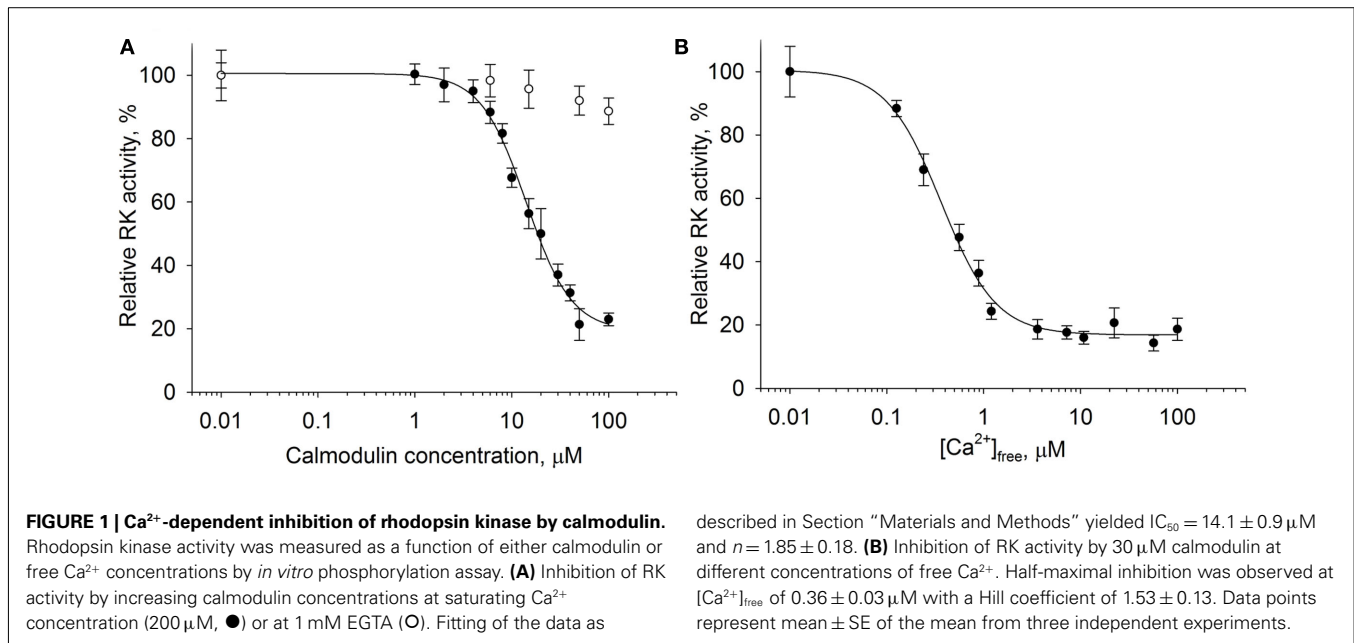
Surface plasmon resonance measurements were performed on a BIACORE 2000 instrument (GE Healthcare) at 25°C. Details of the operation principle, the immobilization procedures and of the evaluation of sensorgrams had been described before (Koch, 2000; Komolov et al., 2006). The analysis of how various GST-RK constructs interact with calmodulin was performed as described recently for recoverin (Komolov et al., 2009) with some modifications. GST-RK fusion proteins were captured on the surface of a CM5 sensor chip (GE Healthcare) with pre-immobilized goat anti-GST antibodies (GE Healthcare) resulting in a surface density of approximately 1 ng/mm². Calmodulin was applied in the mobile phase in running buffer (20 mM Tris-HCl pH 7.5, 100 mM NaCl, 2 mM CaCl₂, 1 mM MgCl₂, 0.005% Tween 20) at 10 μ l/min flow rate. Calmodulin concentration in steady-state affinity analysis was varied from 10 to 150 μ M. Each calmodulin response was double-referenced by subtracting a blank (buffer) and control (injection over GST-coated surface lacking the RK fragments) response (Myszka, 1999). The sensorgrams were also normalized to the amount of immobilized GST-RK. The apparent K_D s for calmodulin-RK peptide complexes were determined by equilibrium analysis as follows: the SPR amplitudes recorded at steady-state of the association phase of each calmodulin injection versus calmodulin concentrations in the mobile phase were plotted using SigmaPlot 11 software and the plots were fitted to the one site saturation ligand-binding model (Eq. 2):

$$y = \frac{B_{\max}x}{K_D + x} \quad (2)$$

where K_D is the apparent equilibrium dissociation constant and B_{\max} is a maximal SPR signal. The resulting K_D values were expressed as best-fit value \pm SE of the fit.

ANALYTICAL PROCEDURES

Rhodopsin content in urea-washed ROS membranes was determined as described previously (Senin et al., 1995). Concentrations of purified recoverin and calmodulin were determined spectrophotometrically assuming ϵ_{280} of 27,500 and 3,030 M⁻¹ cm⁻¹, respectively (Wallace et al., 1983; Johnson et al., 1997). Concentrations of GST-RK constructs were measured using Bradford protein assay (Bio-Rad Laboratories) calibrated with bovine serum albumin. The purity of all the proteins obtained was assessed by SDS-PAGE with Coomassie Brilliant Blue R250 staining.



RESULTS

Ca²⁺-DEPENDENT INHIBITION OF RHODOPSIN KINASE BY CALMODULIN

It is well-known that recoverin inhibits RK *in vitro* in the micromolar range of free Ca²⁺ concentrations ([Ca²⁺]_{free}). Taking into account previous data (Pronin et al., 1997; Levay et al., 1998) it can be supposed that the RK activity can be additionally regulated by calmodulin. We performed a detailed investigation of the ability of calmodulin to affect rhodopsin phosphorylation by RK in a Ca²⁺-dependent manner. For this purpose we applied the *in vitro* phosphorylation assay in the reconstituted system consisting of 0.3–0.5 units of RK purified from ROS, 10 μM dark-adapted rhodopsin in the content of urea-washed ROS membranes, and various concentrations of calmodulin ranging from 0.01 to 100 μM (Figure 1A). The experiment was conducted at high (200 μM [Ca²⁺]_{free}) or low (1 mM EGTA) Ca²⁺ levels.

