

2015

Quantification of CSF chemokines and cytokines allows for rapid laboratory detection of CNS infections and further discrimination between viral and non-viral pathogens

Danielle Fortuna, MD

Department of Surgery, Thomas Jefferson University


AM Cardenas

Children's Hospital of Philadelphia, Philadelphia, PA

EH Graf

Follow this and additional works at: <https://jdc.jefferson.edu/pacbresidentposters>

The Children's Hospital of Philadelphia

 Part of the [Medical Anatomy Commons](#), [Medical Cell Biology Commons](#), and the [Medical Pathology Commons](#)
Amity L. Roberts, PhD, D(ABMM)
Thomas Jefferson University

Let us know how access to this document benefits you

Larry A. Harshyne, PhD

Thomas Jefferson University

Recommended Citation

Fortuna, MD, Danielle; Cardenas, AM; Graf, EH; Roberts, PhD, D(ABMM), Amity L.; Harshyne, PhD,
See next page for additional authors

Larry A.; Hooper, PhD, D. Craig; and Curtis, MD, PhD, Mark T., "Quantification of CSF chemokines and cytokines allows for rapid laboratory detection of CNS infections and further discrimination between viral and non-viral pathogens" (2015). *Department of Pathology, Anatomy, and Cell Biology Resident's Posters*. Paper 20.

<https://jdc.jefferson.edu/pacbresidentposters/20>

This Article is brought to you for free and open access by the Jefferson Digital Commons. The Jefferson Digital Commons is a service of Thomas Jefferson University's [Center for Teaching and Learning \(CTL\)](#). The Commons is a showcase for Jefferson books and journals, peer-reviewed scholarly publications, unique historical collections from the University archives, and teaching tools. The Jefferson Digital Commons allows researchers and interested readers anywhere in the world to learn about and keep up to date with Jefferson scholarship. This article has been accepted for inclusion in Department of Pathology, Anatomy, and Cell Biology Resident's Posters by an authorized administrator of the Jefferson Digital Commons. For more information, please contact: JeffersonDigitalCommons@jefferson.edu.

Authors

Danielle Fortuna, MD; AM Cardenas; EH Graf; Amity L. Roberts, PhD, D(ABMM); Larry A. Harshyne, PhD; D. Craig Hooper, PhD; and Mark T. Curtis, MD, PhD

Quantification of CSF chemokines and cytokines allows for rapid laboratory detection of CNS infections and further discrimination between viral and non-viral pathogens

D Fortuna¹, AM Cardenas², EH Graf², A Roberts¹, L Harshyne¹, DC Hooper¹ and MT Curtis¹

¹Thomas Jefferson University Hospital, Philadelphia, PA, USA, ²Children's Hospital of Philadelphia, Philadelphia, PA, USA

ABSTRACT

Background: Prompt diagnosis of central nervous system (CNS) disease is critical to guide intervention and appropriate therapy. Development of novel laboratory approaches to rapidly classify acute-onset CNS disease is in great demand. Serious microbial pathogens, especially viruses, are quickly expanding beyond their historic geographic range and may not even be considered in the clinician's differential diagnosis. Unlike bacterial cultures, current viral testing targets a limited number of viruses. Additionally, despite diversity in etiology, signs and symptoms of both infectious and non-infectious CNS disorders can be remarkably similar, which can confuse the clinical picture and delay treatment. Bacterial, viral, fungal and protozoan CNS pathogens are sensed by pattern recognition receptors of the immune system, stimulating immediate release of measurable levels of chemokines and cytokines into the CSF. Our objective is to use pathogen-specific chemokine/cytokine profiles to classify CNS disease as infectious versus non-infectious and further discriminate between viral and non-viral infections.

