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Unaltered myocilin expression in the blood of primary open angle glaucoma patients

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Purpose: To investigate the expression of the myocilin gene (MYOC) in the blood of primary open angle glaucoma (POAG) patients to determine if altered systemic expression is playing a role.

Methods: Patients (n=47) were eligible for inclusion if they met standard clinical criteria for POAG. Control subjects (n=27) were recruited who were free from glaucoma by examination. RNA was extracted from leukocytes of patients and controls and converted to cDNA by reverse transcriptase enzyme, and quantitative PCR was used to assess expression levels of MYOC and the house keeping gene β-globulin (HBB). The ratio of MYOC expression to HBB expression for POAG patients was compared to that of controls and to clinical characteristics of POAG patients.

Results: Mean gene expression values were statistically similar in POAG patients and controls for both MYOC (p≤0.55) and HBB (p≤0.48). MYOC/HBB ratios were also statistically indistinguishable between POAG patients and controls (p≤0.90). MYOC/HBB ratios were not significantly associated with age, sex, or ethnicity of patients within the POAG group. Similarly, MYOC/HBB ratios were not significantly associated with clinical parameters related to POAG severity, including maximum intraocular pressure, vertical cup-to-disk ratio, static perimetry mean deviation, or static perimeter pattern standard deviation.

Conclusions: MYOC expression is not altered in the blood of POAG patients, unlike MYOC expression in trabecular meshwork (TM) cultures. These results suggest that MYOC expression is not altered systemically but rather that MYOC expression may contribute to POAG pathogenesis in specific tissues such as TM.

Glaucma is one of the leading causes of blindness worldwide [1,2], characterized by chronic degeneration of axons in the optic nerve head. Primary open angle glaucoma (POAG) is the most prevalent type of glaucoma in western countries and has risk factors that include elevated intraocular pressure (IOP) and age [3]. Elevated IOP is associated with increased aqueous humor outflow resistance in the trabecular meshwork (TM) of the eye [2], although the exact mechanism and causative factors for this increase is unclear. Up to half of all patients with POAG have a positive family history [4,5], and these and other observations suggest that genetic factors may contribute to POAG [1,6,7].

Myocilin (MYOC) was the first gene linked to POAG [8] and is the one most studied [9]. It is located in chromosome 1, contains three exons, and codes for a largely extracellular matrix protein. This protein has an NH2-terminal coiled region and a COOH-terminal olfactomedin domain [10], but its function is still not well understood. To date, mutations in MYOC seem most likely to have their pathogenic effect largely because of inability of the protein to fold properly [11]. This may result in an unfolded protein response in TM cells, activating a mitochondria-independent apoptosis pathway which ultimately leads to cell death, breakdown of TM cell structure, obstruction of aqueous humor outflow pathway, ocular hypertension, and ultimately the optic nerve damage of glaucoma [12-14]. MYOC may directly impair optic nerve function when mutated [15]; however, direct evidence for this hypothesis is still lacking [11,15].

Most studies investigating MYOC expression in POAG have employed human cultured TM cells [16,17]. However, whole blood gene expression studies have been used to investigate POAG [18], other hereditary optic neuropathies [19], and diseases affecting brain anatomy and function [20, 21] because the target tissues for these diseases is not readily available. Therefore, the current study investigated MYOC expression in whole blood from POAG patients in hope that this approach will add to our knowledge of whether altered systemic expression of this gene contributes to POAG pathogenesis.

