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Regulation of a duplicated locus: Drosophila sloppy paired is replete with functionally overlapping enhancers.

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Regulation of a Duplicated Locus: Drosophila *sloppy paired* is Replete with Functionally Overlapping Enhancers

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Abstract

In order to investigate regulation and redundancy within the *sloppy paired* (*slp*) locus, we analyzed 30 kilobases of DNA encompassing the tandem, coordinately regulated *slp1* and *slp2* transcription units. We found a remarkable array of stripe enhancers with overlapping activities surrounding the *slp1* transcription unit, and, unexpectedly, glial cell enhancers surrounding *slp2*. The *slp* stripe regulatory region generates 7 stripes at blastoderm, and later 14 stripes that persist throughout embryogenesis. Phylogenetic analysis among drosophilids suggests that the multiplicity of stripe enhancers did not evolve through recent duplication. Most of the direct integration among cis-regulatory modules appears to be simply additive, with one notable exception. Despite the apparent redundancy among stripe enhancers, transgenic rescue suggests that most are required for full function, to maintain *wingless* expression and parasegment boundaries throughout embryogenesis. Transgenic rescue also reveals indirect positive autoregulation by the 7 early stripes, without which alternate stripes within the 14-stripe pattern are lost, leading to embryos with a pair-rule phenotype.

Introduction

The *sloppy paired* (*slp*) locus contains two tandem transcription units, *slp1* and *slp2*, that both encode transcription factors with a forkhead domain (Grossniklaus et al., 1992). During segmentation of the germ band, they act downstream of primary pair-rule genes such as *even skipped* (*eve*) (Fujioka et al., 1995), and have been classified as secondary pair-rule genes (Akam, 1987; Cadigan et al., 1994b; Ingham, 1988).

Like several other pair-rule genes, including *eve*, *slp1* and *slp2* are expressed in both 7- and 14-stripe patterns (Grossniklaus et al., 1992; Macdonald et al., 1986). The 7-stripe pattern is established, in part, through repression by the 7-stripe pattern of *eve*: in *eve* mutants, each *slp* stripe expands posteriorly into the *eve* domain (Fujioka et al., 1995). In turn, *slp* helps to restrict the late *eve* stripe pattern and to maintain the *engrailed* (*en*) pattern of 14 stripes (Jaynes and Fujioka, 2004). More specifically, *slp* expression in 14 stripes helps to maintain the parasegment (PS) boundary by preventing the expansion of *en* stripes anteriorly into the *slp* domain, and by maintaining *wingless* (*wg*) expression in the *slp*-expressing cells (Cadigan et al., 1994a, b; Jaynes and Fujioka, 2004). In turn, En represses *slp* in an interaction that is likely to be direct (Kobayashi et al., 2003). Specific sites of En action in the *slp* locus have not yet been localized. This function in establishing and maintaining PS boundaries is conserved in insects that do not have a clear pair-rule stage of segmentation (Choe and Brown, 2007).

Early transgenic studies suggested that regulatory DNA upstream of *slp1* is required for segmentation function (Grossniklaus et al., 1992). Despite the fact that both *slp1* and *slp2* are expressed in the same striped pattern (with *slp2* appearing to start a bit later), a rescue construct containing this region along with only the *slp1* transcription unit can rescue most of the segmentation defects caused by a deficiency of the entire *slp* locus (Cadigan et al., 1994a), suggesting that *slp2* may be dispensable for segmentation. In further transgenic studies, several cis-regulatory modules (CRMs) were identified. The 6 kb just upstream of *slp1* was shown to contain at least 3 CRMs, producing a head stripe at blastoderm, germband stripes at stages 10-11, and stripes in the ventral ectoderm at stage 11 and later (Lee and Frasch, 2000). A genome-wide search for Bicoid binding site clusters helped to identify 3 *slp* CRMs, of which two (located about 1 kb 5' and 3 kb 3' of *slp1*) were shown to drive head stripes at blastoderm (Ochoa-Espinosa et al., 2005). Using consensus binding site information for segmentation gene products, another head stripe CRM was identified about 2 kb 5' of *slp2* (Schroeder et al., 2004).

Most recently, an interaction between two CRMs further upstream of *slp1* was studied (Prazak et al., 2010). One region drives 14 stripes beginning at blastoderm, and shows ectopic activation in

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some cells within odd-numbered parasegments that normally do not express detectable levels of *slp* RNA. However, when combined with another CRM, which drives properly restricted expression within even-numbered parasegments, ectopic expression is repressed, suggesting that an interaction between distant CRMs plays an important role in *slp* regulation.

Motivated by a desire to more fully understand the regulation and function of the *slp* locus, we conducted a systematic transgenic analysis of a 30 kilobase (kb) region surrounding the *slp* transcription units. This analysis revealed a surprising degree of overlap in both space and time in the striped expression driven by CRMs surrounding *slp1*, as well as unexpected neuronal regulatory CRMs surrounding *slp2*. Phylogenetic analysis suggests that the multiplicity of stripe CRMs did not evolve through recent duplication. Extensive dissection of the regulatory region showed that integration of this CRM information is mostly additive, with the exception noted above. We rescued *slp* mutants with transgenes carrying various CRMs. These experiments confirm the conclusions of Prazak et al. (2010), and show that the improper pattern driven by the upstream region produces significant embryonic defects. These experiments also reveal that autoregulation, through repression of a repressor, is a primary function of the early 7-stripe pattern. They further suggest that the extensive apparent redundancy among stripe elements actually provides for fully functional levels of expression across the many stages of *slp* expression.

Materials and Methods

Plasmids construction and production of transgenic flies

To analyze CRM activities, conventional P-element transgenesis was used (Fujioka et al., 1998; Spradling and Rubin, 1982). To generate DNA fragments, PCR was performed using BAC clone 06H02 as template (obtained from the Berkeley Drosophila Genome Project (Hoskins et al., 2000)). PCR fragments were cloned into a modified P-element vector (Fujioka et al., 1999) upstream of a *lacZ* reporter gene. For *slp1-promoter-lacZ*, the region from −261 (SfiI) to +121 bp relative to the *slp1* transcription start site (TSS), or for *slp2-promoter-lacZ*, the region from −314 to +373 bp (relative to the *slp2* TSS), was fused to the *lacZ* coding region followed by the *eve* 3' UTR from +1306 to +1521 bp (KpnI). The *mini-white* gene is positioned so that the two genes are divergently transcribed. Several independent insertion sites were analyzed for each construct, and the expression patterns shown were seen consistently.

To analyze the rescue ability of u8100, a region from −8.1 to +1.5 kb relative to the *slp1* TSS, which includes 78 bp 3' of the *slp1* mRNA polyA signal, was cloned into a conventional P-element

vector. Five independent insertion sites were analyzed, and showed similar rescue ability. To compare the rescue ability of different CRMs, ΦC31 recombinase-mediated cassette exchange (ΦC31-RMCE) was used (Bateman et al., 2006). Various CRM regions were cloned into attBΔ2 (Fujioka et al., 2008). The regions used for the rescue constructs are described in the figure legends. ΦC31-RMCE was performed as previously described (Bateman et al., 2006), except that chromosomally integrated ΦC31 recombinase (Bischof et al., 2007) was used, instead of co-injection of ΦC31 mRNA. Successful RMCE events were first identified by loss of *mini-white*-dependent eye color. The presence and direction of the exchanged region were confirmed by PCR. The attP-docking site at cytological location 95E5 (Fujioka et al., 2008) was used.

Embryo analysis

Embryos were subjected to *in situ* hybridization using anti-sense RNA probes against *lacZ, slp1* (which may cross-react with *slp2*), and *wg* mRNA, or to antibody staining with anti-β-galactosidase (β-gal, ICN) as previously described (Fujioka et al., 1999). For glial cell expression, anti-β-gal, and anti-Reversed polarity (Repo) (Alfonso and Jones, 2002) obtained from the Developmental Studies Hybridoma Bank, were visualized with DyLight549-conjugated anti-mouse IgG and DyLight488-conjugated anti-rabbit IgG (Jackson Immuno Research). Rescue constructs were analyzed in a *CyO,Δ34* mutant (Grossniklaus et al., 1992) background. Cuticle preparation was performed as previously described (Fujioka et al., 1995).

Sequence comparison and analysis

To identify conserved sequence blocks (CSBs) within each *slp* CRM, we used the phylogenetic analysis application EvoPrinter (Odenwald et al., 2005) on the cis-Decoder web site (Brody et al., 2007, 2008), with default settings. These CSBs were then used in cis-Decoder, with default settings, to identify conserved sequence clusters (CSCs), and to ask whether *slp* CRMs with overlapping expression patterns share CSCs.

We performed BLAST searches from FlyBase (Tweedie et al., 2009) with individual CRM sequences against other drosophilid genome sequences (Clark et al., 2007) using default settings. The most conserved subsequences were then BLAST searched against both the *D. melanogaster* and *A. gambiae* genomes, using an expect value of 1000. Matching sequences were placed on a map of the region to determine their relative positions and orientations. This methodology provided evidence for specific homologous sequences for most of the *slp* CRMs in a common ancestor of the drosophilids, but not between the drosophilids and *A. gambiae*.

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To identify possible transcription factor binding to a 12 bp element shared between two mesodermally expressed CRMs (see Results), we searched *Drosophila* transcription factor binding site matrices in the JASPAR database (Bryne et al., 2008; Portales-Casamar et al., 2010) using each 6 bp subsequence with a relative profile score threshold of either 90% (described as "high stringency" in "Results") or 80% ("low stringency").

Results

Regulatory anatomy of the *sloppy paired* locus

Motivated by a desire to better understand the regulation of the tandem *slp1* and *slp2* transcription units by pair-rule and segment polarity genes, we performed a detailed mapping of enhancer activity throughout the locus. Although several *slp* CRMs have been localized and studied (Lee and Frasch, 2000; Ochoa-Espinosa et al., 2005; Prazak et al., 2010; Schroeder et al., 2004), a locus-wide analysis had not been done. We surveyed the 30.9 kb genomic region from 10.3 kb upstream of *slp1* to 9.4 kb downstream of *slp2* for enhancer activity using reporter transgenes. We initially tested approximately 2 kb DNA fragments overlapping by about 500 bp, producing 18 transgenic constructs (see Fig. 1A for map, Fig. S1 for expression data).

In our CRM names, the initial letter indicates the location relative to transcription units: (u) upstream of *slp1*, (i) internal, between *slp1* and *slp2*, and (d) downstream of *slp2*. The numbers following these letters indicate the end points in hundreds of bp, where the first 2 digits (or 3 for u10382) are the 5' end point, and the remaining digits are the 3' end point. For example, "u8172" extends from about −8.1 to −7.2 kb relative to the *slp1* transcription start site (TSS), "i1523" from +1.5 to +2.3 kb relative to the *slp1* TSS, and d2445 from +2.4 to +4.5 kb relative to the *slp2* TSS. Regions upstream of *slp1* were tested for CRM activity in the context of *slp1-promoter-lacZ*, while regions between *slp1* and *slp2*, and those downstream of *slp2*, were analyzed in the context of *slp2-promoter-lacZ* (see Materials and Methods).

Apparent redundancy among stripe elements

Of our 18 constructs carrying about 2-kb each of regulatory DNA (Fig. 1), 8 showed a 14-stripe pattern (Fig. S1), suggesting a surprising level of redundancy in producing this aspect of *slp* expression. The region represented by these 8 constructs span the *slp1* TSS. Further dissection of these 8 constructs identified 8 non-overlapping CRMs that each give a 14-stripe pattern (Figs. 2, 3; Fig. 1B,C for maps and summary). Two of these (u1609, and i2330) are restricted to the mesoderm.

Unlike early *eve* stripes, which are produced individually or in pairs by distinct CRMs (Fujioka et al., 1999; Goto et al., 1989; Harding et al., 1989; Sackerson, 1995), multiple, non-overlapping *slp* CRMs drive expression of 7 stripes (both u4734 and u3225 drive first 7 then 14 stripes, while u3125 drives only 7 stripes) or 14 stripes (u8172, u5547, u2316, and i1523) in the ectoderm. This is consistent with its role as a secondary pair-rule gene, as it is regulated by other 7- and 14-stripe patterns of primary and secondary pair-rule genes, as well as by segment polarity genes. Although we did identify the 7-stripe-specific u3125 within the 7-plus-14 stripe u3225, we did not identify a 7-stripe-specific subregion of u4734. In recent studies, CRMs u3125 and u8172 were analyzed in greater detail (Prazak et al., 2010), and a binding site for an activator of *slp*, Odd-paired, was found in u8172 (Sen et al., 2010).

The non-overlapping CRMs u1609 (Figs. 1B and 2) and i2330 (Figs. 1C and 3) each drive a 14-stripe pattern in the mesoderm at stage 10. A similar pattern is seen with u8766 (Fig. S1), showing that there is apparent redundancy in mesodermal, as well as ectodermal, stripe expression. Shortening u8766 to create u8772 causes loss of this mesodermal stripe expression (data not shown; see Fig. S2 for detailed locations of *slp* CRMs and a summary of expression data). We note that our CRM u1609 is a more compact version of the previously described slp5'-1 (see Fig. S2), while u3931 corresponds roughly to slp5'-3 (Lee and Frasch, 2000).

After the stripe activities of u8172, u3725, u1609, i1523 (Figs. 2, 3), and u5547 (not shown, its activity is somewhat weaker than the others) fade, expression in some cells in the CNS, probably neuroblasts, becomes apparent. These CRMs are good candidates for providing the known function of *slp* in developing neuroblasts (Bhat et al., 2000). The CRMs u0900 and i2330 (Figs. 2, 3) also drive expression like that previously described for *slp* as ventrolateral cell clusters (Grossniklaus et al., 1992). We also saw this activity in 3 out of 7 independent transgenic lines with u3931 (data not shown). We were not able to separate these activities from the stripe activities, suggesting that they are regulated by overlapping sets of transcription factors.

