

Division of Rheumatology Faculty Papers

Division of Rheumatology

1-15-2024

Associations of Galectin-3 Levels With Measures of Vascular Disease in Patients With Rheumatoid Arthritis

Amanda Nussdorf *Thomas Jefferson University*, amanda.nussdorf@jefferson.edu

Elizabeth Park

Isabelle Amigues

Laura Geraldino-Pardilla

Sabahat Bokhari

See next page for additional authors

Follow this and additional works at: https://jdc.jefferson.edu/rheumatologyfp

Part of the Diseases Commons, and the Rheumatology Commons
<u>Let us know how access to this document benefits you</u>

Recommended Citation

Nussdorf, Amanda; Park, Elizabeth; Amigues, Isabelle; Geraldino-Pardilla, Laura; Bokhari, Sabahat; Giles, Jon T.; and Bathon, Joan M., "Associations of Galectin-3 Levels With Measures of Vascular Disease in Patients With Rheumatoid Arthritis" (2024). *Division of Rheumatology Faculty Papers*. Paper 1. https://jdc.jefferson.edu/rheumatologyfp/1

This Article is brought to you for free and open access by the Jefferson Digital Commons. The Jefferson Digital Commons is a service of Thomas Jefferson University's Center for Teaching and Learning (CTL). The Commons is a showcase for Jefferson books and journals, peer-reviewed scholarly publications, unique historical collections from the University archives, and teaching tools. The Jefferson Digital Commons allows researchers and interested readers anywhere in the world to learn about and keep up to date with Jefferson scholarship. This article has been accepted for inclusion in Division of Rheumatology Faculty Papers by an authorized administrator of the Jefferson Digital Commons. For more information, please contact: JeffersonDigitalCommons@jefferson.edu.

Authors

Amanda Nussdorf, Elizabeth Park, Isabelle Amigues, Laura Geraldino-Pardilla, Sabahat Bokhari, Jon T. Giles, and Joan M. Bathon

This article is available at Jefferson Digital Commons: https://jdc.jefferson.edu/rheumatologyfp/1



Contents lists available at ScienceDirect

Seminars in Arthritis and Rheumatism

journal homepage: www.elsevier.com/locate/semarthrit



Associations of galectin-3 levels with measures of vascular disease in patients with rheumatoid arthritis

Amanda Nussdorf^a, Elizabeth Park^{c,*}, Isabelle Amigues^b, Laura Geraldino-Pardilla^c, Sabahat Bokhari^d, Jon T Giles^{c,e}, Joan M Bathon^c

^a Division of Rheumatology, Department of Medicine, Thomas Jefferson University Hospital, Philadelphia, PA, USA

^b Division of Rheumatology, Department of Medicine, National Jewish Health, Denver, CO, USA

^c Division of Rheumatology, Department of Medicine, Columbia University Irving Medical Center/New York Presbyterian Hospital, New York, NY, USA

^d Division of Cardiology, Lehigh Valley Heart and Vascular Institute, Allentown, PA, USA

e Division of Rheumatology, Department of Medicine, Cedars-Sinai Medical Center, Los Angeles, CA, USA

ARTICLE INFO

Keywords: Galectin-3 Rheumatoid arthritis Cardiovascular Aorta

ABSTRACT

Objectives: Galectin-3 is a beta-galactoside-binding lectin and is a marker of cardiovascular disease (CVD) in the general population. It may also play a role in joint inflammation. We asked whether serum galectin-3 is a useful marker of subclinical vascular disease in patients with rheumatoid arthritis (RA).

Methods: RA patients without clinical CVD underwent assessment of coronary artery calcium (CAC) score, aortic inflammation (using 18Fluorodeoxyglucose positron emission-computed tomography [FDG PET/CT]), and myocardial flow reserve (MFR). Aorta FDG uptake was measured as standardized uptake values (SUV). Generalized linear models were constructed to explore the associations of galectin-3 levels with CAC score, aortic SUV, and MFR.

Results: A total of 124 RA patients (mean age 57; 82 % women, 45 % Hispanic; median RA duration 6.8 years; 75 % seropositive; median CDAI 16; 33 % on prednisone; 89 % on DMARDs; median CAC score 0; median aorta SUV 2.59; mean MFR 2.86; median galectin-3 level 8.54 ng/mL) were analyzed. In univariable analysis, higher galectin-3 levels were associated with higher aortic SUV (p = 0.007) but CAC score and MFR were not. In multivariable analysis, higher galectin-3 level remained significantly associated with higher aortic SUV (β Coefficient=0.1786, p value=0.002).

Conclusion: In our cohort of RA patients without clinical CVD, higher serum galectin-3 levels were independently associated with higher levels of aortic inflammation, but not CAC score or MFR. This suggests that galectin-3 may be a biomarker for an inflammatory and potentially reversible stage, but not a later (calcified) stage, of atherosclerosis in patients with RA.