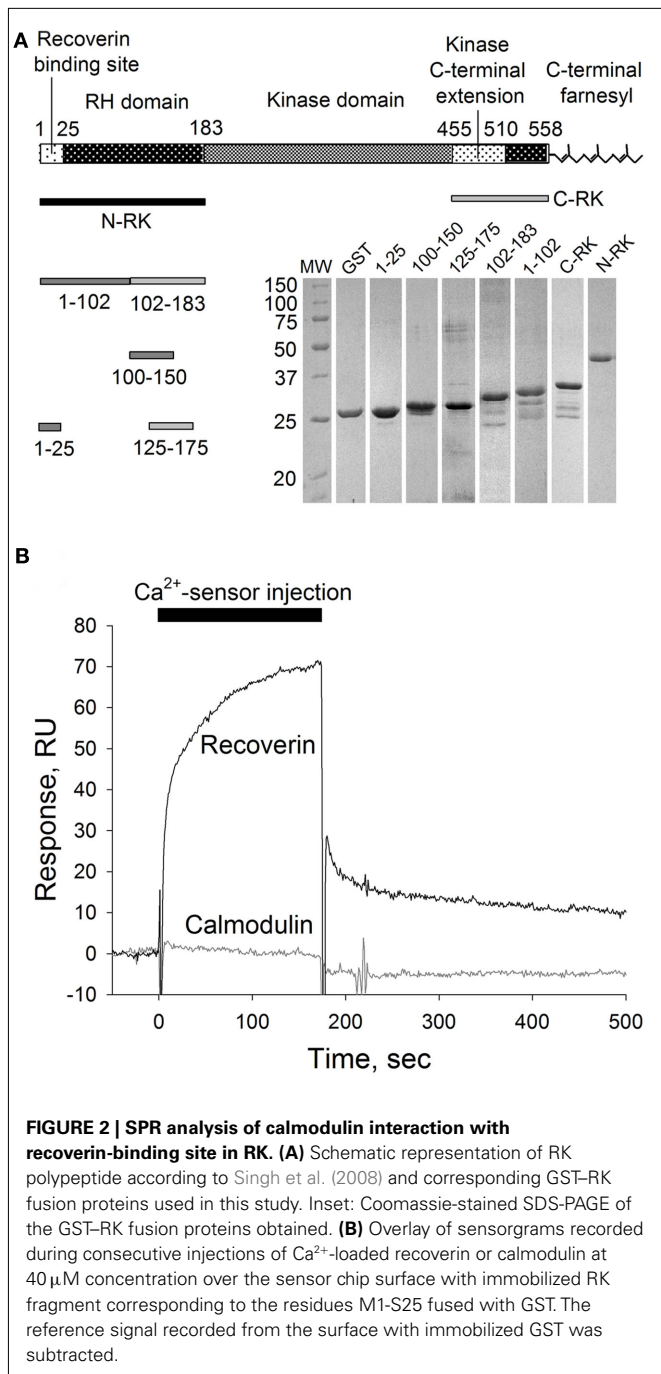
According to our data, calmodulin inhibited RK in the presence of Ca²⁺; the inhibition was half-maximal at 14 μM protein concentration. In the absence of Ca²⁺, calmodulin was ineffective. We further examined rhodopsin phosphorylation by RK as a function of free Ca²⁺ concentration in the presence of saturating concentration (30 μM) of calmodulin (Figure 1B). Inhibition reached half maximum at 0.36 μM [Ca²⁺]_{free} with a Hill coefficient of 1.53. In summary, these findings indicate the ability of calmodulin to operate *in vitro* as a Ca²⁺-dependent inhibitor of RK.

MAPPING OF THE CALMODULIN BINDING SITE IN RHODOPSIN KINASE

The observed ability of calmodulin to inhibit RK in a Ca²⁺-dependent manner poses a question whether it binds to the same site in the RK molecule as recoverin does or whether these two Ca²⁺-sensors interact with separate specific sites in the enzyme. We employed a biosensor-based technique, the SPR spectroscopy, to study the interaction of recoverin and calmodulin with GST-fused N-terminal RK peptide M1-S25 containing the recoverin-binding site (Higgins et al., 2006; here and below, see Figure 2A for a schematic representation of RK fusions constructed and

tested). The RK peptide was captured on the anti-GST antibody-coated sensor chip and the binding of Ca²⁺-loaded forms of calmodulin and recoverin (both at concentration of 40 μM) to the immobilized RK fragment was monitored in real-time resulting in sensorgrams shown at Figure 2B.

As expected, recoverin interacted with the RK fragment containing the first 25 amino acids of the enzyme (Figure 2B). The apparent *K_D* of recoverin for M1-S25 fragment was about 17 μM, as determined in titration series (data not shown). The interaction of calmodulin with the same RK peptide was negligible (Figure 2B). These results suggest the existence of separate binding sites for calmodulin and recoverin. To identify calmodulin binding site(s), we constructed two recombinant GST fusion fragments that encompass amino terminal (N-RK: M1-G183) and carboxy terminal (C-RK: I455-S561) regulatory regions of RK. Only the N-RK fragment readily captured supplied calmodulin (Figure 3A), while the calmodulin binding to C-RK fragment was almost absent. To further localize the calmodulin binding site within N-RK, we tested the ability of calmodulin to interact with N-RK fragments of smaller size: M1-L102 and L102-G183. The interaction occurred in both cases but the response was much more pronounced for M102-G183 (Figure 3B). Moreover, we showed that within the RK sequence L102-G183 the calmodulin binding site is presumably localized within a short stretch of 25 amino acids present in positions 150–175 since the N-RK fragment F125-K175 bound calmodulin, but the fragment D100-V150 did not (Figure 3C). The additional low affinity site for calmodulin is localized within the RK sequence S26-L102 since the interaction of RK fragment M1-S25 with calmodulin was negligible (Figures 2B and 3B). To evaluate the quantitative characteristics of the effects observed, we carried out steady-state affinity analysis of calmodulin interaction to those GST-RK constructs, which displayed a significant binding to the Ca²⁺-sensor. The analysis was performed by recording series of sensorgrams at different calmodulin concentrations (in the range of 10–150 μM) and measuring SPR signal amplitudes at steady-state (Figure 3D). The apparent



equilibrium dissociation constants were determined as concentrations required for half-maximal saturation of the binding signal. The obtained K_D values were 63 μM for the full-length N-RK, 54 μM for the M1-L102 fragment, 23 μM for the L102-G183 fragment, and 17 μM for the F125-K175 fragment.