METHODS

Patients and controls: Patients (n=47) were evaluated in the Glaucoma Service at the Wills Eye Institute, Philadelphia, PA, and enrolled after examination by a glaucoma specialist.
Patients were eligible for inclusion if they met the following clinical criteria for POAG [22-25]: age greater than 40 years; intraocular pressure (IOP) ≥21 mmHg in one or both eyes before initiation of glaucoma treatment; normal-appearing, open anterior chamber angles bilaterally by gonioscopy; optic nerve appearance characteristic of the optic discs typically observed in primary open-angle glaucoma (with localized narrowing or absence of the neuro-retinal rim, with the amount of cupping exceeding the amount of pallor of the rim, and with asymmetric cupping of the optic discs in the two eyes); and static visual field (using a full threshold 24–2 program; Humphrey Field Analyzer II; Carl Zeiss Meditec, Inc., Dublin, CA) showing abnormalities typical of glaucoma (as per Advanced Glaucoma Intervention Study criteria) [26]. Good agreement was required between the appearance of the optic disc and the visual field. Exclusion criteria included historical, neuroimaging, or biochemical evidence of another possible optic neuropathic process affecting either eye, significant visual loss in both eyes not associated with glaucoma, or choosing not to participate. This research adhered to the tenets of the Declaration of Helsinki, and all patients and controls signed an informed consent approved by the Wills Eye Institute institutional review board.

All control subjects (n=27), frequently spouses of patients, had full ophthalmologic examinations documenting IOPs that were <21 mmHg and symmetric in the two eyes, normal anterior chambers, optic discs that were normal and IOPs that were <21 mmHg and symmetric in the two eyes, patients, had full ophthalmologic examinations documenting the Wills Eye Institute institutional review board.

Quantitative RT–PCR: A two-step semi-quantitative reverse transcriptase polymerase chain reaction (RT–PCR) method was used to measure gene expression levels of MYOC and β-globulin (HBB) in POAG patients and controls. Random hexamers were used as primers in the first step of cDNA synthesis. Total RNA (1 μg) was combined with 0.5 μg primers, 200 μM dNTPs, and sterile Milli-Q water (Millipore, Billerica, MA) and preheated at 65 °C for 2 min to denature secondary structures. The mixture was then cooled rapidly to 20 °C and then 10 μl 5× RT Buffer, 10 mM dithiothreitol, and 200 U Moloney Murine Leukemia Virus Reverse Transcriptase (Invitrogen Life Sciences, Grand Island, NY) were added for a total volume of 50 μl. The RT mix was incubated at 37 °C for 90 min then stopped by heating at 95 °C for 5 min. The cDNA stock was stored at −20 °C.

Relative RT–PCR was performed to measure gene expression of MYOC and HBB according to standard guidelines [27]. Primer sequences and optimal PCR annealing temperatures (ta) are listed in Table 2. Primer sequences were designed to span exon regions to insure that no false positive PCR fragments would be generated from pseudogenes and contaminate genomic DNA. In addition, all forward PCR primers were labeled with fluorescein (6-FAM), making quantitation more accurate. Polymerase chain reactions were performed using 100 ng of cDNA, 5 pmoles of each oligonucleotide primer, 200 μM of each dNTP, 1 unit of HotStar Taq-polymerase (Qiagen, Valencia, CA) and 1× PCR buffer in a 20 μl volume. The PCR program initially started with a 95 °C denaturation for 5 min, followed by 25 cycles of 95 °C for 1 min, 55 °C for 45 s, and 72 °C for 1 min. Linear amplification range for each gene was tested on the adjusted cDNA, and 25 cycles were found to be optimal for both