Introduction

Cardiovascular disease (CVD) is the leading cause of excessive deaths in people with rheumatoid arthritis (RA) [1–3]. Current prediction tools for future CVD developed for the general population do not perform well in individuals with RA [4]. Biomarkers that augment prediction tools in identifying RA patients at high risk for CV events could enable better CV risk stratification and earlier intervention in RA patients.

Galectin-3 is a beta-galactoside-binding lectin found in both the intra- and extra-cellular environment in a wide range of tissues that has been implicated as both an indicator of inflammation as well as of CVD

[5,6].

In a murine model of antigen-induced arthritis, galectin-3 knock out resulted in significantly reduced joint inflammation and bone erosion compared to wild-type (WT) mice [7]. In humans, galectin-3 levels were elevated in the sera and synovia of patients with RA compared with healthy controls or individuals with osteoarthritis (OA) [8], and were elevated in patients with undifferentiated arthritis who evolved into classifiable RA compared to those who did not [9].

In the general population, higher galectin-3 levels predicted incident heart failure (HF) [10] and serum galectin-3 level is FDA approved as a diagnostic marker for risk stratification and prognosis in HF evaluation

Available online 15 January 2024

0049-0172/© 2024 The Author(s). Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

^{*} Corresponding author at: 630W 168th P&S 3-450, New York, NY, 10032, USA. *E-mail address*: Ep2899@cumc.columbia.edu (E. Park).

https://doi.org/10.1016/j.semarthrit.2023.152357

[11]. Galectin-3 is also thought to be a key role in atherogenesis. In a murine model of atherosclerosis, there was increased expression of galectin-3 in intimal atherosclerotic plaques versus WT mice, and the atherosclerotic mice had higher serum levels of galectin-3 than WT mice [12]. Galectin-3 levels were detected in both early and advanced atherosclerotic lesions of carotid and limb arteries, localized by immunoperoxidase staining to macrophages and foam cells, suggesting a key role in early generation of atherogenesis- i.e., macrophage migration, lipid loading, and smooth cell proliferation [13].

Although similar studies in human arteries are lacking, galectin-3 levels were positively correlated with carotid intima-media thickness [14] and severity of coronary artery disease [15], and an elevated serum galectin-3 level has been shown to be an independent predictor of CVD mortality risk in individuals with coronary artery disease (CAD) [16].

Research exploring a potential role for galectin-3 in mediating CVD in patients with RA is limited to the work of Anyfanti et al. [17], who found an independent association between galectin-3 levels and cardiac output and systemic vascular resistance. However, the possible association of galectin-3 with joint inflammation [7–9] could confound its relationship with CVD in individuals with RA. We asked whether galectin-3 is an independent marker of subclinical vascular disease in an RA cohort extensively phenotyped for articular and subclinical CV disease.

We used coronary artery calcium (CAC) score, and 18Fluorodeoxyglucose positron emission-computed tomography (FDG PET/CT) of the ascending aorta, as structural and functional measures of atherosclerosis, respectively. FDG uptake in the vascular wall is thought to represent inflammation in atherosclerotic plaques [18], and is associated with future clinical CV events [19-20] but is reversible with treatment such as statins [21]. In contrast, CAC represents a more advanced (calcified) stage of atherosclerosis and does not appear to be reversible with statins [22]. Both measures were previously reported in our RA cohort without known clinical CVD [23]. Myocardial flow reserve (MFR) was used as a measure of dysfunction (impaired vasodilation) of the myocardial microvasculature and it also has been characterized in this same cohort [24]. We hypothesized that higher serum galectin-3 levels would be significantly associated with one or more of these vascular measures (higher CAC score, higher aortic inflammation, reduced MFR) in our cohort of RA patients without known clinical CVD, despite adjustments for levels of systemic and articular inflammation.

Materials and methods

Patients

The study sample consisted of 124 patients with RA enrolled in Rheumatoid Arthritis Study of the Myocardium (RHYTHM) who had a baseline serum galectin-3 level and all three cardiac measures (CAC, FDG PET/CT and MFR). RHYTHM has been described in detail previously [25-27]. Inclusion criteria were age >18 years and fulfillment of the 2010 American College of Rheumatology/European League Against Rheumatism criteria for RA [28]. Exclusion criteria included a prior CV event, defined as a self-reported physician-diagnosed myocardial infarction, HF, coronary artery revascularization, angioplasty, peripheral arterial disease or related procedures, pacemaker, or defibrillator devices. Other exclusion criteria included presence of active cancer and contraindication to pharmacologic study agents (i.e., regadenoson or adenosine). The study was in compliance with the Declaration of Helsinki and was approved by the Columbia University Institutional Review Board. All participants were provided written consent prior to participation.