Thus, both high and low affinity calmodulin binding sites are localized within the N-terminal regulatory part of RK, and they are separated from the recoverin-binding site consisting of M1-S25 sequence of the enzyme. Together, our findings indicate that both Ca²⁺-sensors, recoverin, and calmodulin, are able to bind RK using separate specific sites, suggesting the possibility of their simultaneous binding to and inhibition of RK.

SYNERGETIC INHIBITION OF RHODOPSIN KINASE BY CALMODULIN AND RECOVERIN

We have found that calmodulin displays an inhibitory effect on RK at free Ca²⁺ concentrations ranging from 0.01 to 1 μM (Figure 1B). The range overlaps with the physiological changes of Ca²⁺ concentrations found in the photoreceptor cell (Gray-Keller and Detwiler, 1994). Moreover, the revealed inhibitory action of calmodulin was observed at Ca²⁺ concentrations even below the Ca²⁺-sensitivity of recoverin, the well-established RK regulator. Since both proteins colocalize in photoreceptor cells with RK, we asked whether these two Ca²⁺-sensors could have a joint effect on RK activity. Thus we utilized the *in vitro* phosphorylation assay examining the ability of calmodulin and recoverin to simultaneously regulate the activity of RK. Urea-washed ROS membranes containing 10 μM rhodopsin were incubated with 0.3–0.5 units of RK in the presence of increasing concentrations of either calmodulin, recoverin, or their 1:1 mixture at the saturating free Ca²⁺ concentration (200 μM).

Remarkably, calmodulin and recoverin were able to synergistically inhibit the RK activity (Figure 4A). While the IC₅₀ for calmodulin or recoverin alone were 14.1 and 5.8 μM, respectively, their joint action resulted in half-maximal RK inhibition at 4.6 μM of the total Ca²⁺-sensing protein concentration in the mixture. Approximation of the experimental data was performed with the Hill equation and revealed that the interaction between RK and recoverin was cooperative with Hill coefficient of 1.6. A good curve fit was also possible when approximating the same data with one site (Langmuir) ligand-binding model (data not shown). Collectively, these results point out that the inhibition of RK by recoverin occurs at equimolar stoichiometric ratio of the proteins. In contrast, the inhibition of RK by calmodulin alone as well as the synergetic inhibition by both Ca²⁺ sensors had Hill indexes of about 2. The latter fact evidences for the cooperative binding of two molecules of Ca²⁺-sensor per one RK molecule. The inhibitory effect of calmodulin on RK was found to occur at lower free Ca²⁺ concentrations than the effect of recoverin. Calmodulin inhibited RK at a half-maximal free Ca²⁺ concentration of 0.36 μM while the inhibitory effect of recoverin was at 1.36 μM Ca²⁺ (Figure 4B). The synergetic inhibition of RK by the 30-μM calmodulin/recoverin mixture was observed at a half-maximal free Ca²⁺ concentration of 1.07 μM. Apparently, the synergetic action of both Ca²⁺-sensors caused a shift in the affinity of the target–effector interaction and at the same time an extension of the operating Ca²⁺-range. In Ca²⁺-titration experiments where the RK inhibition by calmodulin or recoverin was studied, the Hill coefficients were within the range of 1.53–1.66, indicating similar cooperative action of the cation in the case of both Ca²⁺-sensors. Thus, our data indicate that calmodulin complements the inhibitory action of recoverin on RK and increases the sensitivity of RK inhibition to Ca²⁺.

MODELING OF EQUILIBRIUM BETWEEN RK, RECOVERIN, CALMODULIN, AND CALCIUM IONS

We further used the parameters obtained to create a model of the synergetic inhibition of RK by calmodulin and recoverin. Since the combined use of both Ca²⁺-sensors increases efficiency of RK inhibition (Figure 4A), the fully competitive scheme of RK