There are 4 non-overlapping CRMs (u3931, u1609, u0900, and i3039) that drive a head stripe at the blastoderm stage (Figs. 2, 3). In addition, several other CRMs drive weak head expression (Fig. S2, data not shown). Both a region spanning the junction between u1609 and u0900 and a region within i3039 were identified by searching for Bicoid binding site clusters genome-wide (Ochoa-Espinosa et al., 2005). These clusters are presumably involved in activating the head stripes. The early head-stripe generating u3931 was not identified at high stringency by this method (Ochoa-Espinosa et al., 2005; Schroeder et al., 2004). However, at lower stringency, clustered Bicoid binding sites can be found there (Hongtao Chen and Stephen Small, personal communication). As u3525

also drives this head expression (data not shown, see Fig. S2), the region common to these constructs

(−3455 to −3056 bp) is a good candidate for functional Bicoid binding sites. Bicoid binding in the vicinity of these CRMs has been confirmed in a genome-wide study using chromatin immunoprecipitation (Li et al., 2008), where one binding region encompasses u3931, another spans u1609 and u0900, and a third spans i3039.

Some stripe CRMs described above also drive expression in 3rd instar larvae. CRM u8766 drives reporter gene expression in dorsal and ventral sections of the eye disc (Fig. S3 A-E), and also affects *mini-white* expression within the transgene, causing patterned eye color (Fig. S3 F-J). In other lines with the same CRM, reporter gene expression behind the morphogenetic furrow was stronger, and was not associated with patterned eye color (Fig. S3 K-N). Perhaps strong, uniform late expression of *mini-white* masks the effect on eye color of earlier patterned *mini-white* expression.

Intriguingly, u8781 drives a ring of expression in the brain of 3rd instar larvae (Fig. 3). Although the *slp* locus is not known to have a function in this part of the nervous system, *slp* RNA is also seen there in a pattern similar to that of u8781 (Fig. 3). CRMs i1530, i2330, and i2339 each drive a stripe of expression closer to the ventral midline in the larval CNS and brain (Fig. S3 O-Q). However, we were unable to clearly detect endogenous *slp* expression there. Nonetheless, such a similar activity of multiple CRMs suggests functional significance.

Stripe element rescue of the *slp* mutant phenotype

None of the *slp* stripe CRMs drive an expression pattern that continues until stage 13 (data not shown), when endogenous *slp* RNA can still be seen (Grossniklaus et al., 1992). A 9.6 kb construct spanning from −8.1 to +1.5 kb, including the *slp1* transcription unit, was tested for the ability to rescue the *slpΔ34* mutant chromosome, which is a modified *CyO* balancer chromosome with a deletion that removes the *slp1* coding region, the intergenic region, and the 5' half of the *slp2* coding region, and thus is null for both transcripts (Grossniklaus et al., 1992). Consistent with a loss of expression at stage 13 driven by the individual *lacZ* reporter constructs, this rescue construct did not maintain *slp* expression to stage 13 (not shown). Nonetheless, it did rescue the *slpΔ34* segmentation defects almost completely (Fig. 4) at several different P-element insertion sites, although mild abnormalities within the ventral denticle bands could still be seen in many embryos. Because *CyO/CyO* embryos do not hatch (but don't have segmentation defects), it is unclear whether this rescue construct would rescue hatching of a *slp* null mutation in an otherwise wildtype background (which does not exist).

We tested several combinations of apparently redundant stripe CRMs for their ability to rescue the *slp* null mutant phenotype. Comparisons were made at the same chromosomal docking site, using the ΦC31 recombinase system (Bateman et al., 2006; Groth et al., 2004). We first tested the same region used in Fig. 4, spanning from −8.1 to +1.5 kb (u8100, Fig. 5). The rescue ability at this attP-docking site (at cytological location 95E5) was indistinguishable from that seen at several random chromosomal insertion sites using P-element transgenesis (Fig. 4). Therefore, this docking site was used for all subsequent rescue analysis. The rescue ability of this construct was very similar to that seen previously for a longer construct that included the two downstream stripe CRMs i1523 and i2330 (Cadigan et al., 1994a). This suggests that these two CRMs are functionally redundant with the upstream stripe CRMs.

To further test for redundancies within the stripe elements of the *slp* locus, we first tested the upstream-most 3 kb of the 9.6 kb rescue construct u8100, in combination with an extended promoter and *slp1* coding region from −904 bp to +1536 bp (u8150, Fig. 5). This includes CRMs that give 14 stripes beginning at stage 7 (u8172, Fig. 2, which is contained within u8766, Fig. S1) and persisting until stage 12 (u7250, Fig. S1), as well as strong head expression (u900, Fig. 2). Consistent with the mild ectopic expression seen with u8172 (Fig. 2 and (Prazak et al., 2010)), this construct drove clear ectopic expression within the odd-numbered parasegments (confirmed by co-staining for Eve, data not shown). This results in an aberrant *wg* expression pattern at embryonic stage 7 (Fig. 5, 2nd column) that is largely, but not completed, corrected at later stages. Most rescued embryos end up with a pair-rule deletion of naked cuticle between ventral denticle bands at the end of embryogenesis (Fig. 5, bottom rows, 2nd column). Such a pair-rule phenotype was not seen with the u8100 rescue construct (described above). These results are consistent with those described previously (Prazak et al., 2010), and suggest that the activity of the upstream-most stripe CRMs is restricted through Eve-dependent repression, acting through regulatory regions closer to the TSS. Nonetheless, this construct rescues *wg* expression quite well, and the pair-rule defects that remain are relatively mild (compare to *slp⁻* in Fig. 4).

We also tested two other combinations of stripe CRMs for their rescue ability. One extends from −5.5 kb through the *slp1* coding region (to +1536 bp). It also includes both early and later stripe CRMs, as well as a CRM that drives strong head expression. It drives approximately normal *slp1* expression similar to that of the u8100 rescue construct, and rescues the *wg* and cuticle patterns quite well (u5500, Fig. 5). However, many more embryos show mild cuticle defects than with the u8100 rescue construct, indicating that the level of *slp* expression is not sufficient for full rescue. The fact that both of these rescue constructs, which share two CRMs, rescue the mutant phenotype well

shows that there is some redundancy between them, as expected from the reporter analysis. On the other hand, the fact that neither one rescues as completely as the combination of the two (u8100) shows that this redundancy is only partial, when examined at the level of functional rescue.

Finally, we discovered an autoregulatory requirement for the 7 early *slp* stripes in activating 7 of the 14 late stripes. This was revealed when we tested a combination of the stripe CRMs downstream of *slp1* along with an extended *slp1* promoter and coding region (–665 through +1539 bp) for the ability to rescue the *slpΔ34* mutant phenotype. As with the above rescue transgenes, the combination of these elements drives both 14-stripe *lacZ* expression and head expression (Figs. 3 and S1). However, the stripe expression does not begin until slightly later than with the other rescue constructs. In contrast to the other rescue transgenes, this one (i1539, Fig. 5) gives strong expression in only 7 stripes in the *slp* mutant background, and only weak expression in the other 7. This reveals a functional requirement for the early 7-stripe pattern, which is very weak in these embryos. Without these 7 early stripes of *slp* expression, half of the 14 later stripes do not form properly. These are the ones located just anterior to the 7 early stripes of *eve* expression, and in a *slp* mutant, *odd-skipped* (*odd*) stripes have been shown to expand into these cells, preventing activation of half of the *wg* stripes (Jaynes and Fujioka, 2004). This expanded *odd* expression can also prevent activation of 7 of the *slp* stripes within the 14-stripe pattern, accounting for our results. This loss of every other *slp* stripe, and the accompanying loss of *wg*, results in a pair-rule loss of naked cuticle between ventral denticle bands in the odd-numbered parasegments (Fig. 5, 4th column). Clearly, these downstream CRMs are not sufficient for rescue, apparently because they do not drive the 7 early stripes strongly enough. All in all, these results suggest that despite the seeming redundancy when stripe CRMs are tested individually, all of them contribute to full *slp* function in the native context.

Glial cell regulatory elements

Strikingly, several CRMs that do not drive striped expression do drive patterns in the nervous system. CRMs i4053, i5882, d2445, and d5778 (as well as the partially overlapping d6383) drive expression in spindle-shaped cells in both the central and peripheral nervous systems (Fig. 3). Based on the cell shape, we suspected that these were glial cells. To test this, we double stained for expression of our reporter and a glial cell marker, Repo, product of the gene *reversed polarity*, or *repo* (Campbell et al., 1994; Halter et al., 1995; Xiong et al., 1994). As shown in Fig. 6, i4053 drives expression in most, but not all, Repo-expressing cells at embryonic stage 13. Few, if any, strongly reporter-expressing cells are Repo-negative, suggesting that reporter expression is limited to glial cells. Since *slp* CRMs i5882, d2445, and d5776 (a shorter version of d5778) also drive expression in

glial cells (Fig. 6), including some that do not express *i4053-lacZ* (data not shown), it is likely that most, if not all, glial cells express one or more *slp-lacZ* reporter. Furthermore, transgenes carrying i4060, which contains i4053, gave reporter gene expression in eye disc cells in the position of glia (Fig. S3R, S) (Campbell et al., 1994; Xiong et al., 1994). Although we have not been able to see a consistent signal for *slp* RNA or Slp protein in these cells, a transgenic line that carried a BAC clone in which the *slp2* coding region was fused to GFP showed expression (Venken et al., 2009) similar to our constructs (data not shown), suggesting that endogenous *slp2* is expressed there, but either transiently or at a relatively low level. The lack of good Slp antibodies, however, leaves this an open question. The independent activities of several CRMs in glial cells suggest that they have been the subject of considerable evolutionary selection, consistent with the regulation of glial cell fates by *slp*. However, this appears to be a separate function from the previously described negative regulation of glial cell fates by *slp* (Mondal et al., 2007).

Evolutionary origins of the *slp* stripe CRMs

We explored the evolution of the *slp* CRMs using two methodologies. First, we used BLAST searches to identify related regions among the sequenced drosophilid genomes (Clark et al., 2007). Each of the CRMs tested showed strong conservation among the more closely related drosophilids, all of which contain both a *slp1-* and a *slp2*-related coding region in a similar tandem arrangement to that in *D. melanogaster*. Sufficient similarity was found within each tested CRM to identify a related region in most of the drosophilid species. The locations of these sequence similarities are shown on maps of the *slp* locus in Figs. S4A (for the more closely related species) and S4B (for those more distantly related to *D. melanogaster*). Overall, this analysis suggests that separate elements related to each of these CRMs existed in the common ancestor of the drosophilids, about 40 million years ago (Russo et al., 1995).

We also performed BLAST searches with the most conserved elements of each stripe CRM against both the *D. melanogaster* and *Anopheles gambiae* (mosquito) genomes, as mosquitoes are the next most closely related lineage for which sequenced genomes are available. Within *D. melanogaster*, the best match found among co-expressed CRMs was between the two mesodermal stripe CRMs u1609 and i2330. This 12 bp sequence, GACGTCTTCATT, is highly conserved among drosophilids within u1609, but not within i2330. We used this sequence to search the JASPAR transcription factor database (Bryne et al., 2008; Portales-Casamar et al., 2010). The only high stringency predicted binding site was for the homeodomain-containing *ventral veins lacking* gene product, which has not been found to be expressed in a pattern (Tomancak et al., 2002; Tomancak et al., 2007) that overlaps with those driven by these CRMs. At lower stringency, possible binding by

very many transcription factors is predicted. Likewise, many relatively low stringency matches could be found between conserved elements of our *D. melanogaster* CRMs and sequences surrounding the *A. gambiae* transcription unit that is most closely related to *D. melanogaster slp*. However, we did not find a pattern to these matches that suggested the existence of common ancestral regulatory regions. We also did not find another *slp*-related coding sequence on the same chromosome as that of the most closely related sequence. This closest sequence is more similar to *slp2* than to *slp1* (data not shown). This suggests that the twin *slp* coding regions in drosophilids arose from a duplication event occurring after the split from their last common ancestor with mosquitoes.

Lastly, because shared conserved sequence clusters have previously been found in some CRMs with similar expression patterns (Brody 2007), and a genome-wide database of conserved sequence clusters (CSCs) has been generated, we used the interactive resources Evoprinter and cis-Decoder to identified CSCs using each of our 15 minimal *slp* CRMs (listed in Fig. S2 in boldface) as a starting point. Although CSCs were found within all except 3 of them (u3931, i4053, and i5882), none of these CSCs were shared among co-expressed CRMs. The single case of a shared CSC was between the glial cell CRM d5778 and a CSC that spans the junction between u1609 and u2316. However, these three CRMs have little or no overlap in their expression patterns (Figs. 2, 3), so the functional significance of these results is unclear.

Discussion

Individual stripe enhancers

We did not identify CRMs that drive individual *slp* stripes in the germ band at any stage, consistent with the *slp* locus acting strictly downstream of the primary pair-rule genes, which are responsible for converting non-periodic patterns of the maternal and gap gene products into periodic 7- or 8-stripe patterns (Ingham, 1988). However, a head stripe is driven by several separable CRMs (u3931, u1609, u0900, and i3039; Figs. 1-3; see also Fig. S2). The early *slp* head stripe is regulated by the maternal gradient-generating gene *bicoid*, among other genes, and some of these CRMs contain previously identified clusters of Bicoid binding sites (Li et al., 2008; Ochoa-Espinosa et al., 2005; Schroeder et al., 2004).