Clinical covariables

Demographic and lifestyle characteristics were assessed by structured patient interviews using standardized health questionnaires. Current and past use of glucocorticoids and of disease-modifying antirheumatic drugs (DMARDs) was ascertained by patient interview and medical records, along with documentation of prescription and over-the-counter medications taken within the preceding two weeks verified by pill containers supplied by participants. Conventional synthetic DMARDs (csDMARD) included methotrexate, sulfasalazine, hydroxychloroquine, and leflunomide. Biologic DMARDs (bDMARDs) included adalimumab, etanercept, infliximab, certolizumab, golimumab, tocilizumab, abatacept, anakinra, and rituximab. Body mass index (BMI) was calculated as patient weight (in kilograms) divided by height (in square meters). Resting blood pressure (BP) was measured 3 times, with the mean of the last 2 measurements used. Hypertension (HTN) was defined as a systolic BP of \geq 140 mm Hg, diastolic BP of \geq 90 mm Hg, or use of antihypertensive medications. Diabetes was defined as a fasting serum glucose level of \geq 126 mg/dl or use of antidiabetic medications. RA disease duration was measured in years from the date of physician diagnosis. Disease activity was calculated based on the 28 joint count using the Clinical Disease Activity Index (CDAI), and the Disease Activity Score using C-reactive protein (DAS28-CRP) formulas [29-31]. The Health Assessment Questionnaire (HAQ) was used to assess functional impairment [32].

Laboratory measurements

Phlebotomy was performed after an overnight fast. Serum and plasma were separated by centrifugation and stored at -80 °C. All samples were measured in the Biomarkers Core Laboratory of the Columbia University Irving Institute for Clinical and Translation Research. Galectin-3 levels were measured by ELISA, reported in ng/mL (Quantikine®ELISA kit, R&D Systems, Intra-Assay Precision Coefficient of Variations 3.8 %, Inter-Assay Precision Coefficient of Variations 6.3 %). Anti-cyclic citrullinated peptide antibody (anti-CCP) was measured by ELISA (Quanta Lite CCP3 IgG kit, Inova Diagnostics), with seropositivity defined as ≥ 60 units. Rheumatoid factor (RF) was measured by ELISA (IBL America), with seropositivity defined as ≥ 40 units.

Imaging

¹⁸Fluorodeoxyglucose [FDG] positron emission-computed tomography [FDG PET/CT]:

Aortic inflammation was assessed and quantified via FDG-PET/CT scan using reproducible and validated methods [18,33]. The scan was performed on the same visit as the phlebotomy visit, after 18 h of a carbohydrate-free diet and an overnight fast. Analyses were restricted to the ascending aorta. The area of interest was defined as the region of the aorta beginning 1 cm above the origin of the left main coronary artery and ending at the aortic arch in trans-axial view. Ascending aortic inflammation was determined by FDG uptake in the arterial wall, calculated as the standardized uptake value (SUV). The outcome measure used in our multivariable models was maximal SUV of the most diseased segment (SUV MDS max) as defined previously [21,33].

Coronary artery calcium (CAC). Subclinical atherosclerosis was assessed using CAC. CAC was quantified by the Agatston method using the cardiac CT images obtained for attenuation correction and anatomic coregistration with the PET scan [34].

Myocardial flow reserve (MFR). Microvascular dysfunction was assessed by calculating myocardial flow reserve (MFR). This was done by measuring myocardial blood flow (MBF) before (rest) and after vasodilation (stress) using PET/CT with N-13 ammonia as a perfusion tracer, as described previously [24]. The ratio of the MBF measured at maximal vasodilation over the MBF at rest is referred to as MFR, which is a measure of the vasodilatory reserve of the myocardium. Reduced MFR in the absence of flow-limiting CAD is believed to reflect dysfunction in the myocardial microvasculature. Detailed methods for assessing MFR in this cohort have been published previously [24].

Statistical analysis

All analyses were performed on baseline data. Galectin levels and CAC were measured on all 124 patients enrolled, while aortic FDG uptake and MFR were measured in 89 and 80, respectively, at baseline. Summary statistics for variables were calculated. Means and standard deviations for normally distributed, and medians and interquartile ranges for non-normally distributed variables, were calculated. For categorical variables, counts and percentages were calculated. Univariable linear regression models were used to explore the associations of RA patient characteristics, treatment, atherosclerosis risk factors, and measures of cardiac and arterial structure and function, with log transformed galectin-3 levels. To assess the independent association of galectin-3 level and aortic inflammation, variables that were associated with the log transformed galectin-3 level in the univariable models at a significance level of p < 0.30 were carried into the extended multivariate linear regression models. The extended multivariate model was reduced further to a limited multivariable linear regression model by including all the variables from the extended multivariable model with *p* values <0.30, to account for the possibility of residual confounding. We also performed multivariable analyses stratified by age, gender, race, treatment type, CCP or RF positivity, disease activity, smoking status, and CAC. In all tests, a 2-sided alpha level of 0.05 was considered statistically significant. Serum galectin level was modeled as a continuous variable, but split into tertiles in Tables 1 and 2 for the summary statistics, for easier visualization and interpretation. All statistical calculations were performed using Stata software.