Methods: Levels (pg/ml) of chemokines and cytokines were determined in the CSF of 45 patients with documented infectious meningitis or meningoencephalitis (mean age 19.2 years) and in the CSF of 25 patients who were negative for CNS infection (mean age 27.4 years). MILLIPLEX MAP Human Cytokine/Chemokine Magnetic Bead Panels (Millipore) were used to measure CSF chemokines and cytokines levels (pg/ml). Innate immune analytes quantified included IP-10 (CXCL10), IFN γ , IL-15, MDC (CCL22), MCP-1 (CCL2), Fractalkine, and FLT3L. Samples were analyzed in duplicate by a FlexMAP 3D (Luminex). Standard curves were generated for each cytokine and median fluorescent intensities were transformed into concentrations by 5-point, non-linear regression. For univariate analysis, comparisons between groups were made using the Mann-Whitney test. We utilized receiver operating characteristic (ROC) curve analysis to calculate areas under the ROC curve (AUC) for each analyte to access the utility of chemokine/cytokine levels as discriminating tests. The ROC generated sensitivity and specificity values were then used to determine clinically optimal cutoff values for the informative analytes.

Results: Univariate analysis utilizing Mann-Whitney tests demonstrated that median values (pg/ml) of IP-10 (CXCL10), IFN γ , IL-15, MDC (CCL22), MCP-1 (CCL2), Fractalkine, and FLT3L were all significantly higher in CSF from patients with infectious brain disorders than in CSF from patients with non-infectious disorders (p-value < 0.05). MDC (CCL22) demonstrated statistical significance, when comparing viral infections versus non-viral infections (with the non-viral infection group having higher analyte levels). IP10 (CXCL10) can reliably distinguish between an infectious versus non-infectious CNS process (AUC 0.9778) with an optimal cut-off value of 2023 pg/ml (sensitivity, specificity; 93.0%, 92.0%). In the infectious group, MDC (CCL22) can reliably differentiate between viral and non-viral CNS infection (AUC 0.9545) with an optimal cut-off value of 194 pg/ml (sensitivity, specificity; 91.67%, 87.88%).

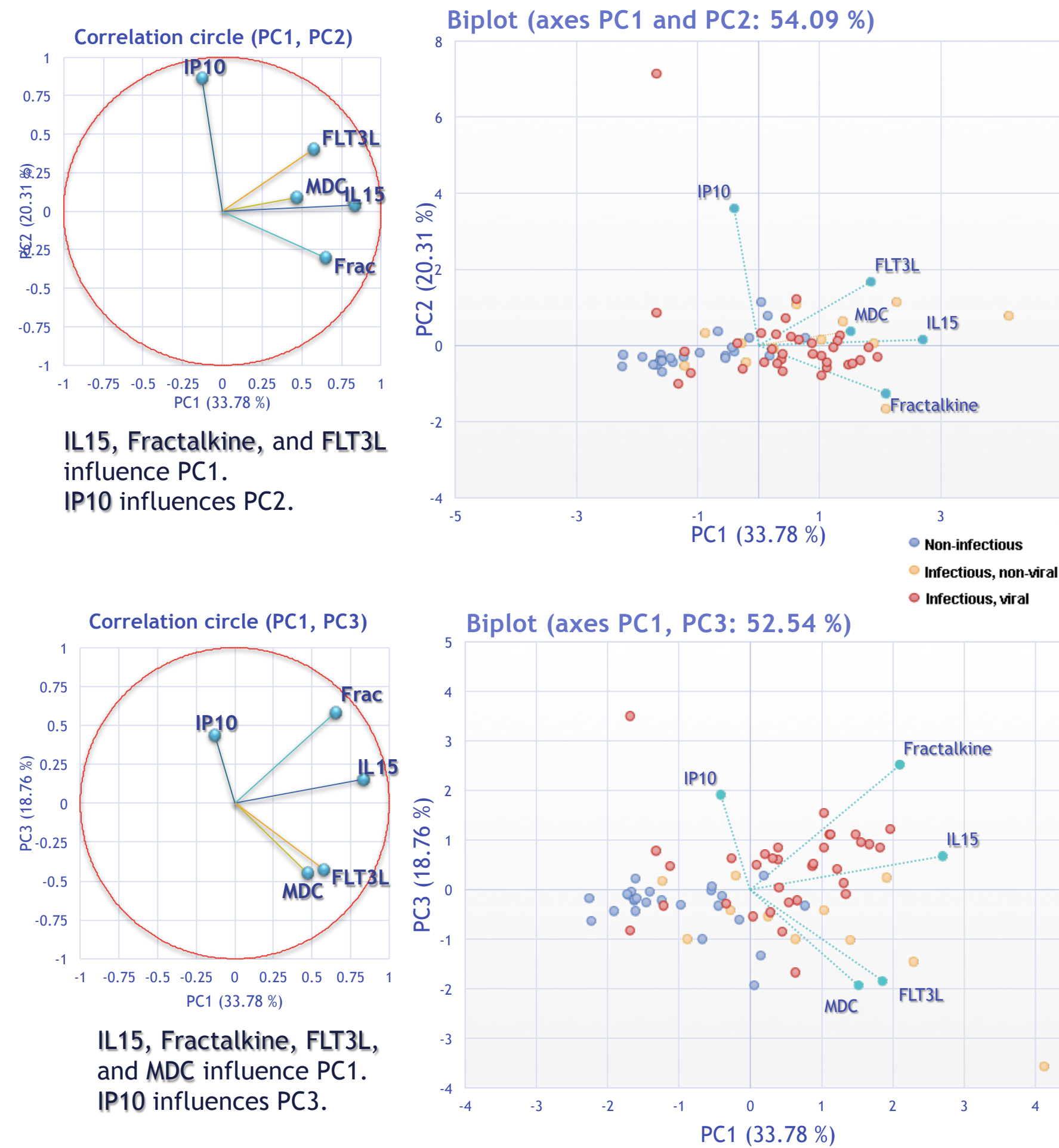
Conclusion: CSF levels (pg/ml) of IP-10 (CXCL10) can reliably distinguish infectious versus non-infectious CNS disorders, and in the infectious group, MDC (CCL22) can reliably distinguish between viral and non-viral CNS infections. These results suggest that CSF chemokine/cytokine quantification can serve as a useful laboratory tool for the rapid triage of CNS diseases to help guide prompt therapy and further testing.