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Two distinct regulatory domains

The *slp1*, but not the *slp2*, transcription unit is surrounded by stripe CRMs. This situation may have arisen following a chromosomal duplication that gave rise to these twin transcription units. However, a simple duplication within an array of unique CRMs cannot explain the current regulatory landscape of this locus. In stark contrast to *slp1*, *slp2* is surrounded by nervous system CRMs. These drive expression mostly, if not exclusively, in glial cells (Fig. 6). Consistent with this expression being dispensable for viability, an earlier study indicated that *slp2* and the flanking region are not required for viability (Cadigan et al., 1994a; Grossniklaus et al., 1992). However, flies can survive in the laboratory with clear CNS defects (Fujioka et al., 2003). A previous study (Mondal et al., 2007) suggested negative regulation of glial cell specification by *slp*. Our data suggest that *slp* is expressed in glial cells alongside Repo. Because 4 non-overlapping CRMs drive expression in glial cells, we suggest that *slp* has a separate, positive function in glial cells following their specification. Previous analysis of *slp*-related protein-coding sequences in non-drosophilid insects and basally branching arthropods suggested that the common ancestral coding sequence of *slp1* and *slp2* was more similar to *slp2* (Choe and Brown, 2007; Damen et al., 2005). We found that this is also true in the mosquito *A. gambiae*. A conserved nervous system function for *slp2* might help to explain why the *slp2* coding region has diverged more slowly than *slp1* from their common ancestral sequence.

Some of the stripe CRMs surrounding *slp1* also drive embryonic CNS expression (Figs. 1-3, S2), possibly in neuroblasts. Previous studies showed that *slp* is involved in specifying neuroblast identity (Bhat et al., 2000). These CRMs are good candidates for providing this function. In addition, multiple CRMs drive expression in the larval brain and in eye discs (Figs. 1, S2, S3).

Standard P-element transgenesis revealed that many of the CRMs surrounding *slp1* can cause pairing-sensitive silencing of *mini-white* in some transgenic lines (Fig. S2), a rare phenomenon that is usually associated with Polycomb-response elements (PREs). This may indicate a set of dispersed PREs in this region that facilitate the association of Polycomb with the locus, and maintain a chromatin domain enriched in histone H3 tri-methylated at lysine 27, which was found to be present throughout the *slp* locus in embryos (Negre et al., 2011; Schuettengruber et al., 2009).

Transgenic rescue suggests minimal redundancy among stripe CRMs

Our rescue construct is shorter at both ends than one previously tested (Cadigan et al., 1994a), yet generates a similar degree of rescue. In addition to containing upstream sequences extending into neighboring genes, the previous construct included our i1523 and i2330 stripe CRMs. Both constructs included the *slp1*, and not the *slp2*, coding region. There are several possible

explanations for the inability of both rescue constructs to completely rescue the denticle defects of the *slpΔ34* null mutant chromosome. One possibility is that the *slp2* transcript may be more stable than that of *slp1*, which would be consistent with the fact that *slp2* RNA normally appears to both begin expression and reach its maximum levels later (Grossniklaus et al., 1992). This could explain not only our inability to completely rescue function, but also the premature disappearance of *slp* RNA from our construct. The sufficiency of this explanation is argued against, however, by the fact that a *slp2*-specific mutation is probably viable (Grossniklaus et al., 1992), although the mild denticle defects resulting from rescue by our transgene might not cause lethality. Finally, maintenance of stripe expression may require sequences in the *slp* locus that do not themselves have enhancer activity, such as the maintenance elements within the *bithorax* complex (Maeda and Karch, 2009) or the *eve* gene (Fujioka et al., 2008). Testing of this possibility will require further study.

Recent studies of genes with apparently redundant enhancers (Frankel et al., 2010; Perry et al., 2010) suggest that true redundancy may be rare, and that distinct enhancers with overlapping activities contribute to phenotypic robustness that is likely to be maintained by natural selection. Our results are consistent with this, although they suggest that there is some redundancy among the stripe CRMs, as those downstream of *slp1* do not noticeably contribute to patterning the cuticle when all the upstream ones are present. However, the sequences within these elements appear to be conserved, suggesting they contribute to function, at least in the wild. Our results further suggest that even following a genomic duplication that generates partially redundant coding regions, redundant CRMs may be rapidly lost during subsequent evolution.

Interactions among CRMs

In almost all cases, our larger elements drive expression in all the places where expression is driven by smaller CRMs that they contain. As an example, the 2.1 kb u8766 drives expression both in the larval brain and in 14 stripes, consistent with the fact that it spans the 600 bp u8781 and the 900 bp u8172, which drive expression in the brain and in 14 stripes, respectively. Furthermore, most of the differences among partially redundant CRMs are consistent with their activities combining additively to generate endogenous *slp* expression. For example, while the regions u8172, u4734, and u3225 each drive a 14-stripe pattern in the ectoderm beginning at embryonic stage 7 or 8 (Figs. 3, 4), the regions u5547, u2316, and i1523 are expressed later, at stages 9–11. Thus, while there is considerable overlap among the striped patterns driven by these elements, they are not all redundant, and each may be important to produce the robust *slp* striped pattern in the endogenous context.

In contrast, some negative positional cues depend on more complex CRM interactions. A recent study (Prazak et al., 2010) described a detailed analysis of the u8172 region (whose 14-stripe

pattern includes some cells outside the normal *slp* expression domain). That study showed that u3125 (which drives a 7-stripe pattern with no ectopic expression), and derivatives of it, can suppress ectopic expression from u8172 when combined in the same construct. Our rescue data show that ectopic expression driven by the upstream CRM disrupts normal function (Fig. 5, u8150), and so must be suppressed within the endogenous locus. Thus, non-additive interactions among individual CRMs have important roles in regulating *slp* expression, even though the general trend is for the activities of *slp* CRMs to combine additively.

Another kind of interaction among CRMs is revealed in *slp* mutants that are rescued using the stripe CRMs located downstream of *slp1*, which do not drive an early 7-stripe pattern. Although i1530 drives a regular 14-stripe pattern in wild-type embryos (Fig. S1), in a *slp* mutant the longer i1539 drives expression strongly in only 7 stripes, and weakly in the other 7 (Fig. 5). This difference is explained by positive autoregulation, in that the early *slp* stripes are required for functional levels of later *slp* expression in the same cells. This is reminiscent of the positive autoregulation of *eve* stripes, which is indirect (Fujioka et al., 1995). Here, the late loss of *slp* expression in the absence of early *slp* stripes can be explained by expanded *odd* expression (Jaynes and Fujioka, 2004), which apparently represses later *slp* expression in every other stripe of the 14 stripe pattern. We have not localized the site of action of this *odd*-dependent repression, which could be either in the stripe CRM region downstream of *slp1*, or within the *slp1* promoter region, both of which are contained within this rescue construct. In either case, it is interesting to note that the 14-stripe pattern driven by these CRMs is regulated, at least initially, in a pair-rule fashion, with independent inputs to two interdigitated sets of 7 stripes.

The fact that there is such an indirect autoregulatory requirement for only half of the *slp* stripes highlights the pair-rule character of *slp* function in its intimate relationship with *eve* and *odd* (Jaynes and Fujioka, 2004), even though it is clearly also required in 14 stripes at later stages, where it has a similar mutual repressive relationship with *engrailed* (Cadigan et al., 1994b; Kobayashi et al., 2003). This example illustrates that the pair-rule genes are difficult to neatly classify into early and late classes because of the complexity of their interactions both with gap genes and with each other. A recent study (Schroeder et al., 2011) placed *odd*, which had traditionally been classified as a secondary pair-rule gene, into the "early" class, while *slp* was assigned to the "late" class. Despite the fact that *odd* participates directly in translating non-periodic pattern information into periodic pattern, while *slp* does not, *slp* nonetheless regulates *odd* after periodic pair-rule patterns have been established. This secondary cross-regulation, which formally goes "backwards" in the hierarchy, is essential for the correct transition to segment polarity gene control. Specifically, without early 7-stripe

slp expression, half of the *wg* stripes are not established (those that coincide with the "missing" *slp* stripes), and the adjacent parasegment borders decay, resulting in pair-rule defects (Fig. 5, i1539 rescue). Thus, complex regulatory interactions occur at both the early pair-rule stage and the late pair-rule stage, and may be the norm for developmental processes.

Stage-dependent regulation of *slp* CRMs

The 7- and 14-stripe *slp* patterns occur at different stages, and are driven in part by separable elements. Among the 14-stripe CRMs, some drive earlier expression, which overlaps in time with expression driven by the later-acting CRMs. This suggests that different combinations of activators, and possibly different repressors, may be responsible for activating, and restricting the activity of, these elements at different stages. This, in turn, provides a rationale for the existence of multiple regulatory elements with temporally overlapping patterns. As the expression of activators change during development, maintenance of expression within a given cell is subject to changing constraints on the relevant CRMs. In particular, the need to maintain both the on state and the off state in the appropriate cells may limit the ability of a single CRM to respond properly at all stages, making it advantageous to utilize different CRMs as the milieu of trans-acting factors changes within the nucleus.

Evolution of *slp* CRMs

We used BLAST searches to map sequence similarities for each stripe CRM among the sequenced drosophilid genomes, all of which contain both *slp1* and *slp2* coding regions, in a similar arrangement to that in *D. melanogaster*. The highest-stringency similarity was found between two CRMs expressed in stripes in the presumptive mesoderm, u1609 and i2330. Analysis of likely transcription factor binding to this 12 bp sequence based on known specificities did not reveal any specific factors with a pattern of expression suggesting regulation of these CRMs. However, the arrangements of best-match sequences to each stripe CRM in the most distantly related drosophilids suggest that ancestral sequences for each stripe CRM existed separately in their common ancestor (see Fig. S4 for a map of the relative locations of these cross-species similarities). However, whether these apparently conserved sequences represent distinct, ancestral CRMs with functions similar to those in *D. melanogaster* remains an open question.

We also tried, without success, to find clear evidence of homologies to stripe CRMs in the next-most closely related sequenced genome, that of *A. gambiae*, which might indicate an ancestral element from which more than one drosophilid CRM evolved. Although numerous short sequence similarities were found, their arrangements did not suggest any specific relationship to a drosophilid

CRM. Presumably, future analysis will reveal how the locus evolved, when sequenced genomes become available for species that diverged from the drosophilids more recently than mosquitoes.

Shared conserved sequence clusters have previously been found in some CRMs with similar expression patterns (Brody et al., 2007, 2008). Therefore, we used Evoprinter and cis-Decoder to look for CSCs both within *slp* CRMs and between different CRMs. CSCs were found within many of them, but none of these CSCs were shared among co-expressed CRMs. We suggest that this may be different from the situation among enhancers active in neuroblasts, for example, because the *slp* CRMs may have evolved by convergent evolution under conditions where the available pool of DNA binding activators was large enough to preclude convergence to a similar set of sequence clusters.

Acknowledgements

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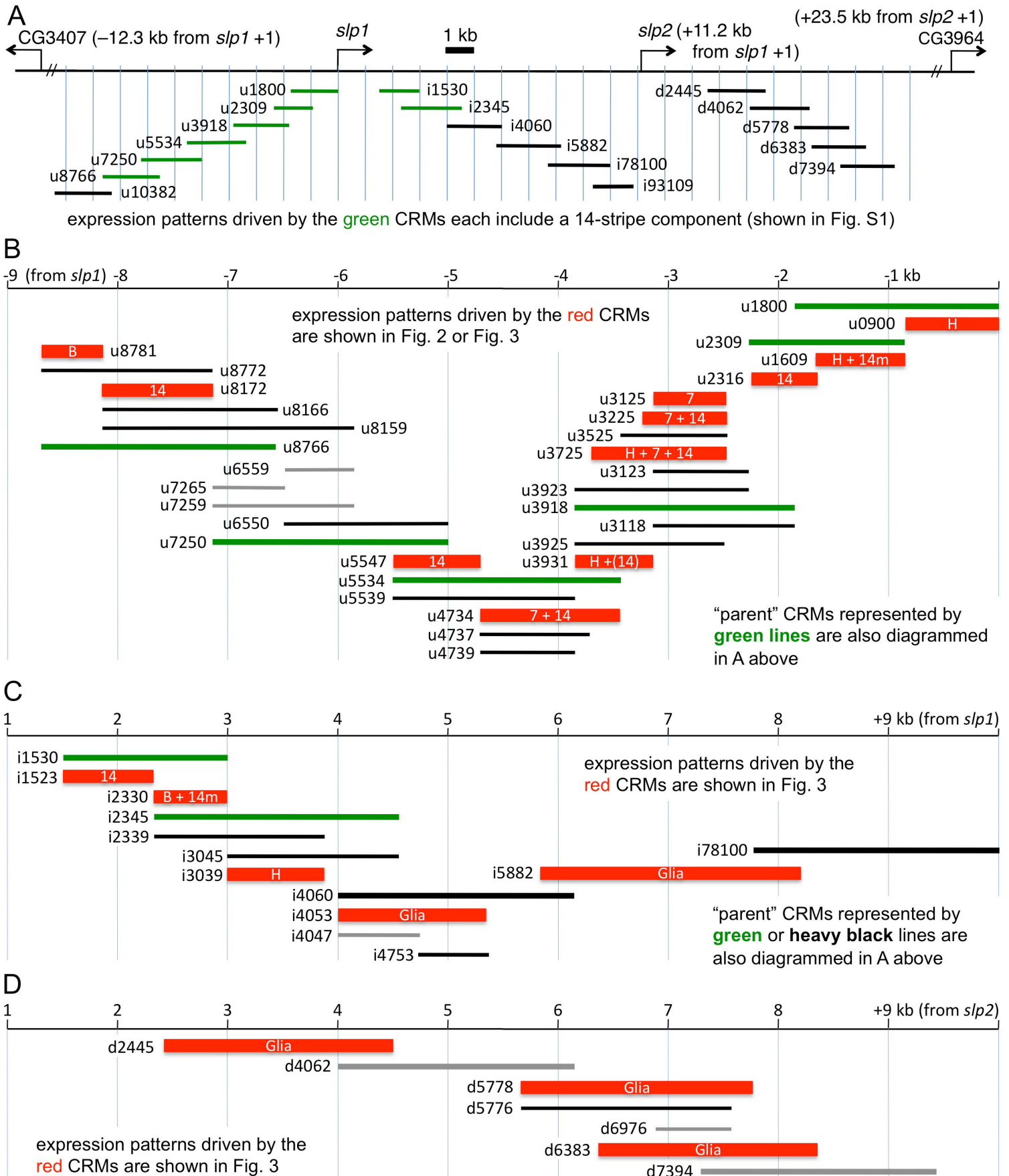
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Figures