Results

Patient characteristics

Baseline characteristics of the 124 RA patients are summarized in Tables 1 and 2. The mean age was 57 years. The majority were women (82%), of diverse race and ethnicity (45% Hispanic), and overweight on average (mean BMI of 29). Median RA duration was 6.8 years. Seventyfive percent were RF or CCP positive. The majority of patients had low to moderate disease activity, with median DAS-CRP of 3.91 and CDAI of 16. One third of patients were currently on prednisone, with median dose of 5 mg daily. The majority (79 %) of patients were prescribed a csDMARD and 28 % of those were also prescribed a bDMARD. Ten percent of patients were on a bDMARD only, and 11 % of patients were not on any DMARD. Forty-three percent were current or former smokers, 38 % had hypertension, 11 % had diabetes, 15 % were on a statin, and 10 % were on aspirin. The majority (66 %) of participants had a CAC score of zero with only 10 % of patients having a CAC score > 300. The median ascending aorta SUV MDS max was 2.59. The mean MFR was 2.86. Median galectin-3 level was 8.54 ng/mL.

Univariable associations of patient characteristics and galectin-3 level

Baseline characteristics are summarized in Tables 1 and 2. In univariable analyses, older age, race, longer RA disease duration, current prednisone use, type of treatment, higher triglyceride level, presence of diabetes, and higher levels of ascending aorta inflammation as measured by SUV were all associated with increased galectin-3 levels. Measures of disease activity (DAS28-CRP, CDAI) and systemic inflammation (IL-6

Table 1

Baseline	characteristics	of RA	cohort ((demos	graphics	, RA	characteristics	and	current.

	All natients	Patients in 1st tertile of galectin-	Patients in 2nd tertile of	Patients in 3rd tertile of galectin-	n value
	riii puticitis	3 level	galectin-3 level	3 level	† value
		[3.3–7.33 ng/mL]	[7.44–9.51 ng/mL}	[9.54-38.44 ng/mL]	
	(n = 124)	(n = 42)	(n = 42)	(n = 42)	
Demographics					
Mean Age – years	$56.5 \pm 12.26.$	51.76 ± 13.38	55.07 ± 11.43	57.10 ± 11.55	0.008
Female – no. (%)	101 (81.45)	31 (73.81)	35 (85.37)	35 (85.37)	0.745
Race/Ethnicity					
Non-Hispanic White – no. (%)	43 (35.25)	18 (43.90)	15 (37.50)	10 (24.39)	ref
Non-Hispanic Black – no. (%)	19 (15.57)	4 (9.76)	7 (17.50)	8 (19.51)	0.344
Hispanic – no. (%)	55 (45.08)	15 (36.59)	18 (45.00)	22 (53.66)	0.039
Other – no. (%)	5 (4.10)	4 (9.76)	0	1 (2.44)	0.904
RA Characteristics					
Median RA duration – years	6.8 (2–14.2)	6.9 (2–13)	6.6 (1.5–17.6)	6.9 (2.8–16.6)	0.013
RF positive – no. (%)	68 (54)	22 (52.38)	26 (63.41)	20 (48.78)	0.774
Anti-CCP positive – no. (%)	84 (67.74)	28 (66.67)	31 (75.61)	25 (60.98)	0.566
Nodules Present – no. (%)	10 (8.13)	3 (7.14)	1 (2.50)	1 (2.44)	0.425
Median IL-6 pg/mL [#]	2.30	1.96 (1.10–3.73).	2.48 (1.24-8.71)	2.81 (1.88-8.24)	0.096
	(1.38 - 7.53)				
Median CRP mg/L [#]	2.48	2.24 (1.09-6.6)	2.09 (0.77-6.06)	3.26 (1.21-8.32)	0.617
	(1.09-6.67)				
Median DAS28-CRP	3.91	3.68 (2.63–4.44)	3.68 (2.81-4.70)	4.30 (3.35–4.84)	0.122
	(2.98-4.69)				
Median CDAI	16.4 (8.6–28.1)	15.5 (6.5–24.9)	12.7 (8.1–31.3)	21.8 (12.1–32.1)	0.075
Median HAQ		0.94 (0.38–1.5)	1.34 (0.88–1.88)	1.25 (0.63–2.0)	0.058
Current Treatment With:					
Prednisone use- no. (%)	41 (33.06)	9 (21.43)	15 (36.59)	17 (41.46)	0.038
Median prednisone dose (mg)	5 (4–10)	5 (4–7.5)	5 (2.5–10)	5 (4.5–7.5)	0.406
Conventional DMARD with no biologic	63 (50.81)	23 (54.76)	20 (48.78)	20 (48.78)	ref
– no. (%)					
Conventional DMARD with biologic -	35 (28.23)	13 (30.95)	13 (31.71)	9 (21.95)	0.748
no. (%)					
Biologic only – no. (%)	12 (9.68)	2 (4.76)	2 (4.88)	8 (19.51)	0.006
No DMARD – no. (%)	14 (11.29)	4 (9.42)	6 (14.63)	4 (9.76)	0.932