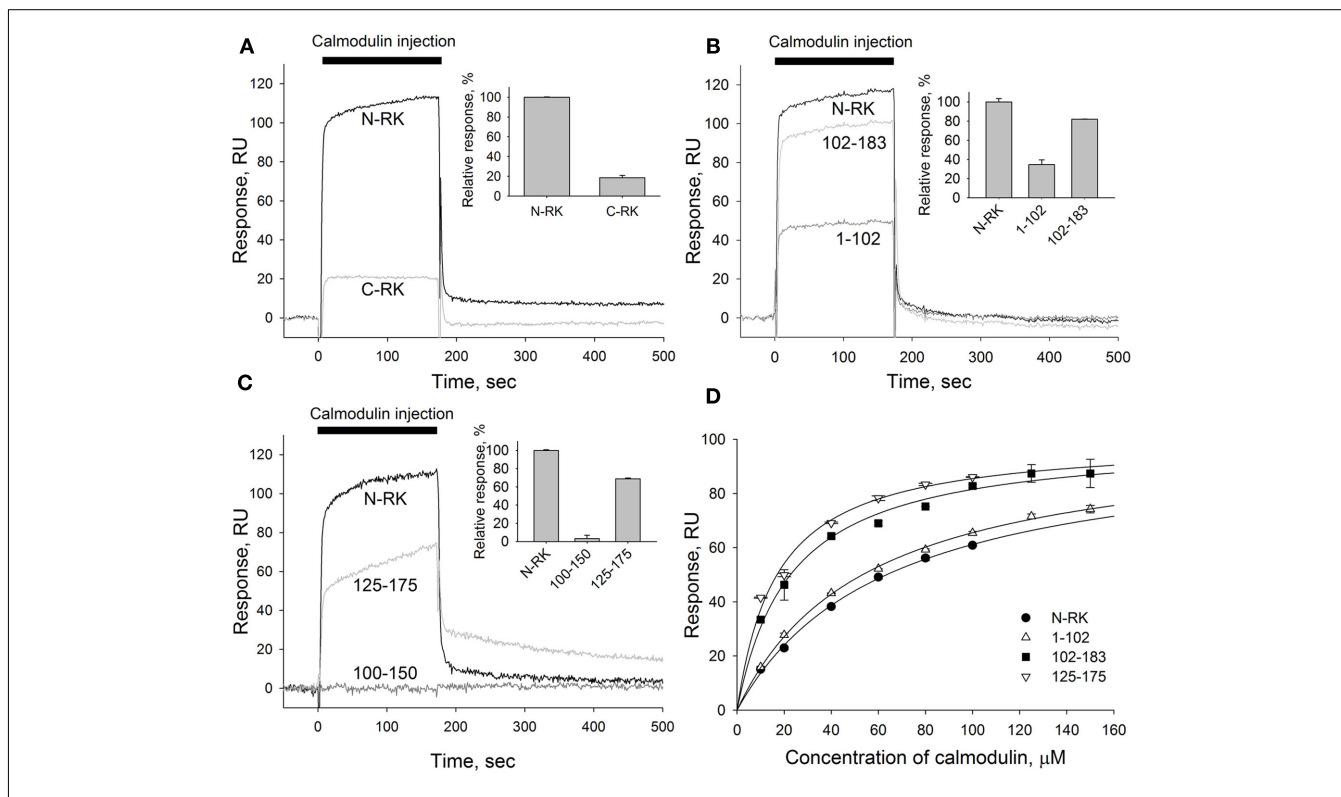
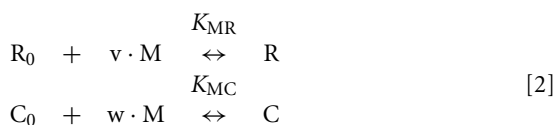
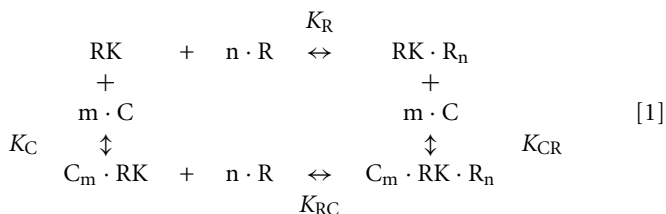


FIGURE 3 | Mapping of the calmodulin binding site in the N-terminal region of RK. (A–C) A representative overlay of SPR sensorgrams showing real-time binding of calmodulin to RK fragments anchored on the sensorchip surface via GST tags using anti-GST antibodies covalently coupled to the dextran matrix. Sixty micromolar calmodulin in running buffer containing 2 mM Ca^{2+} was injected over the surfaces with immobilized N-terminal or C-terminal regions of RK (termed N-RK and C-RK, respectively) (A), N-RK and its fragments corresponding to the residues M1-L102 and L102-G183 (B), or N-RK and its fragments corresponding to the residues D100-V150 and F125-K175 (C). The sensorgrams were normalized to the amounts of immobilized fragments, and the reference signal from a control surface with immobilized GST was subtracted. *Insets* show relative responses at the end

of the injection observed for each GST-RK fragment indicating the amount of calmodulin bound. (D) Steady-state affinity analysis of calmodulin binding to the full-length N-RK or its fragments M1-L102, L102-G183, or F125-K175. Binding of calmodulin was recorded at concentrations ranging from 10 to 150 μM. The amplitudes of binding signals at equilibrium were determined, normalized, and are shown as a function of calmodulin concentration. Apparent K_D s were determined as half-maximal calmodulin concentrations required for saturation of SPR amplitudes as described under Section “Materials and Methods.” The obtained values for N-RK, M1-L102, L102-G183, and F125-K175 are 63 ± 3, 54 ± 2, 23 ± 2, and 170 ± 1.5 μM, respectively. Data points represent mean ± SE of the mean from two independent experiments. The calculated K_D s are expressed as best-fit value ± SE of the fit.

regulation was excluded. The SPR experiments strongly evidence for the presence of separate recoverin- and calmodulin binding sites. The simplified scheme of chemical equilibria describing this case looks as follows (the rationale behind the use of this scheme is explained in the Modeling of Equilibrium Between RK, Recoverin, Calmodulin, and Calcium Ions in Appendix):



Here, M denotes calcium ion, R and C denote Ca^{2+} -bound states of recoverin and calmodulin, respectively, while R_0 and C_0 denote their Ca^{2+} -free states. Each of the equilibria is characterized by its own association constant (see Modeling of Equilibrium Between RK, Recoverin, Calmodulin, and Calcium Ions in Appendix; Table 1). Fitting of the experimental data shown in Figure 4 to the Hill equation gives estimates of the most of the constants and binding stoichiometries of Schemes [1] and [2]. Notably, these values implicitly take into account the presence of membranes, which are known to increase the affinity of recoverin to Ca^{2+} (Zozulya and Stryer, 1992) and to affect the distribution of RK between soluble and membrane-bound forms (Sanada et al., 1996). Therefore, these values should be considered as effective. The analysis of the experimental data shown in Figure 4A (see Modeling of Equilibrium Between RK, Recoverin, Calmodulin, and Calcium Ions in Appendix) enabled us to estimate all parameters of the Scheme [1] (Table 1). According to our analysis, the binding of Ca^{2+} -loaded recoverin molecules to RK facilitates its further association with Ca^{2+} -bound calmodulin, as manifested by 1.55-fold increase in