### Table 1. Primer sequences, PCR annealing temperature and amplicon size for the MYOC gene.

<table>
<thead>
<tr>
<th>Exon</th>
<th>Primer sequence</th>
<th>Annealing temp (°C)</th>
<th>Amplicon size (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pro-F</td>
<td>TGTAAAAAGCACGGCCAGTCGGCGCTTGTTATTTTCTCTGTT</td>
<td>55</td>
<td>592</td>
</tr>
<tr>
<td>Pro-R</td>
<td>CAGGAAACCAGCTATGACC</td>
<td>77</td>
<td>775</td>
</tr>
<tr>
<td>1F</td>
<td>TGTAAAAAGCACGGCCAGTCGGCGCTTGTTATTTTCTCTGTT</td>
<td>57</td>
<td>325</td>
</tr>
<tr>
<td>1R</td>
<td>CAGGAAACCAGCTATGACC</td>
<td>850</td>
<td>850</td>
</tr>
<tr>
<td>2F</td>
<td>TGTAAAAAGCACGGCCAGTCGGCGCTTGTTATTTTCTCTGTT</td>
<td>325</td>
<td>325</td>
</tr>
<tr>
<td>2R</td>
<td>CAGGAAACCAGCTATGACC</td>
<td>592</td>
<td>592</td>
</tr>
<tr>
<td>3AF</td>
<td>TGTAAAAAGCACGGCCAGTCGGCGCTTGTTATTTTCTCTGTT</td>
<td>57</td>
<td>700</td>
</tr>
<tr>
<td>3BR</td>
<td>CAGGAAACCAGCTATGACC</td>
<td>775</td>
<td>775</td>
</tr>
<tr>
<td>3BF</td>
<td>TGTAAAAAGCACGGCCAGTCGGCGCTTGTTATTTTCTCTGTT</td>
<td>700</td>
<td>700</td>
</tr>
<tr>
<td>3AR</td>
<td>CAGGAAACCAGCTATGACC</td>
<td>57</td>
<td>57</td>
</tr>
</tbody>
</table>

F=forward; R=reverse; Pro – promoter. Bold and underlined sequences are those of the M13.
**TABLE 2. PRIMER SEQUENCES AND ANNEALING TEMPERATURE β-GLOBULIN AND MYOCILIN FLUORESCENT LABELED PRIMERS.**

<table>
<thead>
<tr>
<th>Primer name</th>
<th>Primer sequence</th>
<th>Annealing temp (°C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>β-globulin-F</td>
<td>(6-FAM)AGCCTGCTTTTGGCGA</td>
<td>57</td>
</tr>
<tr>
<td>β-globulin-R</td>
<td>CTGTCGCTGGGCG</td>
<td></td>
</tr>
<tr>
<td>MYOC-LAB-F</td>
<td>(6-FAM)TTTCTACGTGGAATTTGGACA</td>
<td>59</td>
</tr>
<tr>
<td>MYOC-R</td>
<td>GTAGGGCCTGGGGTCT</td>
<td></td>
</tr>
</tbody>
</table>

F=forward; R=reverse. The forward primers were labeled with 6-FAM.

**TABLE 3. MYOCILIN GENE EXPRESSION IN POAG PATIENTS AND CONTROLS.**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Number POAG/control</th>
<th>POAG</th>
<th>Control</th>
<th>p≤</th>
</tr>
</thead>
<tbody>
<tr>
<td>MYOC expression; mean (SD)</td>
<td>47:26</td>
<td>7476 (4325)</td>
<td>6839 (4271)</td>
<td>0.55</td>
</tr>
<tr>
<td>β-globulin expression; mean (SD)</td>
<td>44:24</td>
<td>109533 (31355)</td>
<td>103885 (31047)</td>
<td>0.48</td>
</tr>
<tr>
<td>MYOC/β-globulin; mean (SD)</td>
<td>42:23</td>
<td>0.0685 (0.0520)</td>
<td>0.0667 (0.0597)</td>
<td>0.90</td>
</tr>
<tr>
<td>MYOC expression in Caucasians; mean (SD)</td>
<td>26:22</td>
<td>7132 (4526)</td>
<td>7268 (4121)</td>
<td>0.90</td>
</tr>
<tr>
<td>β-globulin expression in Caucasians; mean (SD)</td>
<td>22:20</td>
<td>104941 (31552)</td>
<td>99392 (32171)</td>
<td>0.58</td>
</tr>
<tr>
<td>MYOC/β-globulin in Caucasians; mean (SD)</td>
<td>21:19</td>
<td>0.0622 (0.0487)</td>
<td>0.0730 (0.0619)</td>
<td>0.54</td>
</tr>
</tbody>
</table>

POAG=primary open angle glaucoma; MYOC=myocilin gene expression; SD=standard deviation.