RA = rheumatoid arthritis; RF = rheumatoid factor; anti-CCP = anti-cyclic citrullinated peptide; IL-6 = interleukin-6; CRP = C-reactive protein level; DAS28-CRP = Disease Activity Score in 28 joints using the CRP level; HAQ = Health Assessment Questionnaire; DMARD = disease-modifying antirheumatic drug. Means reported with (Standard Deviation); Medians reported with (Interquartile range).

 † *p* value of univariable linear regression association between patient characteristics against the log transformed value of galectin-3 level. Values <0.05 represent statistical significance.

[#] Values log-transformed when included in univariable regression model.

Table 2

Baseline characteristics of RA cohort (cardiovascular risk factors and outcome.

	All patients $(n = 124)$	Patients in 1st tertile of galectin-3 level [3.3-7.33 ng/mL] (n = 42)	Patients in 2nd tertile of galectin-3 level [7.44-9.51 ng/mL] (n = 42)	Patients in 3rd tertile of galectin-3 level [9.54- 38.44 ng/mL] (n = 42)	p value
Cardiovascular risk factors					
Ever smoker – no. (%)	53 (43.09)	16 (38.10)	18 (45.00)	19 (46.34)	0.193
Current smoker – no. (%)	15 (12.20)	5 (11.90)	6 (15.00)	4 (9.76)	0.775
HTN – no. (%)	48 (38.71)	9 (21.43)	22 (53.66)	17 (41.46)	0.119
Mean LDL mg/dL	110.35 ± 32.36	112.62 ± 26.07	115.68 ± 35.75	102.70 ± 33.92	0.200
Mean HDL mg/dL	59.9 ± 19.15	61.29 ± 22.40	59.24 ± 18.51	57.29 ± 16.20	0.179
Median Triglycerides mg/d #	91.5 (75–132)	84.5 (74–112)	96 (77–138)	90 (75–140)	0.019
Diabetes – no. (%)	14 (11.29)	1 (2.38)	8 (19.51)	5 (12.20)	0.033
Statin use – no. (%)	19 (15.32)	5 (11.90)	6 (14.63)	8 (19.51)	0.259
Aspirin use – no. (%)	12 (9.68)	2 (4.76)	8 (19.51)	2 (4.88)	0.99
Mean BMI kg/m2	28.56 ± 5.79	$\textbf{28.67} \pm \textbf{5.47}$	26.92 ± 4.98	30.05 ± 6.53	0.194
Median Galectin level ng/mL [#]	8.54	5.87 (4.53-6.76)	8.55 (7.9–9.05)	13.4 (10.04–17.5)	n/a
	(6.64–10.03)				
Measures of atherosclerosis and vascular					
inflammation					
CAC Score Median CAC score [#]	0 (0–34)	0 (0–28)	0 (0–9.5)	0 (0–139)	0.121
CAC score of zero – no. (%)	80 (65.57)	29 (72.50)	28 (70.00)	22 (53.66)	0.093
CAC score $\geq 100 - no.$ (%)	23 (19.01)	5 (12.50)	6 (15.00)	12 (29.27)	0.057
CAC score \geq 300 – no. (%)	12 (9.92)	3 (7.50)	3 (7.50)	6 (14.63)	0.345
Standardized Uptake Values (SUV) at the Aorta [‡]					
Median SUV max [#]	2.43	2.30 (1.97-2.83)	2.52 (2.18-2.75)	2.54 (2.32-2.73)	0.007
	(2.15 - 2.77)				
Median SUV MDS max [#]			2.62 (2.36-2.97)	2.82 (2.4–3.32)	0.007
Median SUV mean [#]	1.83 (1.6-2.09)	1.68 (1.49–2.01)	1.87 (1.64–2.1)	1.93 (1.71–2.19)	0.003
Median SUV MDS mean [#]	1.91	1.73 (1.5–2.04)	1.92 (1.71-2.18)	2.06 (1.74-2.29)	0.004
	(1.64-2.19)				
Mean MFR*	$\textbf{2.86} \pm \textbf{0.73}$	$\textbf{2.98} \pm \textbf{0.85}$	2.81 ± 0.66		0.749

HTN: hypertension; LDL = low-density lipoprotein cholesterol; HDL = high-density lipoprotein cholesterol; BMI = body.

mass index; CAC = coronary artery calcification; SUV = standardized uptake value in units; max = maximum; MDS = most diseased segment; MFR= myocardial flow reserve.