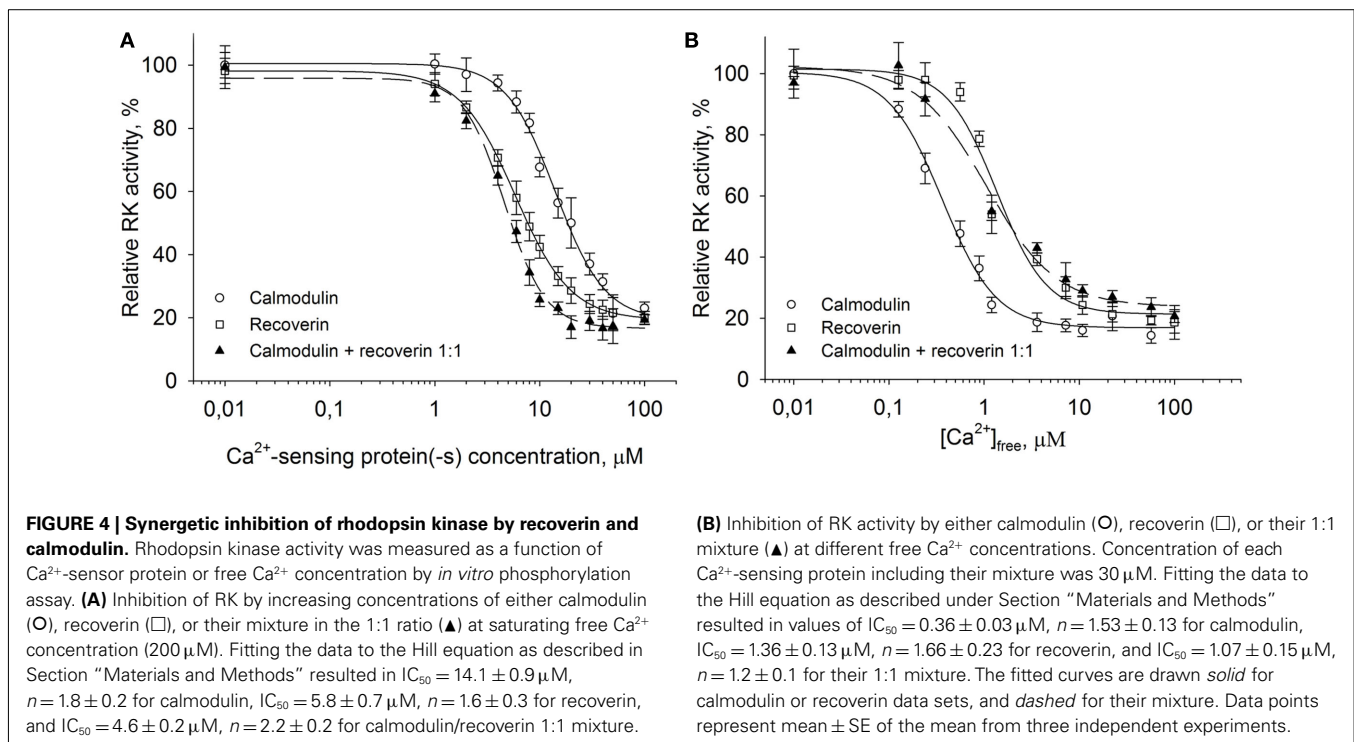


Table 1 | The parameters of the equilibrium between RK, recoverin, calmodulin, and calcium ions.

Equilibrium association constant	Value, M^{-1}	Hill coefficient	Value
K_R	1.7e5	n	1.6
K_C	7.1e4	m	1.8
K_{RC}	2.7e5	–	–
K_{CR}	1.1e5	–	–
K_{MR}	7.4e5	v	1.66
K_{MC}	2.8e6	w	1.53

The parameters of Schemes [1] and [2] are derived from fitting of the experimental data shown in **Figure 4** to the Hill equation, except for the K_{RC} and K_{CR} values, which were estimated from analysis of the data shown in **Figure 4A** using Eqs A11 and A12 in Appendix (see Modeling of Equilibrium Between RK, Recoverin, Calmodulin, and Calcium Ions in Appendix), respectively.

the constant of calmodulin binding to RK. Similarly, association of calmodulin with RK increases its recoverin-binding constant 1.59-fold (**Table 1**). Thus, the modeling results suggest that recoverin and calmodulin could act in a concerted manner, mutually facilitating inhibition of RK.

The changes in RK-inhibiting activity shown in **Figure 4B** reflect association of the Ca^{2+} -sensing proteins with calcium ions. These processes can be approximated by the Hill equation based model (Scheme [2]), which is widely used for simplified description of such complex processes. Alternatively, the Ca^{2+} -dependence of interaction between calmodulin and its target could be described by the model of sequential filling of EF-hands (Dagher et al., 2011). However, this approach requires

the introduction into the Scheme [2] of additional chemical equilibria, which would drastically complicate modeling of the system. According to our analysis based on Scheme [2] (see Modeling of Equilibrium Between RK, Recoverin, Calmodulin, and Calcium Ions in Appendix), the half-maximal synergetic inhibition of RK by calmodulin and recoverin occurs at free Ca^{2+} concentration of 0.71 μM . This value is between the respective values obtained for calmodulin (0.36 μM) and recoverin (1.36 μM). The corresponding experimental value equals to 1.07 μM (**Figure 4B**) which is close to the theoretical estimate.

DISCUSSION

In the present work we have demonstrated that the ubiquitous Ca^{2+} -sensor calmodulin is able to function as an inhibitor of RK. Moreover, we found that the inhibition of RK by calmodulin can complement the inhibitory action of the well-known RK regulator recoverin. Previous attempts to study the ability of calmodulin to regulate GRKs, including GRK1 (RK), were performed using relatively low concentrations of calmodulin (not exceeding 10 μM ; Chuang et al., 1996; Pronin et al., 1997) at which its inhibitory effect could not be seen to the full extent. According to our data, the inhibition of RK by calmodulin at saturating Ca^{2+} concentrations was observed in the range of calmodulin concentration from 2 to 30 μM and was half-maximal at 14 μM of the Ca^{2+} -sensor, whereas recoverin was a more effective inhibitor, as it showed a half-maximal inhibitory effect on the RK activity at 5.8 μM . The calcium range of RK inhibition by calmodulin (0.05–1 μM of $[\text{Ca}^{2+}]_{\text{free}}$) overlapped with the physiological range of the cation concentration (0.05–0.55 μM of $[\text{Ca}^{2+}]_{\text{free}}$) in ROS (Gray-Keller and Detwiler, 1994). The synergetic action of calmodulin and recoverin on the RK activity was within 0.05–1 μM of $[\text{Ca}^{2+}]_{\text{free}}$.