BACKGROUND INFORMATION

CYTOKINE	FUNCTION IN THE INFLAMMATORY RESPONSE	NON-INFECTIOUS CASES	INFECTIOUS CASES
MCP1/CCL2	marker of non-specific inflammation	HEADACHE, IDIOPATHIC INTRACRANIAL HYPERTENSION (IIH), SUBARACHNOID HEMORRHAGE, POSSIBLE AUTOIMMUNE DISEASE	VIRAL: ENTEROVIRUS, HUMAN PARECHOVIRUS, WEST NILE VIRUS, JC VIRUS, HHV6
IP10/CXCL10	produced by a wide range of CNS cells in response to microbial pathogens; stimulated by multiple pathways		
IL15	growth/survival factors for both NK cells and cytotoxic CD8+ T-cells		
MDC/CCL22	produced in response to various microbial products; down-regulated by IFN γ		
FLT3L	supports maturation of antigen presenting cells		NON-VIRAL: S.EPIDERMIDIS, S.PNEUMONIAE, M.TUBERCULOSIS, B.BURGDORFERI, TOXOPLASMOSIS, CRYPTOCOCCUS NEOFORMANS
FRACTALKINE/CX3CL1	support monocyte adhesion to endothelium		
IFN γ	major chemokine product of NK cells; stimulates phagocytosis and pathogen killing in macrophages		

RESULTS: PRINCIPAL COMPONENT ANALYSIS (PCA)

Principal Component Analysis (PCA) with the variables IP10, IL15, MDC, FLT3L, and Fractalkine



We used PCA to visually represent the underlying structure of our data and examine the variability of specific cytokines/chemokines to help distinguish among CNS disease states (non-infectious and infectious, as well as viral versus non-viral infections).

RESULTS: MANN-WHITNEY TEST OF SIGNIFICANCE

	NON-INFECTIOUS VS INFECTIOUS	VIRAL VS NON-VIRAL
MCP1	* (p=0.0494)	
IFN γ	** (p=0.0035)	
IP10	**** (p<0.0001)	NS
IL15	**** (p<0.0001)	NS
MDC	** (p=0.0028)	**** (p<0.0001)
FLT3L	** (p=0.0057)	NS
Fractalkine	**** (p<0.0001)	NS