MYOC and HBB. The PCR samples were electrophoresed on the 3130xl Genetic Analyzer (Applied Biosystems).

Statistical analysis: Absolute RT–PCR values were used to calculate a ratio of the MYOC peak area in the selected linear amplification cycle divided by that of HBB, creating an MYOC/HBB ratio. All clinical and genetic data were analyzed using SPSS v17 (IBM, Chicago, IL).

**RESULTS**

Age (POAG patients 67.3 years; controls 63.6 years; p≤0.17) and sex (POAG 26 males/21 females; controls 12/15; p≤0.18) of the 47 unrelated POAG patients were similar to the 27 control individuals, but ethnicity differed between the POAG group (25 Caucasian/22 African American) and the control group (23 Caucasian/4 African American; p≤0.003).

After aligning and reading all sequences neither POAG patients nor controls were found to have any significant mutation or polymorphism in the coding or promoter regions of MYOC.

Mean gene expression values for both MYOC and HBB (p≤0.48) were statistically similar in POAG patients and controls (Table 3). MYOC/HBB ratios (p≤0.90) were also indistinguishable between POAG patients and controls. Because of ethnic differences between the POAG group and controls, gene expression values and ratios were also compared between Caucasian POAG patients and Caucasian controls. Mean MYOC (p≤0.90) gene expression and MYOC/ HBB (p≤0.54) ratios also did not differ between these groups.

**DISCUSSION**

The 47 patients reported here met rigorous clinical criteria for POAG [22-25] with elevated IOP, normal anterior chamber, and evidence on funduscopy of typical glaucomatous optic nerve damage. They did not have evidence of other types of glaucoma or alternative causes of optic nerve injury by clinical criteria, and none had dysmorphism or an obvious genetic syndrome. They were compared to 27 control individuals in whom POAG and other evidence of optic nerve damage were carefully excluded.

Screening the full MYOC gene and its promoter region revealed no mutations or significant polymorphisms in POAG patients or controls. These results are not surprising, since the prevalence of MYOC mutations is generally less than 5% in adult POAG populations [28]. Currently, there are 85 glaucoma causing mutations listed in the comprehensive myocilin database. They were classified as a glaucoma causing mutations based on the following criteria: i) predicted disruption of protein translation (e.g., frame-shift mutations and premature stop codons); ii) sequence variant frequency in control (unaffected) populations (those with a frequency >1% were classified as polymorphisms); iii) variant location (i.e., protein homology domain; cross species conservation of coding sequence); iv) evidence for partial segregation with the phenotype within a family and v) results of solubility studies. Interestingly, several sequence changes have been reported in the MYOC promoter region, but they were defined as neutral polymorphisms based on the pathologic characteristics
**Table 4. Correlation between clinical parameters and MYOC/β-globulin ratios.**

<table>
<thead>
<tr>
<th>Clinical parameter</th>
<th>MYOC/β-globulin</th>
<th>p≤</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age in years</td>
<td>0.204</td>
<td>0.21</td>
</tr>
<tr>
<td>Sex</td>
<td>0.040</td>
<td>0.81</td>
</tr>
<tr>
<td>Ethnicity</td>
<td>0.122</td>
<td>0.45</td>
</tr>
<tr>
<td>Visual acuity OD</td>
<td>0.016</td>
<td>0.92</td>
</tr>
<tr>
<td>Visual acuity OS</td>
<td>0.075</td>
<td>0.64</td>
</tr>
<tr>
<td>Maximum IOP OD</td>
<td>0.114</td>
<td>0.48</td>
</tr>
<tr>
<td>Maximum IOP OS</td>
<td>0.237</td>
<td>0.14</td>
</tr>
<tr>
<td>Vertical c/d ratio OD</td>
<td>0.129</td>
<td>0.43</td>
</tr>
<tr>
<td>Vertical c/d ratio OS</td>
<td>0.171</td>
<td>0.29</td>
</tr>
<tr>
<td>MD OD</td>
<td>−0.042</td>
<td>0.80</td>
</tr>
<tr>
<td>MD OS</td>
<td>−0.025</td>
<td>0.88</td>
</tr>
<tr>
<td>PSD OD</td>
<td>0.211</td>
<td>0.19</td>
</tr>
<tr>
<td>PSD OS</td>
<td>0.024</td>
<td>0.89</td>
</tr>
</tbody>
</table>

MYOC/β-globulin column contains correlation coefficients; OD=right eye; OS=left eye; IOP=intraocular pressure; c/d=cup to disk; MD=Humphrey visual field mean deviation; PSD=Humphrey visual field pattern standard deviation.