These values are slightly above the physiological range of Ca^{2+} concentration in ROS. However, the recoverin affinity for Ca^{2+} is significantly increased in the presence of RK and ROS membranes (Zozulya and Stryer, 1992; Ames et al., 1995; Senin et al., 1995, 2004). Similarly, the affinity of calmodulin for Ca^{2+} increases upon binding to target enzymes (Shifman et al., 2006). Therefore, extrapolation to the *in vivo* conditions (including high membrane content and presence of target proteins) indicates that the apparent Ca^{2+} -dependence of the synergetic action of calmodulin and recoverin on RK activity could reach the physiological (i.e., submicromolar) range of the calcium concentration. Our experimental results were supplemented with theoretical validation of the ability of calmodulin and recoverin to synergistically bind RK. Different approaches could be applied to modeling of equilibria between RK, recoverin, calmodulin, and calcium ions. Full description of the system would require characterization of each step of the calcium-binding to the calcium-sensing proteins (Dagher et al., 2011), as well as characterization of all protein–protein interactions in the system. Considering that membranes are known to affect the distribution of RK and recoverin between soluble and membrane-bound fractions, as well as to modulate calcium affinity of recoverin (Zozulya and Stryer, 1992; Senin et al., 1995; Sanada et al., 1996), it can be concluded that the total number of chemical equilibria is significantly higher (see Modeling of Equilibrium Between RK, Recoverin, Calmodulin, and Calcium Ions in Appendix). This objective difficulty can be compensated via usage of the Hill model-based representation of the system, which describes the multiple processes of ligand-binding as a single event. Despite being phenomenological, this approach enabled us to estimate the synergy of binding of calmodulin and recoverin to RK. According to our analysis (see Modeling of Equilibrium Between RK, Recoverin, Calmodulin, and Calcium Ions), binding of either calcium sensor to RK results in 1.5-fold increase of the affinity of the complex to another Ca^{2+} -sensing protein. The analysis also showed that the half-maximal synergetic inhibition of RK by calmodulin and recoverin occurs at free Ca^{2+} concentration of $0.71 \mu\text{M}$, which is close to the corresponding experimental value of $1.07 \mu\text{M}$.

The structural aspects underlying the synergetic regulation of RK by calmodulin and recoverin can be identified from the experiments on mapping of the calmodulin binding site in RK using a biosensor-based approach (SPR spectroscopy). Our findings indicate that the interaction of calmodulin and recoverin with RK occurs by means of separate binding sites in the N-terminal region of the enzyme, which makes the co-inhibitory action of the two Ca^{2+} -sensors possible. The recoverin-binding site is known to be located in the first 25 amino acid residues of RK (Ames et al., 2006; Higgins et al., 2006). According to our data, RK contains two calmodulin binding sites. The first, low affinity site is located within the sequence S26-L102 of N-terminal regulatory region of RK, which is in agreement with a previous report (Levay et al., 1998). The second high affinity site was identified within the residues V150-K175 of the same RK region. In other studies this fragment was not tested for the ability to bind calmodulin. The C-terminal regulatory region of RK (I455-S561) did not interact with calmodulin both in the current and in the earlier studies. In addition, the report by Levay et al. (1998) suggested another calmodulin binding site within the residues A341-E400. This site

is located in the large lobe of the catalytic domain of the enzyme (Singh et al., 2008). However, calmodulin apparently has no effect on the catalytic activity of RK since in our experiments it did not affect the ability of the enzyme to phosphorylate the C-terminal peptide of rhodopsin containing phosphorylation sites (data not shown). Thus, the suggested interaction of calmodulin with A341-E400 site seems to be out of the physiological relevance. Moreover, it was shown that the recombinant fragment corresponding to the full-length catalytic domain of RK (E184-D454) is not able to interact with rhodopsin (Komolov et al., 2009), although the domain function should include this interaction. The latter suggests that this fragment does not fully acquire its native structure. These observations led us not to consider parts of the catalytic domain in the context of Ca^{2+} -dependent regulation of RK by calmodulin.

It should be mentioned that the affinities of calmodulin and recoverin to the RK peptides determined in our SPR experiments were lower than in the previous work (Levay et al., 1998). The possible reason of this discrepancy can be the different strategies applied for the immobilization of the interaction partners on the sensor chip. Levay et al. (1998) immobilized calmodulin and recoverin on the chip via cysteine residues. Calmodulin was biotinylated via the cysteine residue and further captured on the streptavidin-coated surface while recoverin was directly attached to the sensor chip surface via thiol coupling with its cysteine residue. These cysteine residues were shown to play a critical role in the Ca^{2+} -dependent operating of both proteins (Lukas et al., 1984; Permyakov et al., 2007, 2012). Thus, the functional state of calmodulin and recoverin on the sensor chip could be significantly affected upon the immobilization procedure. We used an experimental configuration opposite to their approach with RK fragments being retained on the sensor chip surface and recoverin/calmodulin being supplied in the mobile phase. RK fragments in this case were captured on the surface under mild conditions using antigen–antibody interaction. This strategy might better preserve the protein integrity in BIACORE experiments and therefore affinity constants determined by SPR for calmodulin interaction with its binding site ($17 \mu\text{M}$) correlate better with the IC_{50} for calmodulin-mediated RK inhibition ($14 \mu\text{M}$).