Means reported with (Standard Deviation); Medians reported with (Interquartile range).

[†] *p* value s of univariable linear regression association between patient characteristics against the log transformed value of galectin-3 level. Values <0.05 represent statistical significance.

[#] Values log-transformed when included in univariable regression model.

 ‡ n = 89.

* n = 80.

and CRP) were not associated with galectin-3 level. CAC score and MFR were not associated with galectin-3 level.

Multivariable association of galectin-3 level with ascending aortic inflammation

The association between galectin-3 level and arterial inflammation, as measured by SUV MDS max, was further explored using multivariable models. In our final reduced multivariable model, higher galectin-3 level remained significantly associated with greater arterial inflammation as measured by SUV MDS max in the ascending aorta, after adjusting for treatment type and BMI (Table 3).

Interestingly, the association of galectin-3 level with aortic SUV MDS max differed by gender. Each log unit higher galectin-3 level was associated with a 0.22 unit higher aortic SUV MDS max in women (p =0.001), but was but was not significantly associated in men (p = 0.720) (p for interaction = 0.030) (Fig. 1). The association of galectin-3 level with aortic SUV MDS max also differed by race. Each log unit higher galectin-3 level was associated with a 0.27 unit higher aortic SUV MDS max in Non-Hispanic Whites (p = 0.004), but was not significantly associated in Non-Whites (p = 0.254) (p for interaction = 0.089) (Fig. 2). The association of galectin-3 level with aortic SUV MDS max also differed by treatment type. Higher galectin-3 level was significantly associated with higher aortic SUV MDS max in patients who were not receiving biologics (B coefficient= 0.2955, p value = <0.001), but was not significantly associated in patients receiving biologics (B coefficient = -0.0034, p value = 0.961) (p-value for interaction = 0.004). The association of galectin-3 level with aortic SUV MDS max did not differ as

a function of age, CCP or RF positivity, DAS28-CRP, smoking status, or presence of CAC (data not shown).

Discussion

Previous research suggests a potential association between galectin-3 level and joint inflammation in patients with RA, while research in the general population supports an association between galectin-3 level and various outcome measures of CVD. The association of galectin-3 with disease activity could hence confound the relationship between galectin-3 and CVD in individuals with RA and thus limit its usefulness as a marker of CVD in RA. Research exploring serum galectin-3 level and measures of CVD in RA cohorts is limited. In our cohort of RA patients without known clinical CVD, we found that higher serum galectin-3 levels were associated with higher levels of aortic inflammation but not with articular or systemic inflammation, a distinguishing feature from the few prior RA studies. Moreover, galectin-3 level was not associated with advanced atherosclerosis, as measured by CAC score, or with microvascular dysfunction, as measured by MFR. These results suggest that galectin-3 may be a biomarker for an earlier, inflammatory and potentially reversible stage, but not a later (calcified) stage, of atherosclerosis in patients with RA.

Although early studies in mice and in humans suggested a positive signal between galectin-3 levels and joint inflammation [7–9], galectin-3 was not associated with any measures of RA disease activity or systemic inflammation in our cohort. Oshima et al., one of the first groups to study galectin-3 in RA, found a positive association between

Table 3

Multivariable associations of RA patient characteristics and aortic SUV MDS max.

Log Aortic SUV MDS max	Extended multivariable model n = 89		Reduced multivariable model $n = 89$	
	ß Coefficient	<i>p</i> - value	ß Coefficient	<i>p</i> - value
Log galectin-3 level per ng/mL	0.1856	0.007	0.1786	0.002
Age – years	0.0025	0.308		
Race/Ethnicity				
Non-Hispanic White	ref	ref		
Non-Hispanic Black	-0.0027	0.974		
Hispanic	0.0127	0.844		
Other	0.1047	0.445		
RA duration – years	-0.0009	0.717		
Log IL-6 level per pg/mL	-0.0273	0.523		
DAS28-CRP	0.0036	0.893		
HAQ	0.0022	0.955		
Current Prednisone use	-0.0521	0.409		
Type of Treatment Receiving				
Conventional DMARD with no biologic	ref	ref	ref	ref
Conventional DMARD and biologic	0.1287	0.039	0.1048	0.183
Biologic only	-0.0712	0.434	-0.1001	0.051
No DMARD	-0.0034	0.968	-0.0293	0.693
Ever smoker	0.0560	0.322		
HTN	0.0026	0.966		
Diabetes	-0.0509	0.633		
Statin use	-0.0477	0.533		
BMI kg/m ²	0.0115	0.015	0.2411	0.123
Prob > F	0.0918		0.0004	
Adjusted R2	0.1049		0.1876	

The extended multivariable model includes variables from Table 1 with a univariable association *p* value <0.30. Statin use was selected as a surrogate for HDL, LDL and triglyceride levels. The reduced multivariable model includes all of the variables from the extended model with *p* value s <0.3. See Tables 1 and 2 for abbreviations.