Fig. 1. Mapping of CRMs in the *slp* locus. The 30.9 kb genomic region from 10.3 kb upstream of *slp1* to 9.4 kb downstream of *slp2* was analyzed for enhancer activity in transgenic reporters. **A**: Large-scale mapping, using approximately 2 kb fragments overlapping by about 500bp. Bars indicate locations of the fragments. Locations are based on Flybase coordinates (Tweedie et al., 2009) (see Fig. S2). **B-D**: Diagram of fragments used for finer scale dissection of the region upstream of *slp1* (B), between *slp1* and *slp2* (C), and downstream of *slp2* (D). The smallest fragments found to drive consistent reporter gene expression in transgenic lines are diagrammed as red boxes, with internal lettering indicating the pattern: "B", larval brain; "7", 7 stripes (at stage 7); "14", 14 stripes (at stage 7 or later); "14m", 14 stripes restricted to the mesoderm; "(14)", 14 weak stripes; "H", strong embryonic head stripe (stage 6); "Glia", glial cells (in late embryonic CNS). These expression patterns are shown in other figures. Those regions found to drive no consistent expression are indicated as gray lines. Thin black lines are regions that drive expression like the sum of the smaller elements that they contain, except where noted in the text.

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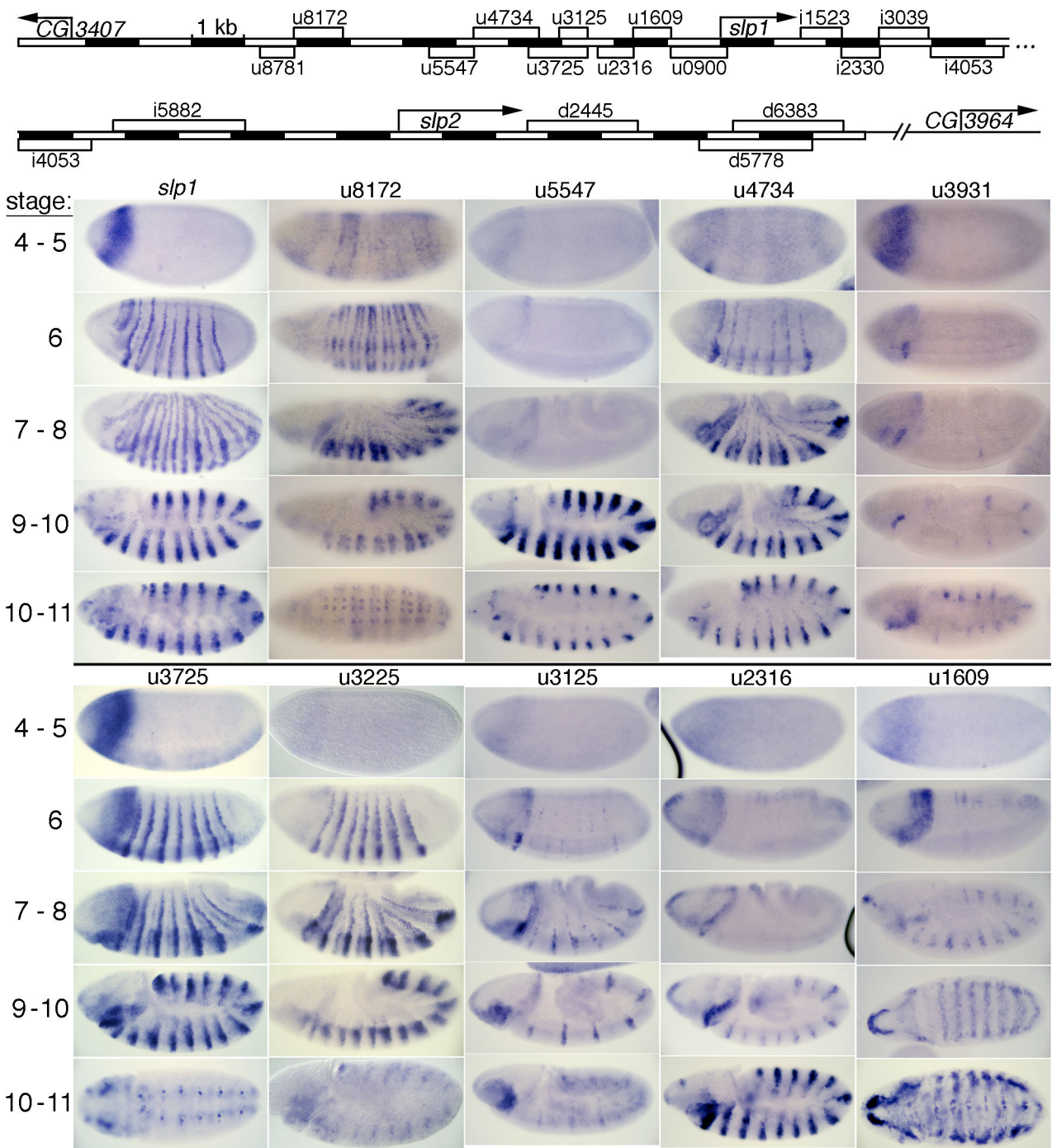
Fig. 2. CRMs upstream of *slp1* drive germband stripe and head expression. Map at the top shows the locations of the shortest identified sequence blocks that drive aspects of the *slp* pattern. The 1st column shows endogenous *slp* expression. The other columns show *lacZ* expression from transgenes carrying each indicated CRM upstream of *slp1-promoter-lacZ* (see Materials and Methods) at the 5 embryonic stages shown at the left. Note that u3225 (which has the same 3' end point as u3125) is not shown on the map.
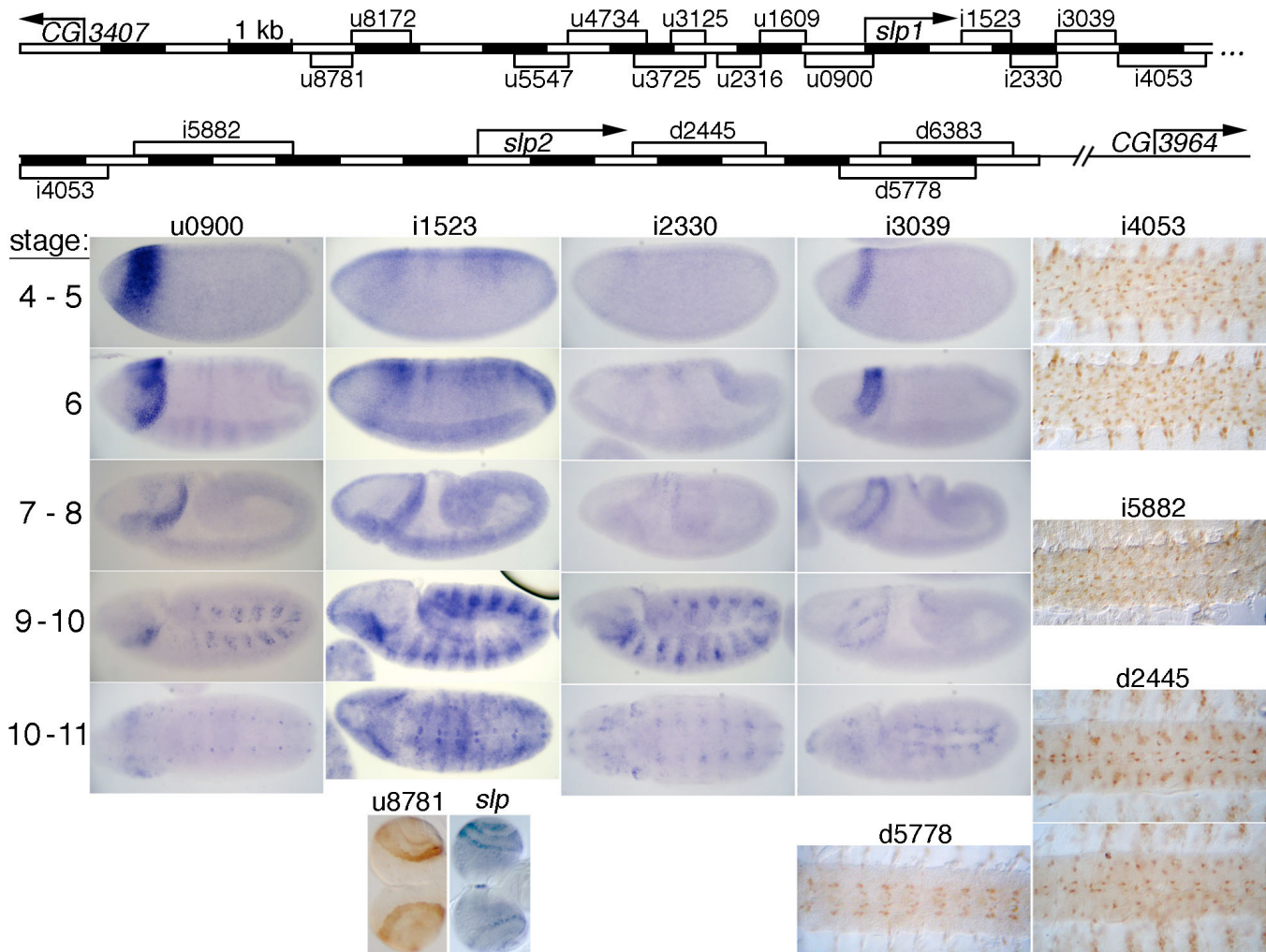
Fig. 3. CRMs downstream of but nearby *slp1* drive germband stripe and head expression, while those closer to *slp2* drive CNS expression. Map at the top shows the locations of the shortest identified sequence blocks that drive aspects of the *slp* pattern. Both "i" and "d" CRMs were analyzed upstream of *slp2-promoter-lacZ,* while u8781 was upstream of *slp1-promoter-lacZ.* In the first 3 columns, expression patterns of *lacZ* RNA from transgenes carrying each CRM are shown at 5 (or 6) embryonic stages (listed at the left), while the last 2 columns show expression of β-galactosidase (β-gal) in the embryonic CNS, or, for u8781, in 3rd instar larval optic lobes. The panel next to u8781 shows endogenous *slp* expression in the optic lobes.