Based on the location of the recoverin and calmodulin binding sites in RK one may suggest the putative mechanisms underlying the intrinsic inhibition activity in the case of each Ca^{2+} -sensor. As it was previously mentioned, recoverin inhibits RK by affecting the conformational transitions of the enzyme required for the formation of the efficient enzyme-substrate complex between RK and rhodopsin C-terminus. Such a mechanism is due to the ability of recoverin to bind to the N-terminal site M1-S25 of RK that is critical for the function of the enzyme (Higgins et al., 2006; Huang et al., 2009; Komolov et al., 2009; Zernii et al., 2011). It is likely that the mechanism of RK inhibition by calmodulin involves a Ca^{2+} -dependent interference with the interaction between RK and photoexcited rhodopsin. This is possible, if the calmodulin binding site in RK overlaps with the receptor docking site. The sequence F150-K175 in RK identified in our work as the high affinity calmodulin binding site contains amino acid residues Y167, Q173, W174, L177 which could be involved in rhodopsin recognition since the corresponding residues in the GRKs 5 and 6 were recently

found to be critical for the phosphorylation of the activated receptors (Baameur et al., 2010). Thus, calmodulin may be suggested to inhibit RK by blocking sterically the association of the enzyme with the responsive sites in its receptor substrate rhodopsin. Preliminary observations made by us further support this conclusion, namely that the fragment F150-K175 of RK is indeed capable of direct binding to rhodopsin in a light-dependent manner. Such a mechanism does not contradict the current hypothesis of the formation of the heteromeric complex between RK, rhodopsin, and Ca^{2+} -sensor(s) (Komolov et al., 2009), since it is known that the receptor binding involves not only the N-terminal regulatory region of RK containing the above mentioned fragment but also the C-terminal farnesyl modification of the enzyme (McCarthy and Akhtar, 2000).

Considering the above speculations the question arises what could be the mechanism underlying the synergetic inhibition of RK by both Ca^{2+} -sensors. The SPR experiments strongly evidence that the recoverin- and calmodulin binding sites in RK are separate. Since combined action of recoverin and calmodulin increases efficiency of RK inhibition, the competitive scheme of the enzyme regulation by two Ca^{2+} -sensors is unlikely. The modeling of equilibrium between RK, recoverin, calmodulin, and calcium ions revealed that the binding of Ca^{2+} /recoverin to RK results in 1.5-fold facilitation of binding of the enzyme to Ca^{2+} /calmodulin, and *vice versa*. Taking into account the higher affinity of recoverin to RK and its higher intracellular concentration as compared to calmodulin it can be suggested that the binding of Ca^{2+} /recoverin to the enzyme precedes that of Ca^{2+} /calmodulin. Presumably, the binding of recoverin to the first 25 amino acid residues of RK allosterically increases the availability of the high affinity calmodulin binding site V150-K175 for the interaction with calmodulin. This is consistent with the fact that the presence of recoverin shifts the binding of calmodulin to RK (and consequent inhibition of the enzyme) to the lower calmodulin concentration. The suggested sequential filling of Ca^{2+} -sensor binding sites in RK, namely recoverin followed by calmodulin, is in agreement with the fact that the Ca^{2+} -dependence of the synergetic inhibition effect is close to the recoverin Ca^{2+} -dependent inhibition

profile. Thus, the following consecutive mechanism of RK regulation by recoverin and calmodulin in photoreceptors can be suggested. At high calcium concentration (0.5–0.6 μM of $[\text{Ca}^{2+}]_{\text{free}}$) present in dark-adapted photoreceptors, RK is strongly inhibited by the synergetic action of both Ca^{2+} -sensors. Shortly after the light absorption, when the Ca^{2+} -concentration has not yet decreased, recoverin still allosterically inhibits phosphorylation of rhodopsin by RK however not affecting the interaction between the enzyme and the cytoplasmic loops of the receptor. In the same time calmodulin prevents this interaction thus allowing the receptor to effectively activate transducin. This is important for the ability of rod photoreceptors to respond to even low illumination levels. During the lowering of the intracellular Ca^{2+} concentration associated with phototransduction, recoverin dissociates from the complex with RK, which decreases the affinity of the enzyme to calmodulin. In these conditions calmodulin alone can still inhibit the enzyme though less effectively than it does in combination with recoverin. Further light-induced decrease in free Ca^{2+} concentration down to 0.025–0.05 μM triggers the dissociation of calmodulin from RK switching the enzyme to the full active form, which causes desensitization of the receptor.

In summary, our study suggested a novel mechanism of regulation of rhodopsin desensitization by rhodopsin kinase involving Ca^{2+} -sensor proteins. Further studies are required for firm establishment of the physiological role of this phenomenon and unraveling the molecular details of the proposed mechanism.

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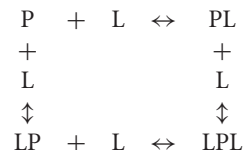
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APPENDIX

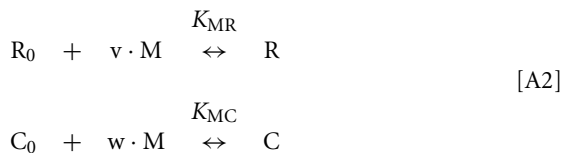
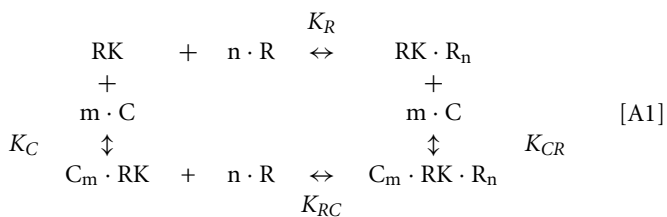
MODELING OF EQUILIBRIUM BETWEEN RK, RECOVERIN, CALMODULIN, AND CALCIUM IONS

In the simplest case the process of binding of two ligand (L) molecules by a protein (P) is described with the “four-states” scheme:



This scheme may be approximated by the scheme of sequential filling of the ligand-binding sites of the protein upon certain ratio of the equilibrium binding constants, describing the scheme. In the case of equilibrium between RK, recoverin, calmodulin, and calcium ions the following equilibria should be taken into account: (1) two four-states schemes, describing the calmodulin–calcium and RK–calmodulin interactions (overall, eight equilibria); (2) one sequential binding scheme, describing the recoverin–calcium interaction (two equilibria); (3) one equilibrium between RK and recoverin; (4) additional equilibria related to the presence of mixed RK forms with simultaneously bound calmodulin and recoverin molecule(s). Overall, the number of chemical equilibria describing this system exceeds 11. Furthermore, the experiments were performed in the presence of membranes, which affect the distribution of recoverin and RK between soluble and membrane-bound forms (Zozulya and Stryer, 1992; Sanada et al., 1996). Hence, the number of equilibria describing this system is actually much higher. The complexity of this system prevents from usage of microscopic constants for its description. The alternative approach is the use of effective equilibrium constants, describing separate stages of the process. Although this approach is phenomenological, it greatly facilitates description of the system and enables to establish the synergy between calmodulin- and recoverin-binding sites of RK.