Fig. 1. Multivariable association of galectin-3 level with ascending aorta inflammation stratified by gender.

serum galectin-3 level and CRP, but the treatment status was not reported in the paper, and this study was published almost two decades ago in a small sample size, when biologic DMARD treatment options were limited, [8]. Congruent to our findings, Anyfanti et al. did not find any association between galectin-3 level and ESR, CRP, or DAS score in their cohort, in which all patients were receiving DMARD treatment [17]. Mendez-Heurgo et al. also did not find an association between galectin-3 levels and measures of RA disease activity including ESR, DAS-28, or VAS in their cohort, and their patients were treated with at least one DMARD [35].



Fig. 2. Multivariable association of galectin-3 level with ascending aorta inflammation stratified by race.

Higher FDG uptake in the aortic vessel wall has been shown to correlate with macrophage infiltrates and is a risk factor for plaque rupture and future CV events [19,20]. Moreover, statin therapy and RA disease modifying agents have been shown to reduce vascular inflammation as measured by FDG/PET CT [21,36].

The higher serum concentrations of galectin-3, which correlated with higher aortic FDG uptake in our cohort, may reflect active arterial inflammation and plaque instability. In a murine model of atherosclerosis, higher intra-plaque galectin-3 level correlated with higher degrees of active inflammation, measured by macrophage content in histopathologic samples, and treatment with atorvastatin decreased the intraplaque galectin-3 concentration [12]. In clinical studies of patients with angiography proven CAD, serum galectin-3 levels were significantly higher in patients with unstable angina and acute MI compared with those with stable angina [37].

In contrast, CAC score appears to correlate with more stable plaque [38,39]. This could be a potential reason why no statistically significant relationship was found between galectin-3 level and CAC score in our RA cohort. Although in the RHYTHM cohort we did not measure carotid intimal thickness (cIMT), another measure of atherosclerosis, Anyfanti et al. reported a positive association between galectin-3 level and cIMT in RA patients [17].

Interestingly, the independent relationship between higher galectin-3 level and higher levels of aortic inflammation differed based on sex, race, and treatment type, but not on age, seropositivity, DAS28-CRP, smoking status, or presence of CAC. When the cohort was stratified by sex, the association was only significant in women. This is similar to a non-RA study in which galectin-3 level was significantly associated with incident HF only in women, not men [40]. We acknowledge the possibility of being unable to detect significant associations in men, however, due to the small number enrolled in RHYTHM (n = 23). Race also influenced the relationship, as the association between galectin-3 and aortic inflammation was significant only in whites compared to non-whites. This finding has also been seen in HF, where in a cohort of 1375 white and 434 black individuals, galectin-3 level was independently associated with composite HF or death in whites, but not in blacks [41].

Treatment type also appears to play a role. In our cohort, higher galectin-3 level was significantly associated with higher aortic inflammation in patients who were not on biologics, but not in patients who were on biologics. The majority of biologics prescribed to the RHYTHM patients were TNF-inhibitors. Previous studies showed that galectin-3's relationship with TNF is complex, and dependent on tissue type and disease. For example, galectin-3 induced higher levels of TNF in synovial fibroblasts than in skin fibroblasts [42]; however, in a model of viral induced hepatitis, galectin-3 attenuated TNF-mediated death of

hepatocytes [43]. When an RA patient is being treated with a biologic, the complex interaction between galectin-3 and other cytokines is disrupted and may explain why no relationship was found in this subgroup.

In addition to investigating the association between galectin-3 level and arterial inflammation and CAC as outcome measures of atherosclerosis, we also examined the association between galectin-3 level and MFR, where reduced MFR is thought to reflect microvasculature dysfunction (reduced vasodilation) of the myocardium. We found no association between serum galectin-3 level and MFR in our RA cohort; however, this is the first study to ever look at the association between galectin-3 level and MFR, and more research is needed in this area to draw any conclusions.

Additionally, comparing galectin-3 levels from our study across other non-RA studies remains challenging due to wide variabilities in assay used as well as cohort characteristics . One large general population study of over 3000 patients reported serum galectin-3 levels (first quartile) between 3.9–11.1 ng.ml for men and 5.0 to 12.0 ng/ml for women, which would fit most of our patients in this study [10].