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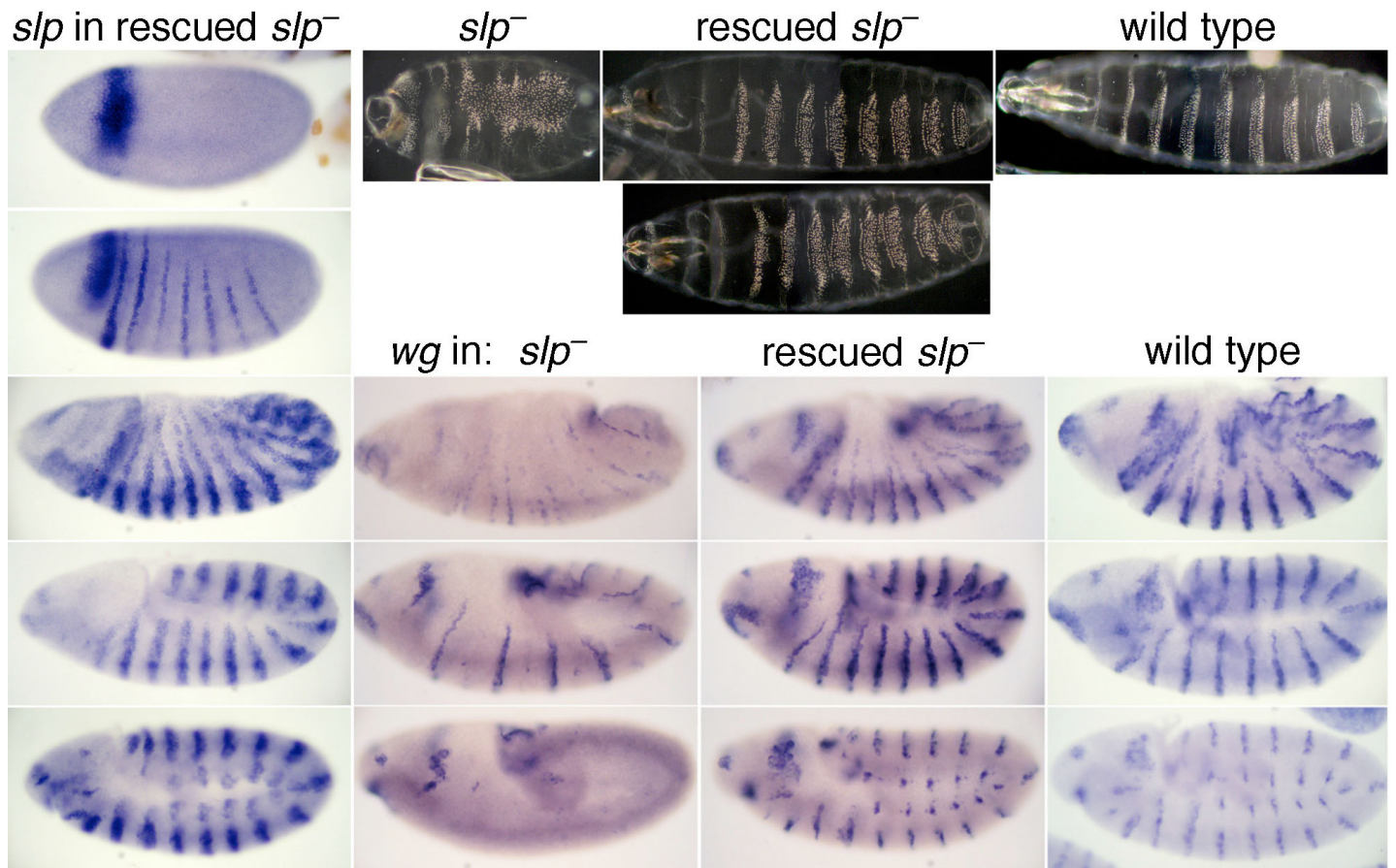
Fig. 4. Transgenic rescue of *slp* expression and function. **1st column**: embryonic expression of *slp* RNA in a *slp* RNA null mutant (*slpΔ34*) with a transgene carrying the *slp* locus from −8.1 to +1.5 kb relative to the *slp1* TSS, which includes 78 bp 3' of the *slp1* mRNA polyA signal. **2nd column**: cuticle pattern (top) and *wg* RNA expression (lower) in the same *slp* RNA null mutant as in column 1, with no rescue construct. **3rd column**: two cuticles representing the range of phenotypes seen (top), and the *wg* RNA pattern (lower), in the rescued *slp* mutant of column 1. Note the near-complete rescue. **4th column**: Cuticle pattern and *wg* RNA expression in wild type.
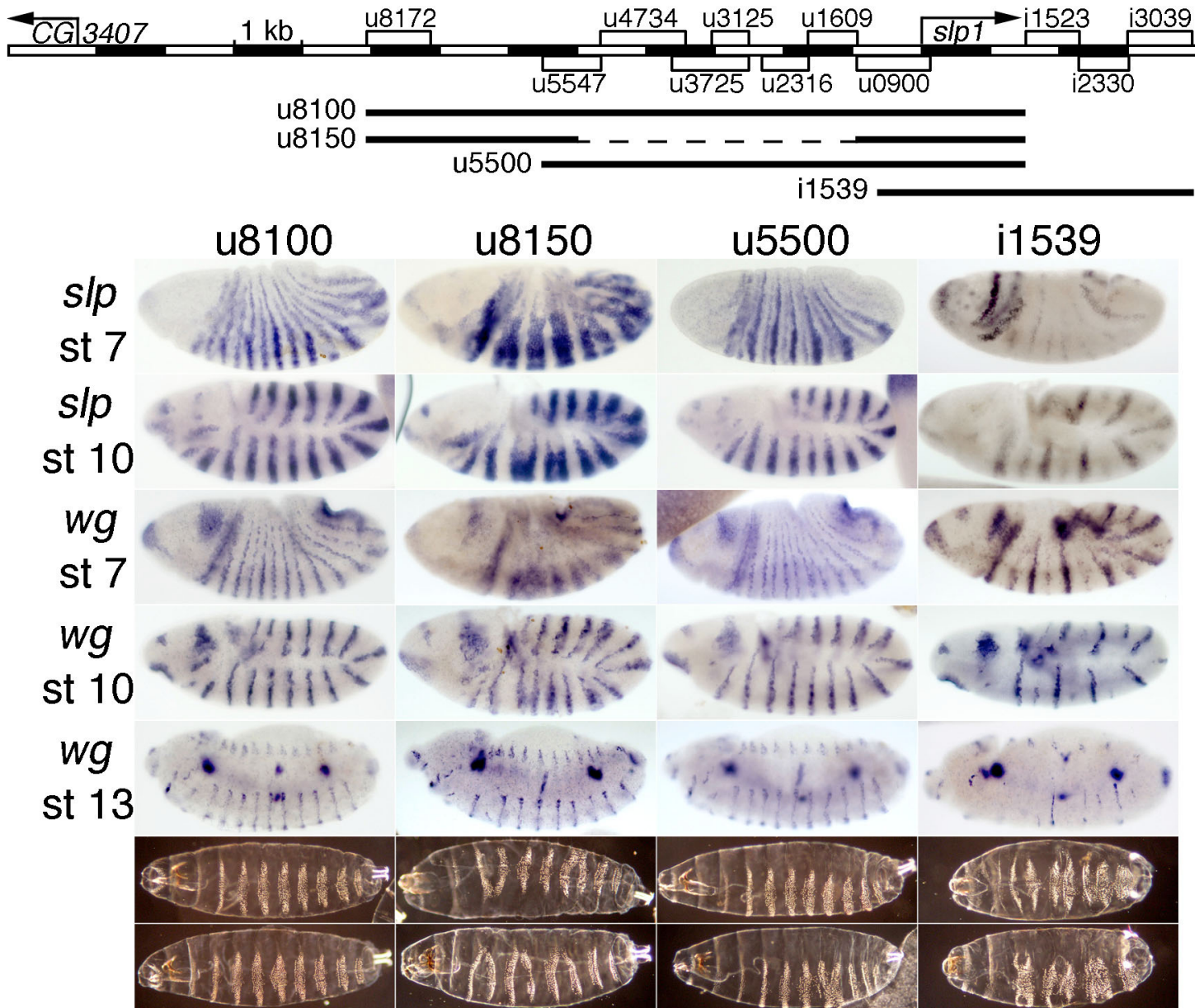
Fig. 5. Transgenic rescue of *slp* expression and function by subsets of CRMs. The map at the top indicates the extent of each rescue construct as a line below the map of all the stripe CRMs of the *slp* locus. The panels below show embryonic expression of *slp* RNA at stages 7 (top row) and 9-10 (2nd row), *wg* expression at stages 7, 10, and 13-14 (indicated on the left), and cuticles at the end of embryogenesis (bottom rows show the range of patterns seen) in a *slp* RNA null mutant (*slpΔ34*) with a transgene carrying the *slp* locus from: **1st column**: −8076 to +1539 bp relative to the *slp1* TSS, which includes 78 bp 3' of the *slp1* mRNA polyA signal. Note the near-complete rescue (see Fig. 4 for wild type). **2nd column**: −8076 to −5000, fused with −940 to +1539 bp. Note the ectopic expression of both *slp* and *wg* at stages 7 and 10, and the partial loss of *wg* and naked cuticle at later stages. **3rd column**: −5510 to +1539 bp. Note the near-complete rescue, with more severe defects in the denticle pattern in some embryos, relative to column 1. **4th column**: −665 to +3934 bp. Note the loss of expression of both *slp* and *wg*, more severely in alternate parasegments, at all stages (see text), and the pair-rule loss of naked cuticle. All constructs were analyzed at the same chromosomal location (see Material and Methods).
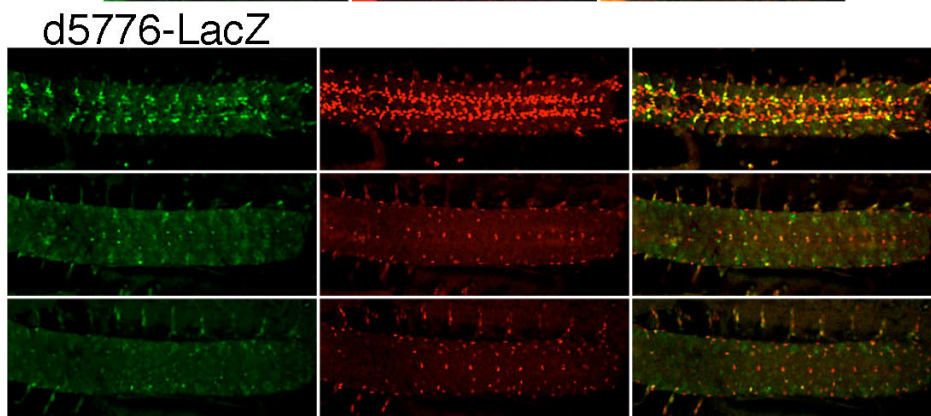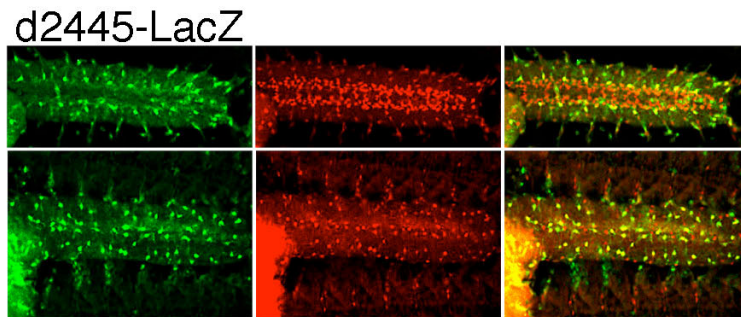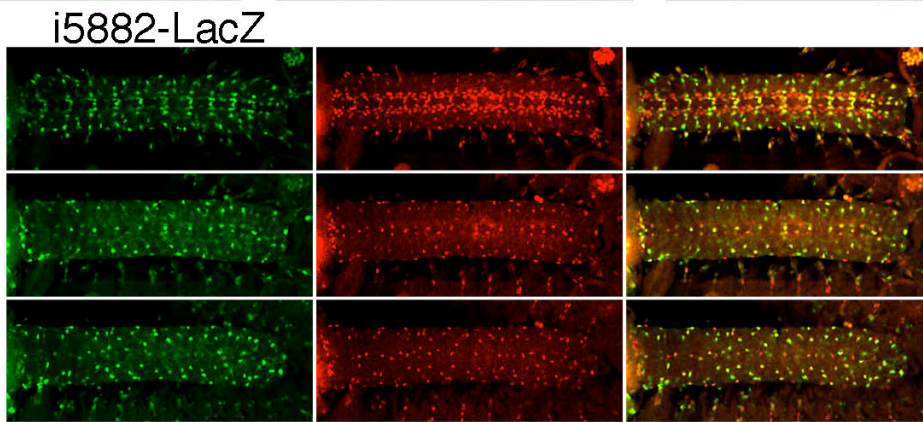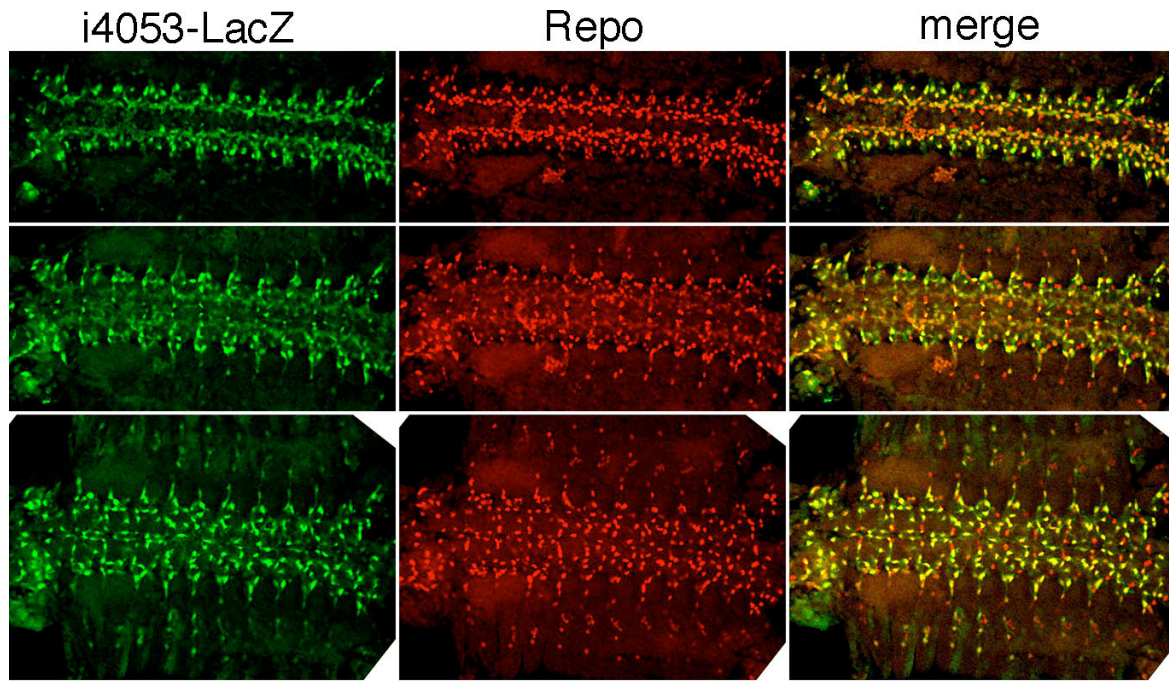
Fujioka, M., & Jaynes, J. B. (2012). Regulation of a duplicated locus: Drosophila *sloppy paired* is replete with functionally overlapping enhancers. *Developmental Biology, 362*(2), 309-319.