The following simplified scheme of chemical equilibria describing the interactions between RK, recoverin, calmodulin, and calcium ions can be suggested:



Here, M denotes calcium ion, R and C denote Ca²⁺-bound states of recoverin and calmodulin, respectively, while R₀ and C₀ denote

their Ca²⁺-free states. Each of the equilibria is characterized by its own effective association constant:

$$K_R^n = [RK \cdot R_n] / ([RK] \cdot [R]^n) \quad [A1]$$

$$K_C^m = [C_m \cdot RK] / ([RK] \cdot [C]^m) \quad [A2]$$

$$K_{RC}^n = [C_m \cdot RK \cdot R_n] / ([C_m \cdot RK] \cdot [R]^n) \quad [A3]$$

$$K_{CR}^m = [C_m \cdot RK \cdot R_n] / ([RK \cdot R_n] \cdot [C]^m) \quad [A4]$$

$$K_{MR}^v = [R] / ([R_0] \cdot [M]^v) \quad [A5]$$

$$K_{MC}^w = [C] / ([C_0] \cdot [M]^w) \quad [A6]$$

Combining the Eqs A1–A4, the following relationship between the parameters of the Scheme [A1] can be derived:

$$K_{RC}^n / K_{CR}^m = K_R^n / K_C^m \quad [A7]$$

The changes in RK-inhibiting activity (see **Figure 4A**) reflect association of the Ca²⁺-sensing proteins with RK under saturating Ca²⁺ level, so at the middle of the transition the following condition takes place:

$$[RK \cdot R_n] + [C_m \cdot RK] + [C_m \cdot RK \cdot R_n] = [RK] \quad [A8]$$

Using Eqs A1–A3, the latter equation can be rewritten:

$$K_R^n \cdot [R]^n + K_C^m \cdot [C]^m \cdot (1 + K_{RC}^n \cdot [R]^n) = 1 \quad [A9]$$

Assuming that the synergetic inhibition of RK by recoverin/calmodulin mixture occurs at [C] = [R] = 4.6 μM (see **Figure 4A**; other numerical values are taken from **Table 1**),

$$K_R^n + K_C^m \cdot [R]^{m-n} \cdot (1 + K_{RC}^n \cdot [R]^n) = [R]^{-n} \quad [A10]$$

$$K_{RC}^n = [R]^{-n} \cdot [(R)^{-n} - K_R^n] \cdot [R]^{n-m} / K_C^m - 1 \quad [A11]$$

$$K_{RC} = 2.7e5 M^{-1}$$

Hence, from Eq. 7,

$$K_{CR} = K_C \cdot (K_{RC} / K_R)^{n/m} = 1.1e5 M^{-1} \quad [A12]$$

The Ca²⁺-dependence of RK-inhibiting activity of recoverin and calmodulin (see **Figure 4B**) reflects association of the Ca²⁺-sensing proteins with calcium ions, so at the middle of the transition the following equation takes place:

$$\begin{aligned}
 [R] + [C] + n \cdot [RK \cdot R_n] + m \cdot [C_m \cdot RK] + (m + n) \\
 \cdot [C_m \cdot RK \cdot R_n] = [R_0] + [C_0]
 \end{aligned} \quad [A13]$$

Since total concentration of RK was about 20 nM, the concentrations of all RK states may be neglected in comparison with concentrations of the RK-free forms of recoverin and calmodulin:

$$[R] + [C] = [R_0] + [C_0] \quad [A14]$$

Considering the material balance in the system,

$$[R] + [R_0] = R_0 \quad [A15]$$

$$[C] + [C_0] = C_0 \quad [A16]$$

where R₀ and C₀ are total concentrations of recoverin and calmodulin, respectively.

The system of Eqs A14–A16 and A5–A6 can be solved with respect to $[M]$.

Substituting Eqs A5–A6 into Eqs A14–A16,

$$K_{MR}^v \cdot [R_0] \cdot [M]^v + K_{MC}^w \cdot [C_0] \cdot [M]^w = [R_0] + [C_0] \quad (A17)$$

$$K_{MR}^v \cdot [R_0] \cdot [M]^v + [R_0] = [R_0] \quad (A18)$$

$$K_{MC}^w \cdot [C_0] \cdot [M]^w + [C_0] = [C_0] \quad (A19)$$

From Eqs A17–A19:

$$[R_0] \cdot (K_{MR}^v \cdot [M]^v - 1) + [C_0] \cdot (K_{MC}^w \cdot [M]^w - 1) = 0 \quad (A20)$$

$$[R_0] = R_0 / (1 + K_{MR}^v \cdot [M]^v) \quad (A21)$$

$$[C_0] = C_0 / (1 + K_{MC}^w \cdot [M]^w) \quad (A22)$$

Substituting Eqs A21–A22 into Eq. 20, and considering that $R_0 = C_0$:

$$\begin{aligned} & (K_{MR}^v \cdot [M]^v - 1) / (1 + K_{MR}^v \cdot [M]^v) \\ & + (K_{MC}^w \cdot [M]^w - 1) / (1 + K_{MC}^w \cdot [M]^w) = 0 \end{aligned} \quad (A23)$$

$$\begin{aligned} & K_{MR}^v \cdot [M]^v - 1 + K_{MR}^v \cdot K_{MC}^w \cdot [M]^{v+w} - K_{MC}^w \cdot [M]^w \\ & + K_{MC}^w \cdot [M]^w - 1 + K_{MR}^v \cdot K_{MC}^w \cdot [M]^{v+w} \\ & - K_{MR}^v \cdot [M]^v = 0 \end{aligned} \quad (A24)$$

$$K_{MR}^v \cdot K_{MC}^w \cdot [M]^{v+w} = 1 \quad (A25)$$

$$[M] = (K_{MR}^{-v} \cdot K_{MC}^{-w})^{1/(v+w)} = 0.71 \mu\text{M} \quad (A26)$$