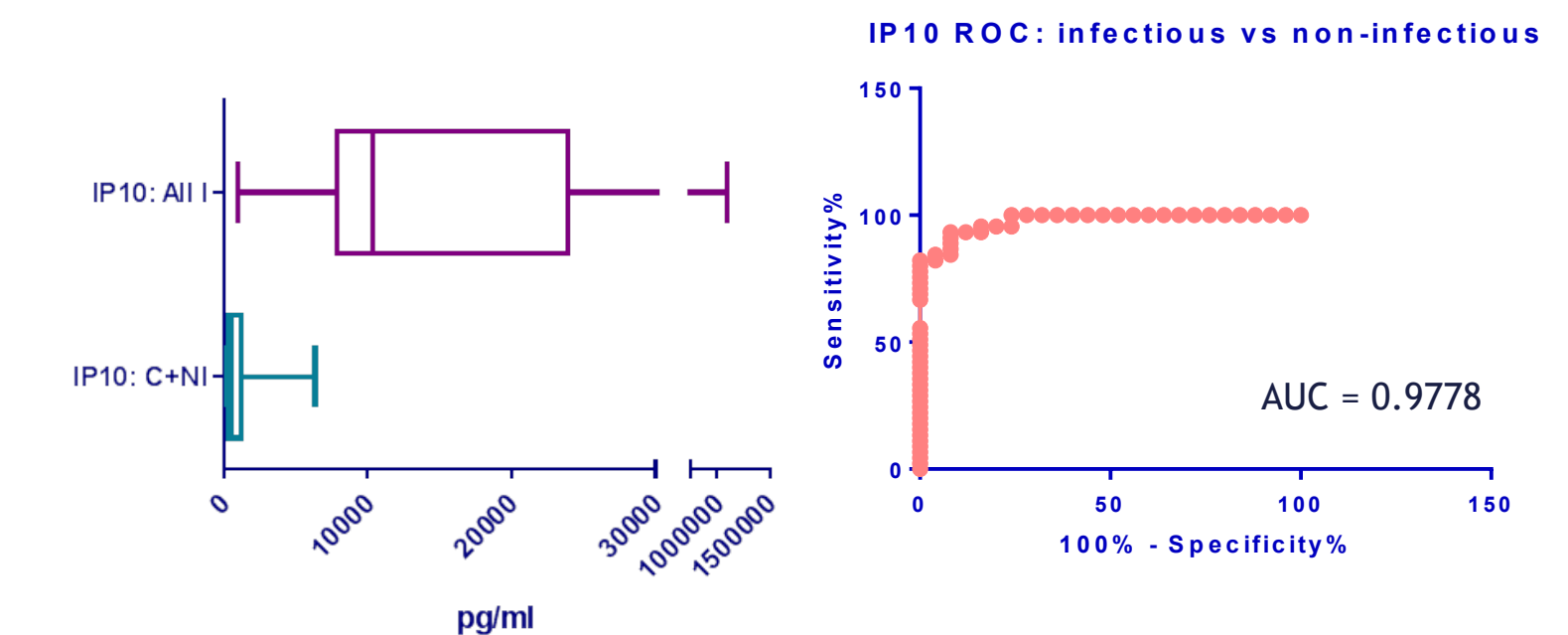
* indicates significant difference between two groups. Number of * represents p-value summary. "NS" indicates no significant difference.