Described above (myocilin). This may indicate that the promoter region is free of mutations which could alter its expression. Alternatively, assessing potential pathogenicity for promoter region sequence changes can be challenging and may not follow the same pathological criteria as sequence changes found in coding regions [29].

Expression of MYOC in blood of POAG patients was unchanged compared to that of controls (Table 3). Expression was statistically similar to that of the housekeeping gene HBB, and the normalized expression of MYOC (MYOC/HBB) also did not differ between patients and controls. MYOC expression did not differ between Caucasian and African American POAG patients and ethnicity matched controls. Finally, MYOC/HBB did not correlate with age or with various clinical factors associated with POAG such as visual acuity, IOP, and C/D ratio (Table 4).

The lack of significant change in MYOC expression in blood of POAG patients stands in contrast to previous studies documenting decreased expression of MYOC in cultured human TM cells [16]. Gene expression studies in whole blood are clearly capable of documenting changes pertinent to complex neurologic diseases such as autism [30], amyotrophic lateral sclerosis [31], schizophrenia [32], and other psychoses [33]. Patients with Leber hereditary optic neuropathy, another lateral sclerosis [31], schizophrenia [32], and other psychoses [33], as well as patients with Leber hereditary optic neuropathy, another complex neurologic disease such as autism [30], and various clinical parameters within the POAG group may be subject to type II statistical errors. The population studied was predominantly Caucasian and African-American, and different results might be obtained in other ethnicities.

A potential limitation of this study is that the number of individuals studied was relatively small, bringing up the possibility that the lack of a statistical difference in MYOC expression and MYOC/HBB ratio between POAG patients and controls might be due to a type II statistical error because of inadequate power. This same patient group was adequate to confirm statistically significant differences in optic atrophy type 1 (OPA1) expression and the OPA1/HBB ratio between POAG patients and controls [18], but it is possible that differences in MYOC expression between POAG patients and controls are smaller, although still present. Similarly, the lack of correlation between the MYOC/HBB ratio and various clinical parameters within the POAG group may be subject to type II statistical errors. The population studied was predominantly Caucasian and African-American, and different results might be obtained in other ethnicities.

We found that systemic MYOC expression was unchanged in these POAG patients compared to controls. One interpretation of these results is that the MYOC protein plays a particularly important role in the globe and that regulation of MYOC expression that might be pertinent to POAG, congenital glaucoma, and/or steroid-induced glaucoma is relatively specific to the TM and may not be reflected in a significant extent in bone marrow or other non-ocular tissues. It is also possible that POAG is not altered by wild-type MYOC expression in any tissue [37]. A gain-of-function disease model was suggested after identification of mutant, misfolded forms of the MYOC protein were found aggregated in the endoplasmic reticulum of TM cells [38]. TM cells are essential for homeostatic regulation of aqueous humor, and their disruption may cause elevated intraocular pressure. A mutation-dependent, gain-of-function association between human MYOC and the peroxisomal targeting signal type 1 receptor (PTS1R) led to the [39] hypothesis that specific MYOC mutations may cause different amounts of MYOC mutations in the TM.
misfolding, with corresponding varying degrees of recognition by the ubiquitin degradation pathway. A greater opportunity for mutant MYOC to interact with PTS1R may allow for poorer clearance from the TM endoplasmic reticulum and greater trabecular cell dysfunction, culminating in a higher IOP phenotype [39].

REFERENCES