Fig. 6. Central nervous system CRMs both upstream and downstream of *slp2* drive expression in glial cells. **1st column**: β-gal expression from the indicated CRM-carrying transgene. **2nd column**: expression of the glial cell-specific protein Repo. **3rd column**: merged view of β-gal (green) and Repo (red). Either 2 or 3 focal planes within a dissected embryonic CNS are shown for each: i4053 is at stage 13, the others are at stage 15 (when the CNS has become condensed). Note that each of these *slp* CRM activities overlap extensively with Repo expression.
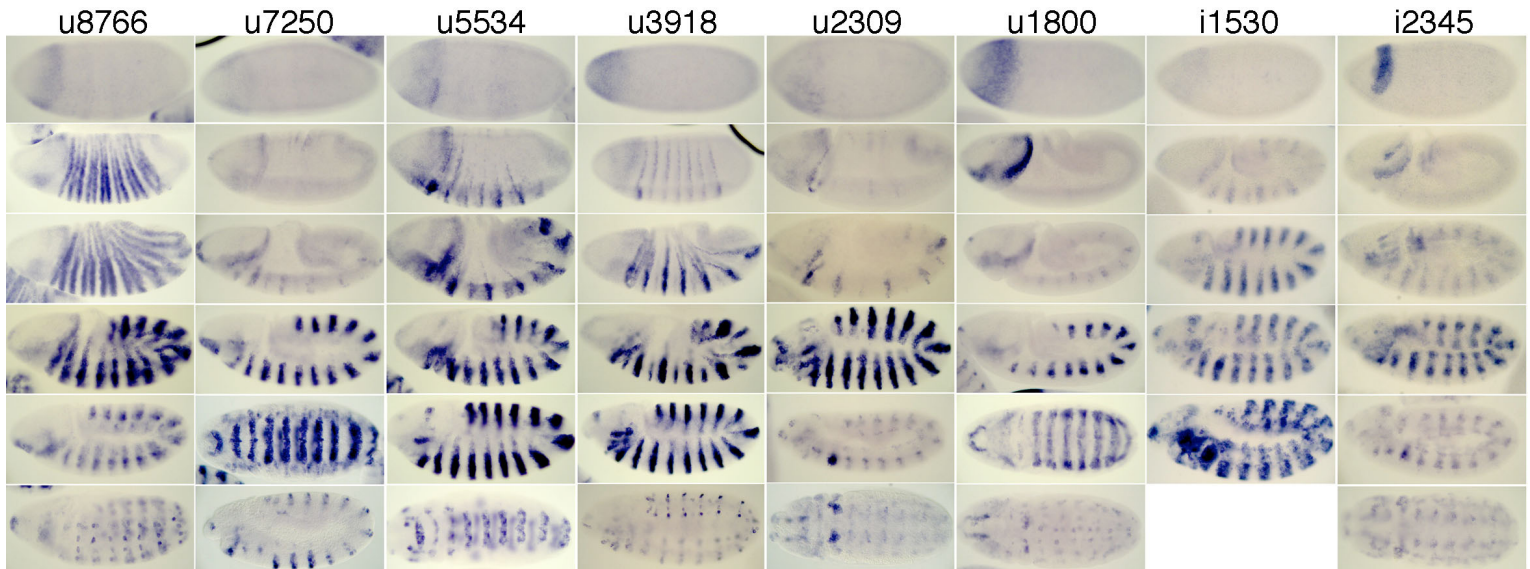
Fig. S1. Transgenic dissection of regulatory DNA upstream of *slp1* and in the *slp1-slp2* intervening region. Each column shows the *lacZ* RNA expression pattern driven by the CRM listed at the top, in the context of a reporter transgene, at various embryonic stages, depending on when the CRM is active. The "u" fragments were placed upstream of the *slp1-promoter-lacZ* reporter, while the "i" fragments were upstream of *slp2-promoter-lacZ* (see Material and Methods for details). See the main text for our naming scheme and Fig. 1A for a map of the CRMs used.

Fig. S2 Fujioka and Jaynes, 2012   this file contains 2 pages

This paper transgene	relative to TSS: of slp1 or slp2 5'	3'	R. enzyme sites 5'	3'	Flybase (r5.22) coordinates		sequence of: 5' end	3' end, reverse complement	summary of embryonic expression in **stripes**
u10382	-10254	-8175			3815426	3817505	CGCAAAAAGACTGCGGCTGTCAACATC	CACGGCTCAGTGGCCTTCGAAACAATC	No stripe expression.
u8766	-8713	-6509			3816967	3819171	TAGAGGCCTTCAAAGGTTTGCTAAAGTTT	GGCTAACTGACTCAAGTACTAGTTGAAATC	See Fig. S1.
u7250	-7174	-4993			3818506	3820687	GTGAAAATCCTAGCACTGCGACGAATTC	CGCCTTATCATCGTCGCCGTGATTTTC	See Fig. S1.
u5534	-5512	-3440			3820168	3822240	GCGTCCACAGTCCACAGTCCGCAGGA	CTGCACTTCAGGTAGTGGGGGTAAATC	See Fig. S1.
u3918	-3929	-1777			3821751	3823903	TGTCTGGCAATCCCGATGCCAACCGCA	ACAATGGAGGAGTTGGGAAGAAAAAA	See Fig. S1.
u2309	-2324	-941		Nru1	3823356	3824739	CCCATTACTCTATGCAGTGTGCATAAA	CGACAGATTTGCGAATTAGTTCTGGG	See Fig. S1.
u1800	-1829	121			3823851	3825800	CTACTGCGTTAAAAGTGCGCTGCCTAC	ATTTCACTTTTCACCATTTTTCTGATT	See Fig. S1.
i1530	1517	3007			3827196	3828686	GGATCGAGTTATCGACACTTGAAAAA	GGTAACCCCAACCTGTTAACTGATTTA	See Fig. S1.
i2345	2291	4545		Spe1	3827970	3830224	AGGGCCTCAATTTGAAAGATG	CTAGTGGTGCACCCATGTGGTGGATTT	See Fig. S1.
i4060	3950	6140			3829629	3831819	CGGATCCTGGCATATAGATCATACATA	GTCACGCGTAAGACTTTCGAGCATTA	No stripe expression.
i5882	5761	8249			3831440	3833928	GCTCGAAAGATCTGGCTGATGGAATCCCCAC	ATGCTAGCTACCGTTTCGAAAGAGTTAAG	No stripe expression. See i78100 note.
i78100	7797	9941			3833476	3835620	ACGCGTCAGAGCGAAAAACGAGAGG	ACTCGTCGGTCGTGCGATTGGATCCCA	Weak head stripe expression. Contains Bcd binding site cluster according to Ochoa-Espinosa et al., 2005 (slpC, see below). A head stripe is driven by Schroeder et al. 2004's slp2-3, which contains slpC and spans the junction of i5882 and i78100.
i93109	9279	10851			3834958	3836530	ACTGAATACCACCCTCATAGC	GGGTCACCCTTTTCCGGATTGTGGC	No stripe expression.
d2445	**2444**	**4523**			3839285	3841364	CGCGACAAACTTGTCGGCAATCCGAC	GTCCCGCTCCAGATTAAGCTGCGACTA	No stripe expression.
d4062	**3985**	**6174**			3840826	3843015	GCAGCCCAACAACTGTGACTGGCAT	CTGCGACGGAGTTGTGCCACTGAATTT	No stripe expression.
d5778	**5689**	**7819**			3842530	3844660	GGCCAGAAGTTGGCCAATAAGTC	GACATCAGTGGCTCCTTTTGGTGCCAC	No stripe expression.
d6383	**6273**	**8346**			3843114	3845187	TGTGCGCACTGTGGGAACTTTGGAT	TCCAGATTCTGCGCTTTGTGCACTGTA	No stripe expression.
d7394	**7257**	**9429**			3844098	3846270	CCGAGCTGTCAACACAGATTCCAAAC	GAAAACACTGCGCCCGTAGCATTCAC	No stripe expression.
u8781	-8713	-8072		Bgl2	3816967	3817608	TAGAGGCCTTCAAAGGTTTGCTAAAGTTT	GATCTTCAATCACATCCCTTTGCG	No stripe expression.
u8772	-8713	-7148		EcoRI	3816967	3818532	TAGAGGCCTTCAAAGGTTTGCTAAAGTTT	AATTCGTCGCAGTGCTAGGATTTTC	Similar to u8172.
u8172	**-8075**	**-7148**	Bgl2	EcoRI	3817605	3818532	GATCTTCAGGATATTTCAGCACATGCAT	AATTCGTCGCAGTGCTAGGATTTTC	See Fig. 2. Slightly weaker than u8766. No mesodermal stripes.
u8166	-8075	-6509	Bgl2		3817605	3819171	GATCTTCAGGATATTTCAGCACATGCAT	GGCTAACTGACTCAAGTACTAGTTGAAATC	Similar to u8766, except mesodermal stripes are much weaker.
u8159	-8075	-5868	Bgl2	Pvu2	3817605	3819812	GATCTTCAGGATATTTCAGCACATGCAT	CTGGCAAATTTTGATGACCAATGGTTGAG	Similar to u8166.
u6559	-6530	-5868	Spe1	Pvu2	3819150	3819812	CTAGTACTTGAGTCAGTTAGCCATATACTT	CTGGCAAATTTTGATGACCAATGGTTGAG	Weak head stripe expression.
u7265	-7174	-6527	EcoRI	Spe1	3818506	3819153	GTGAAAATCCTAGCACTGCGACGAATTC	CTAGTTGAAATCAATCAATATAATGGGTATT	Weak head stripe expression.
u7259	-7174	-5868	EcoRI	Pvu2	3818506	3819812	GTGAAAATCCTAGCACTGCGACGAATTC	CTGGCAAATTTTGATGACCAATGGTTGAG	Weak head stripe expression.
u6550	-6530	-4993	Spe1		3819150	3820687	CTAGTACTTGAGTCAGTTAGCCATATACTT	CGCCTTATCGTCGTCGCCGTGATTTTC	Similar to u5547.
u5547	**-5512**	**-4671**		Nhe1	3820168	3821009	GCGTCCACAGTCCACAGTCCGCAGGA	CTAGCTCCCTTCACTCTTCTTCGAT	See Fig. 2.
u5539	-5512	-3937		Nsi1	3820168	3821743	GCGTCCACAGTCCACAGTCCGCAGGA	TAAAAACCTGGAAGGATCTCGTCGGCT	Similar to u5534.
u4734	**-4675**	**-3440**	Nhe1		3821005	3822240	GCTAGCTGGGAGATTGATCTACGAAT	CTGCACTTCAGGTAGTGGGGGTAAATC	See Fig. 2.
u4737	-4675	-3720	Nhe1	Nde1	3821005	3821960	GCTAGCTGGGAGATTGATCTACGAAT	TATGGAAGCAGCTGGAGATCGTAAGCAACC	SImilar to u4734.
u4739	-4675	-3937	Nhe1	Nsi1	3821005	3821743	GCTAGCTGGGAGATTGATCTACGAAT	TAAAAACCTGGAAGGATCTCGTCGGCT	Similar to u4734, except that stripes expand posteriorly into the En domain.
u3931	**-3929**	**-3056**			3821751	3822624	TGTCTGGCAATCCCGATGCCAACCGCA	ATCTTCGGCATTTGTCGCGCACAGC	See Fig. 2.
u3125	**-3055**	**-2522**	EcoRV	Aat2	3822625	3823158	ATCCTGCTGATGTCGCAGCCATGTGACA	CGTGGTAAAACGAATTGATTAGCGA	See Fig. 2.
u3925	-3929	-2522		Aat2	3821751	3823158	TGTCTGGCAATCCCGATGCCAACCGCA	CGTGGTAAAACGAATTGATTAGCGA	Similar to u3725.
u3118	-3055	-1777	EcoRV		3822625	3823903	ATCCTGCTGATGTCGCAGCCATGTGACA	ACAATGGAGGAGTTGGGAAGAAAAAA	Similar to u3125, + extra weak stripes in odd-numbered parasegments.
u3923	-3929	-2303			3821751	3823377	TGTCTGGCAATCCCGATGCCAACCGCA	GCACACTGCATAGAGTAATGGGTAATTCGA	Similar to u3725.
u3123	-3055	-2303	EcoRV		3822625	3823377	ATCCTGCTGATGTCGCAGCCATGTGACA	GCACACTGCATAGAGTAATGGGTAATTCGA	Similar to u3125.
u3725	-3679	-2522	Nde1	Aat2	3822001	3823158	TATGGTTAGGAAATAAAATAAAATTTAGAT	CGTGGTAAAACGAATTGATTAGCGA	
u3525	-3455	-2522		Aat2	3822225	3823158	ACTACCTGAAGTGCAGATTTGGCAGCAGTAA	CGTGGTAAAACGAATTGATTAGCGA	Similar to u3725, except that stage 10 is weaker, and CNS expression is missing.
u3225	-3243	-2522	Nsi1	Aat2	3822437	3823158	GCATAAAAATCACTGCACATTTTTTTCATCG	CGTGGTAAAACGAATTGATTAGCGA	See Fig. 2. Similar to u3918, except that stripes are expanded anteriorly (indicating possible removal of Cubitus Interruptus binding sites).
u2316	**-2324**	**-1648**		EcoRV	3823356	3824032	CCCATTACTCTATGCAGTGTGCATAAA	ATCTTAGACAACTTGACAAAACAATCA	See Fig. 2.
u1609	**-1647**	**-941**	EcoRV	Nru1	3824033	3824739	ATCAAACGCACAGTGCCGCCGACAAGTA	CGACAGATTTGCGAATTAGTTCTGGG	See Fig. 2.
u0900	**-940**	**121**	Nru1		3824740	3825800	CGAAACTAGAACAAATCCCGG	ATTTCACTTTTCACCATTTTTCTGATT	See Fig. 3. The weak, dorsally restricted stripes may be caused by sequences within pCaSpeR.
i1523	**1517**	**2293**		Stu1	3827196	3827972	GGATCGAGTTATCGACACTTGAAAAA	CCTAGAAAAGTCCCTGGCTTACGCT	See Fig. 3. Stripes are expanded anteriorly relative to both i1530 and endogenous *slp*.
i2330	**2291**	**3007**	Stu1		3827970	3828686	AGGGCCTCAATTTGAAAGATG	GGTAACCCCAACCTGTTAACTGATTTA	See Fig. 3.
i2339	2291	3930		Sph1	3827970	3829609	AGGGCCTCAATTTGAAAGATG	CTGGGTAATCAGTGCTCGGCATCTGCTG	Combined expression of i2330 and i3039.
i3039	**3002**	**3930**	BstE2	Sph1	3828681	3829609	GTTACCTCATCTTTTTGTAATGAAATGCT	CTGGGTAATCAGTGCTCGGCATCTGCTG	See Fig. 3.
i4053	**3950**	**5319**		Xba1	3829629	3830998	CGGATCCTGGCATATAGATCATACATA	TCCAGCTGGAGAATAACATGAGGAGCTA	See Fig. 3 and Fig. 6.
i4047	3950	4710		Dra3	3829629	3830389	CGGATCCTGGCATATAGATCATACATA	GTGGCTGCCACTTGGTTTGGTTCTCCATGG	No stripe expression.
i4753	4714	5319	Dra3	Xba1	3830393	3830998	GTGACCACAAAATGTGTTTTTAATTGAAACTG	TCCAGCTGGAGAATAACATGAGGAGCTA	No stripe expression.
d5776	**5689**	**7567**			3842530	3844408	GGCCAGAAGTTGGCCAATAAGTC	ATTTAGGTCTTGAAGCACTGTTCCTGCTGT	No stripe expression.
d6976	**6915**	**7567**			3843756	3844408	TCATCATTGAGATGCCCCGAGATCTCTCGGC	ATTTAGGTCTTGAAGCACTGTTCCTGCTGT	No stripe expression.
slp1-lacZ	-261	121			3825419	3825800	TCCCAAACAACGTCTACAAACATATA	ATTTCACTTTTCACCATTTTTCTGATT	
slp2-lacZ	**-314**	**373**			3836528	3837214	CCCACTACATGAGAAATGGGGTGTAC	TCCTCGATCTTCACCATGATTCTTAGG	
Lee and Frasch (2000)									
slp5'-4	-5400	-4664			3820280	3821016	GCCCAAAGTCGGTCGAACAG	CTCCCAGCTAGCTCCCTTCA	
slp5'-3	-3986	-3344			3821694	3822336	CCCTTTATAATTCGATCAACTC	GAATGAATCCTGCTACTTGG	
slp5'-2	-2557	-985			3823123	3824695	AGGATCTCGAATCGCTAATC	ATGTGGAATCTCTGGCAATCC	
slp5'-1	-1910	-985			3823770	3824695	CTATCGCTCGATTGCCGTCTC	ATGTGGAATCTCTGGCAATCC	
Ochoa-Espinosa et al. (2005)									
slpA	-1077	-711			3824603	3824969	GCCAGCCAACACTG	CTGCGAACCACTGCACTAGCG	
slpB	3139	3931			3828818	3829610	GCAGCTCCCAAAGAAGCAC	GCTGGGTAATCAGTGCTCGGCATC	
slpC	7997	9005			3833676	3834684	CTGTCTAGGGAGCTTCAAGCG	GGCTAAATAGCAGCCAAATCC	Bcd binding site cluster identified here, but no expression data shown. Our i78100 contains this, and drives weak head expression.
Schroeder et al. (2004)									
slp2-3	7018	9657			3832697	3835336	GGTGTTGTTGGCTCTGCGTGAGTT	CGAGCAAATAGCCCCCACATTCAA	Drives head stripe expression, and contains Ochoa-Espinosa et al.'s slpC. Spans the junction between our i5882 and i78100, the latter of which drives weak head expression.