Mann-Whitney tests of significance for non-parametric data

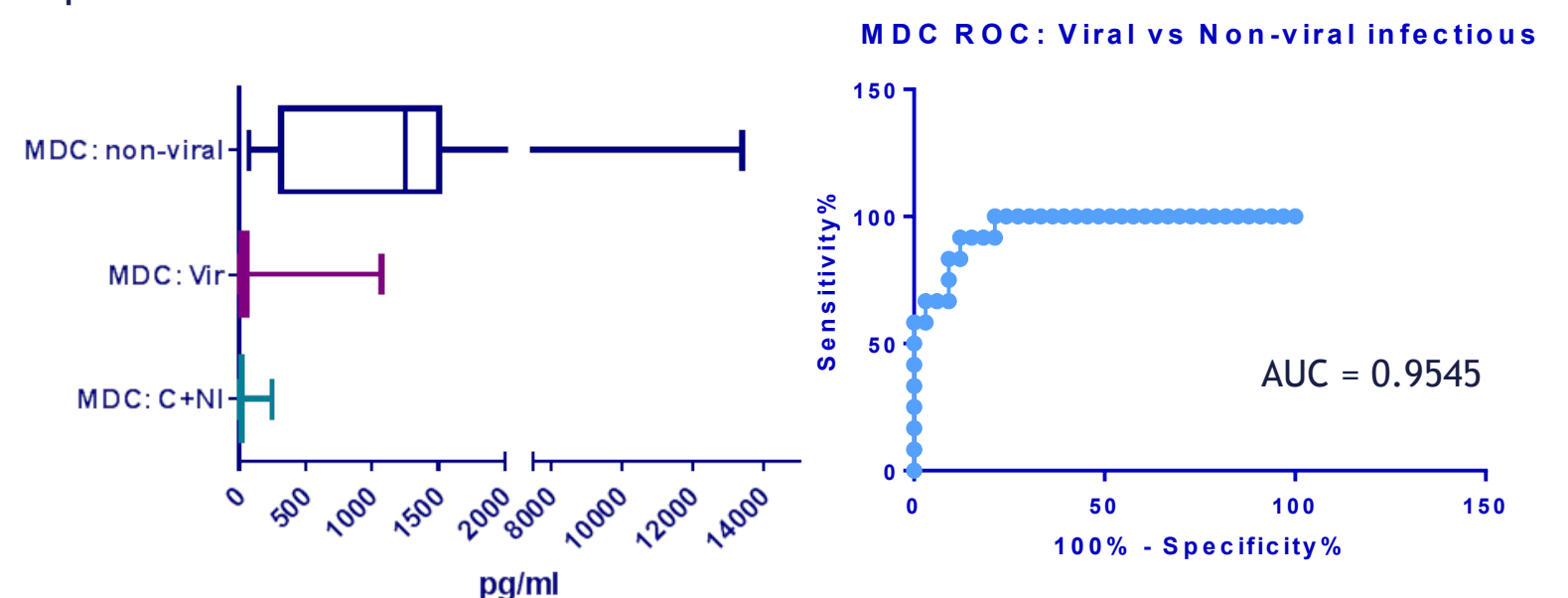
Levels of all studied cytokines (MCP1, IFN γ , IP10, IL15, MDC, FLT3L, and Fractalkine) were **significantly higher in the infectious** compared to the non-infectious group.

RESULTS: DESCRIPTIVE STATISTICS AND RECEIVER OPERATOR CURVES (ROC)

IP10/CXCL10 can reliably distinguish between an **infectious (All I) versus non-infectious (C+NI)** CNS process (AUC 0.9778) with an optimal cut-off value of 2023 pg/ml (sensitivity, specificity; 93.0%, 92.0%).



Among the infectious cases, **MDC** distinguishes **viral from non-viral** infectious-type processes.



CASE STUDY

16 year old woman with history of lupus presented with CNS symptoms (ataxia with progression to altered mental status and paralysis).

CSF studies were normal. CSF microbiology tests were negative.

MCP1: 5678.88 pg/ml
IP10: 23247.43 pg/ml
MDC: 46.99 pg/ml
FLT3L: 69.63 pg/ml
Fractalkine: 100.63 pg/ml

The cytokine profile supports infectious, viral-type etiology.

Four weeks following admission, admission CSF was positive for human parechovirus (HPeV).

MDC LEVEL	SENSITIVITY	SPECIFICITY	LIKELIHOOD RATIO
> 194.0	91.67	87.88	7.563
> 238.3	83.33	87.88	6.875
> 267.3	83.33	90.91	9.167
> 355.9	75.00	90.91	8.250
> 487.1	66.67	90.91	7.333
> 556.6	66.67	93.94	11.00
> 632.1	66.67	96.97	22.00
> 884.2	58.33	96.97	19.25

CONCLUSIONS

- CSF levels (pg/ml) of IP-10/CXCL10 can reliably distinguish **infectious versus non-infectious** CNS disorders
- In the infectious group, **MDC/CCL22** can reliably distinguish between **viral and non-viral** CNS infections.
- CSF chemokine/cytokine quantification can serve as a useful laboratory tool for the rapid triage of CNS diseases to help guide prompt therapy and further testing.