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comments on **embryonic CNS** expression	comments on expression in **optic lobes** at larval stage 3 (3L)	comments on expression in **eye discs** at 3L	comments on patterned eve color and **pairing sensitive silencing** (PSS) of *mini-white*	transgene
No consistent CNS expression.	Similar to u8781 (Fig. 3). Confirmed in 3 lines.	Similar to u8766 (expression stronger dorsally and ventrally, as in Fig. S2). Confirmed in 3 lines.	1 out of 8 lines showed patterned eye.	u10382
Fig. S1. Expression, possibly in neuroblasts, after stripes fade.	7 out of 9 lines similar to u8781.	Fig. S2. Expression stronger dorsally and ventrally. Confirmed in 9 lines.	7 out of 21 lines showed patterned eye.	u8766
No consistent CNS expression.	No consistent expression.	No consistent expression in eye discs.	3 out of 15 lines showed patterned eye. 4 out of 11 lines showed PSS.	u7250
Weak expression, possibly in neuroblasts, after stripes fade.	No consistent expression.	No consistent expression in eye discs.	6 out of 13 lines showed patterned eye.	u5534
Fig. S1. Expression, possibly in neuroblasts, after stripes fade. Similar to u2309.	No consistent expression.	No consistent expression in eye discs.	1 out of 16 lines showed patterned eye. One variegated.	u3918
Fig. S1. Expression, possibly in neuroblasts, after stripes fade.	No consistent expression.	No consistent expression in eye discs.	4 out of 10 lines showed patterned eye. 4 out of 7 lines showed PSS.	u2309
Fig. S1. Expression, possibly in neuroblasts, after stripes fade. Similar to i1523.	No data for 3L.	No data.	4 out of 19 lines showed patterned eye. 2 out of 15 lines showed PSS.	u1800
No data.	Fig. S2. Confirmed in 2 lines.	Expressed posterior to MF (morphogenetic furrow).	0 out of 5 lines showed patterned eye.	i1530
Fig. S1. Expression in ventrolateral cell clusters. Similar to i2330.	Similar to Fig. S2. 3 out of 4 lines showed the pattern.	Expressed posterior to MF.	0 out of 10 lines showed patterned eye.	i2345
Similar to i4053.	No consistent expression.	Fig. S2. Expression stronger dorsally and ventrally. Confirmed in 2 lines.	0 out of 6 lines showed patterned eye.	i4060
Fig. 3 and Fig. 6. Expression in glial cells.	No data for 3L.	No data for 3L.	2 out of 17 lines showed patterned eye. 2 out of 14 lines showed PSS.	i5882
No consistent CNS expression.	1 line showed no expression.	1 line gave expression posterior to MF.	0 out of 10 lines showed patterned eye. 2 out of 9 lines showed PSS.	i78100
Weak ubiquitous expression in CNS at late st 11-12. Expression similar to i2330 emerges.	No data for 3L.	No data for 3L.	0 out of 3 lines (2 out of 6 lines when the element was inverted) showed patterned eye. 1 out of 1 line (2 out of 3 lines when the element was inverted) showed PSS.	i93109
See Fig. 3 and Fig. 6. Expression in glial cells.	No consistent expression.	Expressed posterior to MF.	0 out of 13 lines showed patterned eye.	d2445
No consistent CNS expression.	No data for 3L.	No data for 3L.	1 out of 6 lines showed patterned eye.	d4062
Fig. 3. Expression in glial cells.	No data for 3L.	No data for 3L.	0 out of 6 lines showed patterned eye.	d5778
Fig. 3. Expression in glial cells.	No data for 3L.	No data for 3L.	0 out of 6 lines showed patterned eye.	d6383
No consistent CNS expression.	No data for 3L.	No data for 3L.	0 out of 13 lines showed patterned eye.	d7394
No consistent CNS expression.	Fig. 3. Confirmed in 6 lines.	4 out of 6 lines similar to u8766.	3 out of 17 lines showed patterned eye. 1 out of 11 lines showed PSS.	u8781
Similar to u8172.	Similar to u8781. Confirmed in 4 lines.	Similar to u8766. Confirmed in 4 lines.	0 out of 4 lines showed patterned eye.	u8772
Fig. 2. Similar to u8766, except expression appears in En stripes as well.	1 line somewhat similar to u8781, but the position may not be the same.	Expressed posterior to MF.	0 out of 17 lines showed patterned eye.	u8172
Similar to u8766.	No consistent expression.	Expressed posterior to MF.	1 out of 13 lines showed patterned eye.	u8166
Similar to u8766.	No data for 3L.	No data for 3L.	0 out of 11 lines showed patterned eye.	u8159
No consistent CNS expression.	No data for 3L.	No data for 3L.	0 out of 6 lines showed patterned eye.	u6559
No consistent CNS expression.	No data for 3L.	No data for 3L.	0 out of 4 lines showed patterned eye.	u7265
No consistent CNS expression.	No data for 3L.	No data for 3L.	0 out of 5 lines showed patterned eye.	u7259
No consistent CNS expression.	No data for 3L.	No data for 3L.	1 out of 8 lines showed patterned eye. 1 out of 5 lines showed PSS.	u6550
Weak expression, possibly in neuroblasts, after stripes fade.	No data for 3L.	No data for 3L.	2 out of 7 lines showed patterned eye. 1 out of 5 lines showed PSS.	u5547
Weak expression, possibly in neuroblasts, after stripes fade.	No data for 3L.	No data for 3L.	1 out of 10 lines showed patterned eye.	u5539
Similar to u2309.	No data for 3L.	No data for 3L.	0 out of 5 lines showed patterned eye.	u4734
Weak expression, possibly in neuroblasts, after stripes fade.	No data for 3L.	No data for 3L.	0 out of 6 lines showed patterned eye.	u4737
Weak expression, possibly in neuroblasts, after stripes fade.	No data for 3L.	No data for 3L.	0 out of 8 lines showed patterned eye.	u4739
Similar to u2309.	No data for 3L.	No data for 3L.	0 out of 1 lines showed patterned eye.	u3931
Weak expression, possibly in neuroblasts, after stripes fade.	No data for 3L.	No data for 3L.	0 out of 5 lines showed patterned eye. 1 out of 4 lines showed PSS.	u3125
Similar to u2309.	No data for 3L.	No data for 3L.	0 out of 11 lines showed patterned eye.	u3925
Weak expression, possibly in neuroblasts, after stripes fade.	No data for 3L.	No data for 3L.	0 out of 9 lines showed patterned eye.	u3118
Similar to u2309.	No data for 3L.	No data for 3L.	0 out of 9 lines showed patterned eye.	u3923
No consistent CNS expression.	No data for 3L.	No data for 3L.	0 out of 3 lines showed patterned eye.	u3123
Similar to u2309.	No data for 3L.	No data for 3L.	0 out of 6 lines showed patterned eye.	u3725
Similar to u2309.	No data for 3L.	No data for 3L.	0 out of 3 lines showed patterned eye.	u3525
Weak expression similar to u2309.	No data for 3L.	No data for 3L.	0 out of 12 lines showed patterned eye.	u3225
Weak expression, possibly in neuroblasts, after stripes fade. Similar to u1609.	No data for 3L.	No data for 3L.	0 out of 8 lines showed patterned eye.	u2316
Fig. 2. Weak expression, possibly in neuroblasts, after stripes fade. Similar to i1523.	No data for 3L.	No data for 3L.	0 out of 6 lines showed patterned eye.	u1609
Fig. 2. Weak expression, possibly in neuroblasts, after stripes fade. Similar to u2309.	No data for 3L.	No data for 3L.	1 out of 10 lines showed patterned eye.	u0900
Fig. 3. Expression, possibly in neuroblasts, after stripes fade. Similar to u1609.	No data for 3L.	No data for 3L.	0 out of 9 lines showed patterned eye. 1 out of 7 lines showed PSS.	i1523
Fig. 3. Expression in ventrolateral cell clusters.	Fig. S2. Confirmed in 1 line.	Expressed anterior to MF.	2 out of 15 lines showed patterned eye. 1 out of 10 lines showed PSS.	i2330
Similar to i2330.	Fig. S2. Confirmed in 2 lines.	Expressed anterior to MF.	3 out of 15 lines showed patterned eye. 1 out of 9 lines showed PSS.	i2339
No consistent CNS expression.	No data for 3L.	No data for 3L.	0 out of 8 lines showed patterned eye.	i3039
Fig. 3. and Fig. 6. Expression in glial cells.	No data for 3L.	No data for 3L.	0 out of 5 lines showed patterned eye.	i4053
No consistent CNS expression.	No data for 3L.	No data for 3L.	0 out of 13 lines showed patterned eye.	i4047
Similar to i4053, except expressed in fewer cells.	No data for 3L.	No data for 3L.	0 out of 4 lines showed patterned eye.	i4753
Fig. 6. Expression in glial cells.	No data for 3L.	No data for 3L.	0 out of 11 lines showed patterned eye.	d5776
No consistent CNS expression.	No data for 3L.	No data for 3L.	0 out of 14 lines showed patterned eye.	d6976

Fig. S2. Transgenic end point coordinates, sequences, and summary of expression. **Columns A and O:** names of constructs. *slp1-lacZ* and *slp2-lacZ* indicate which promoter region was used for *in vivo* analysis of the construct. **Columns B – C:** coordinates relative to either the *slp1* or *slp2* (bold and underlined) TSS. **Columns D – E:** restriction enzymes used to create end points (if applicable). **Columns F – G:** Flybase sequence coordinates. **Columns H – I:** 5' and 3' end point sequences. **Columns J – N:** Synopses of expression patterns in various tissues.
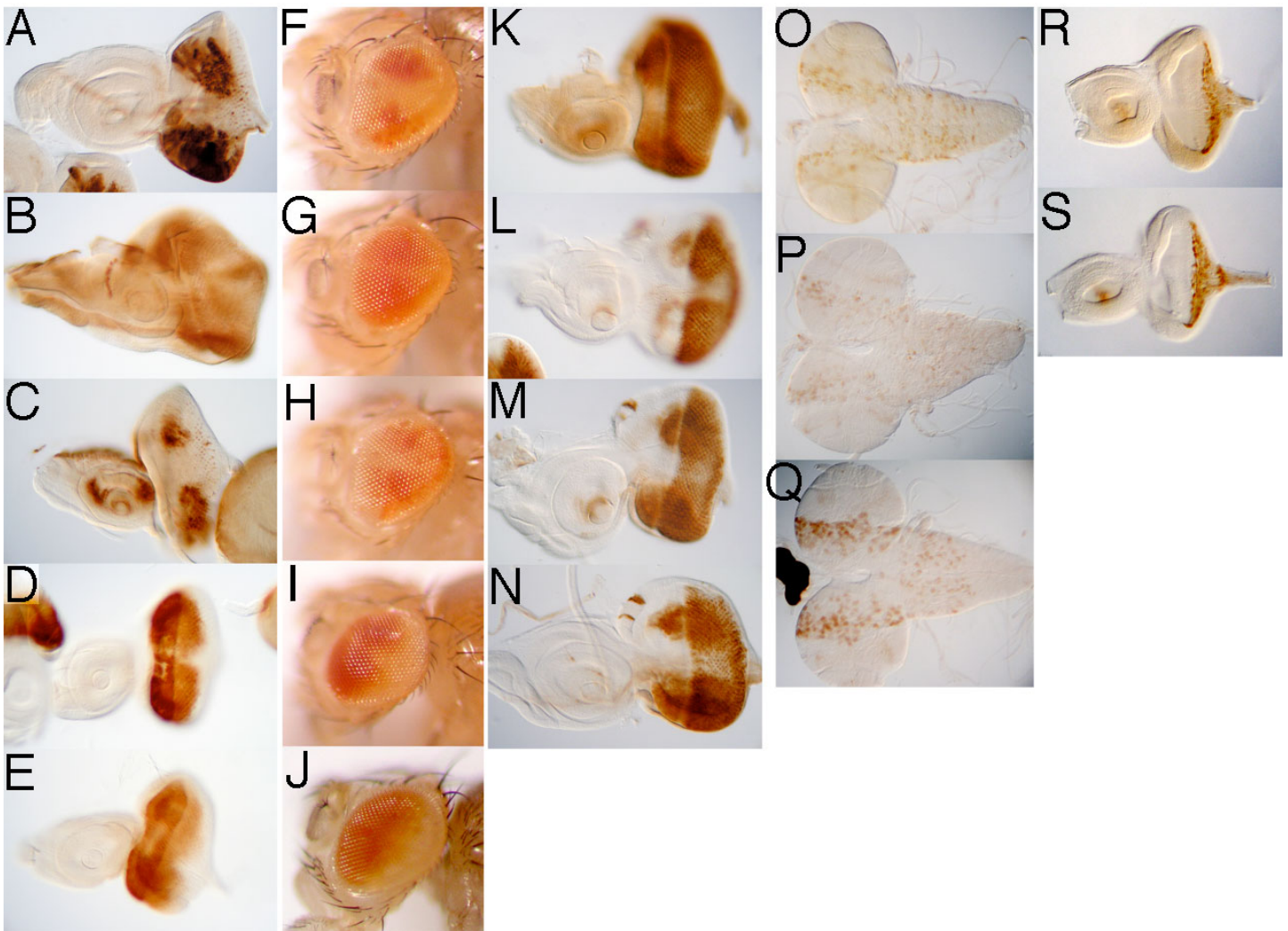
Fig. S3, Fujioka and Jaynes, 2012

Fig. S3. Eye disc and brain expression driven by *slp* CRMs.

A-E, K-N: βgal expression from *u8766-lacZ* in eye discs from different independent insertion lines. **F-J:** patterned eye color in each of the lines shown in A-E, respectively. Note the expression in dorsal and ventral sections of eye discs in A-E, which correlates roughly with the eye color patterns in F-J. These transgenes contain *mini-white*, which produces the eye color. Thus, u8766 may contain an eye disc enhancer that activates both *lacZ* and *mini-white* expression. In lines K-N, *lacZ* expression posterior to the morphogenetic furrow is relatively strong and uniform, and these lines do not have a patterned eye color (they have uniform eye colors ranging from dark orange to red, not shown). We note also that our transgenic vector contains Glass activator binding sites just upstream of the *mini-white* gene, to allow us to identify transformants with regulatory elements that tend to repress *mini-white*, such as Polycomb response elements, and the results might be influenced by these sites. **O-Q:** βgal expression in the 3rd instar larval CNS from *i1530-lacZ*, *i2330-lacZ*, and *i2339-lacZ*, respectively. Comparison with *slp* staining in Fig. 3 suggests that these CRMs may drive expression in cells that normally express *slp*, although we did not clearly detect *slp* expression in precisely these patterns. **R,S:** βgal expression of *i4060-lacZ*. The pattern suggests that these may be eye disc glial cells (Campbell et al., 1994; Xiong et al., 1994).
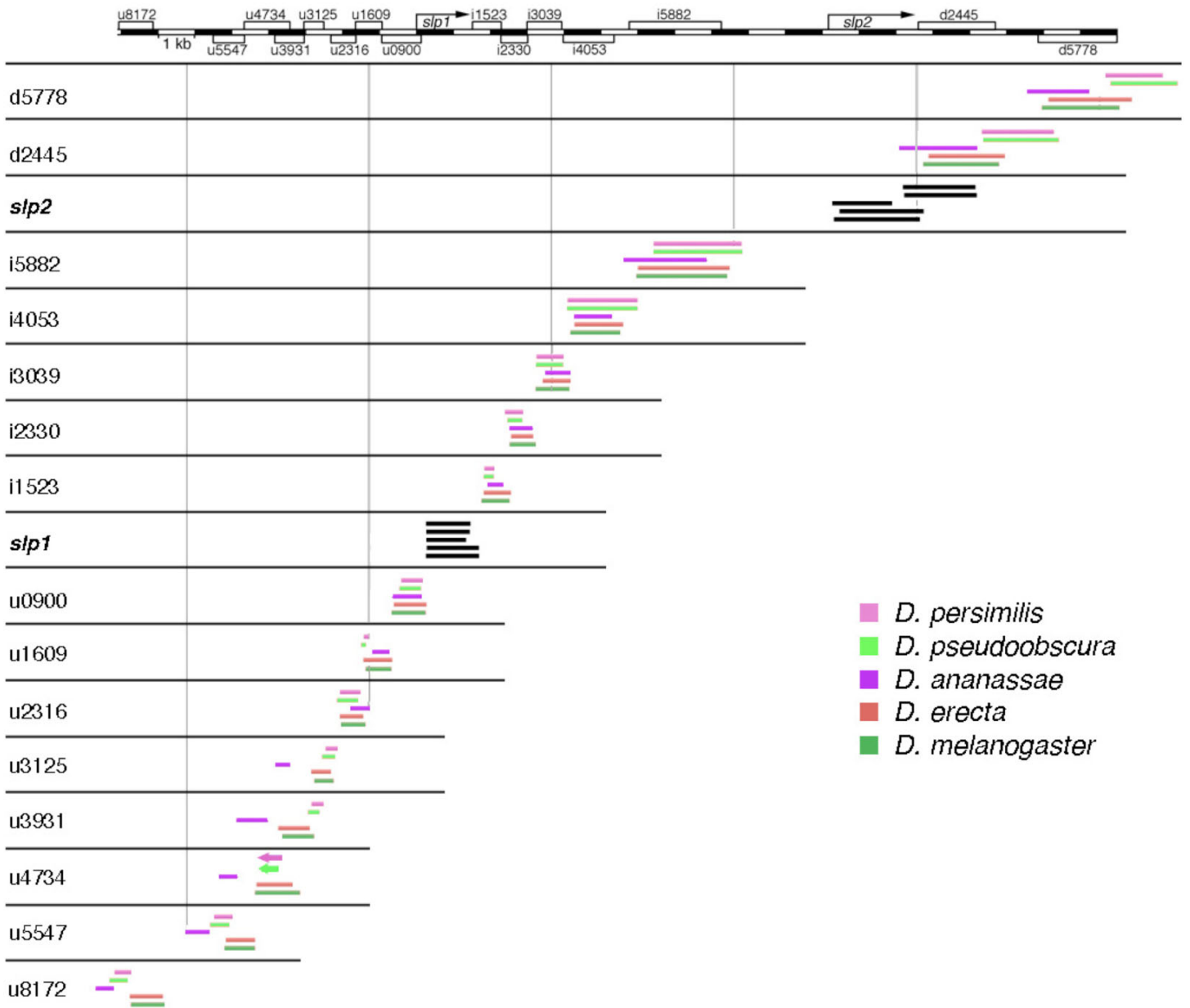
Fig. S4. Arrangements of sequences similar to *D. melanogaster slp* CRMs in 8 other drosophilids. BLAST searches (see Materials and Methods) were used to identify sequences similar to each *D. melanogaster slp* CRM shown on the left, in a representative subset of the 11 other available drosophilid genomes. Colored bars represent the extent of homologous sequences identified, relative to the beginning of the *slp1* coding region in each species. Black bars represent the *slp1* and *slp2* coding regions in the same species, in the order in which they are listed in the inset key. The length of each bar does not indicate the degree of homology, but only the distance over which homologous sequences were identified. That is, the ends of a long bar are defined by short similarities at those positions, rather than by extensive similarity between the ends. Sequence similarities identified in reverse orientation are indicated by arrows (or arrowheads for short sequences). **A:** Map of the locations of sequence similarities among the 5 most closely related species tested. Note that both the order and orientations of these similarities are the same in all these species, except for an inversion of u4734. The fact that this region is inverted in the melanogaster group of species, which includes *D. ananassae* and *D. erecta*, relative to all the other drosophilids (see also B) suggests that the inverted orientation was present in their common ancestor.

Fig. S4B, Fujioka and Jaynes, 2012
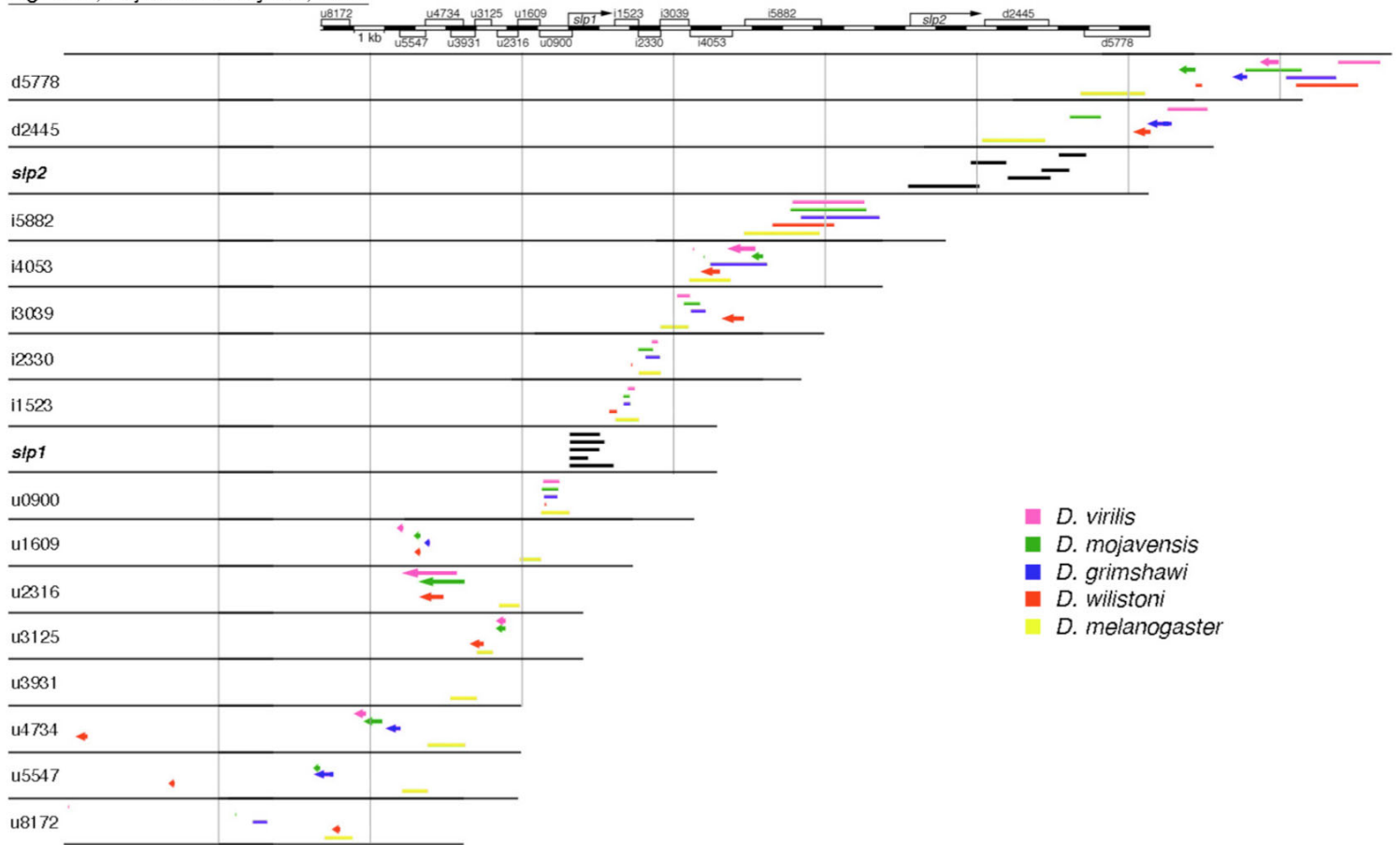
B: Map of the locations of sequence similarities between *D. melanogaster* and 4 drosophilid species more diverged from it. Note that the relative orientations of similarities suggests multiple inversion events involving either individual CRMs (or parts of them), or more than one of them (where both the orientation and relative position of adjacent elements are inverted) during drosophilid divergence. The details of these sequence similarities are available on request.

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