Strengths of our study include a unique cohort with extensive structural and functional CV phenotyping and racial and ethnic diversity, as well as a range of disease activity, disease duration and treatments, making it reflective of a real-world RA population. Limitations of our study include its cross-sectional nature, a relatively small number of patients with arterial FDG uptake and MFR measurements, and the lack of a non-RA control group.

Conclusions

In conclusion, our results suggest that serum galectin-3 level may serve as a biomarker for an early, active, unstable, and potentially reversible inflammatory stage of atherosclerosis, but not a later stage of atherosclerosis, in patients with RA. It is clear from the growing body of literature that galectin-3's role in both atherogenesis and RA is complex and deserves additional study.

Clinical significance

Galectin-3, a beta-galactoside-binding lectin, is a marker of cardiovascular disease (CVD) and heart failure (HF). The role of Galectin-3 has yet been explored in systemic inflammation or joint disease. In our cohort of RA patients without clinical CVD, high levels of galectin-3 were independently associated with higher levels of aortic inflammation.

Funding

This work was supported by the National Institute of Arthritis and Musculoskeletal and Skin Diseases under Award Number AR-050026 (JMB), and by the National Institutes of Health National Center for Translational Science Clinical and Translational Science Award (CTSA) grant UL1 TR000040, of the National Institutes of Health, and by the Rheumatology Research Foundation Award Number RHEUMARF CU12-3892.

Declaration of competing interest

The authors declare there is no conflict of interests.

Acknowledgements

None.

References

- Nicola PJ, Crowson CS, Maradit-Kremers H, Ballman KV, Roger VL, Jacobsen SJ, et al. Contribution of congestive heart failure and ischemic heart disease to excess mortality in rheumatoid arthritis. Arthrit Rheum 2006;54(1):60–7.
- [2] Lee YK, Ahn GY, Lee J, Shin JM, Lee TH, Park DJ, et al. Excess mortality persists in patients with rheumatoid arthritis. Int J Rheum Dis 2021;24(3):364–72.
- [3] Gonzalez A, H Maradit Kremers, Crowson CS, Nicola PJ, Davis III JM, Therneau TM, et al. The widening mortality gap between rheumatoid arthritis patients and the general population. Arthrit Rheum 2007;56(11):3583–7.
- [4] Crowson CS, Matteson EL, VL Roger, Therneau TM, Gabriel SE. Usefulness of risk scores to estimate the risk of cardiovascular disease in patients with rheumatoid arthritis. Am J Cardiol 2012;110(3):420–4.
- [5] Sciacchitano S, Lavra L, Morgante A, Ulivieri A, Magi F, De Francesco GP, et al. Galectin-3: one molecule for an alphabet of diseases, from A to Z. Int J Mol Sci 2018;19(2).
- [6] Dumic J, Dabelic S, Flögel M. Galectin-3: an open-ended story. Biochim Biophys Acta 2006;1760(4):616–35.
- [7] Forsman H, Islander U, Andréasson E, Andersson A, Onnheim K, Karlström A, et al. Galectin 3 aggravates joint inflammation and destruction in antigen-induced arthritis. Arthritis Rheum 2011;63(2):445–54.
- [8] Ohshima S, Kuchen S, Seemayer CA, Kyburz D, Hirt A, Klinzing S, et al. Galectin 3 and its binding protein in rheumatoid arthritis. Arthritis Rheum 2003;48(10): 2788–95.
- [9] Issa SF, Christensen AF, Lindegaard HM, Hetland ML, Hørslev-Petersen K, Stengaard-Pedersen K, et al. Galectin-3 is persistently increased in early Rheumatoid Arthritis (RA) and associates with anti-CCP seropositivity and MRI bone lesions, while early fibrosis markers correlate with disease activity. Scand J Immunol 2017;86(6):471–8.
- [10] Ho JE, Liu C, Lyass A, Courchesne P, Pencina MJ, Vasan RS, et al. Galectin-3, a marker of cardiac fibrosis, predicts incident heart failure in the community. J Am Coll Cardiol 2012;60(14):1249–56.
- [11] Yancy CW, Jessup M, Bozkurt B, Butler J, Casey Jr DE, Colvin MM, et al. 2017 ACC/AHA/HFSA Focused Update of the 2013 ACCF/AHA guideline for the management of heart failure: a report of the American College of Cardiology/ American Heart Association Task Force on Clinical Practice Guidelines and the Heart Failure Society of America. Circulation 2017;136(6):e137–ee61.
- [12] Lee Y-J, Koh Y-S, Park HE, Lee HJ, Hwang B-H, Kang M-K, et al. Spatial and temporal expression, and statin responsiveness of galectin-1 and galectin-3 in murine atherosclerosis. Korean Circ J 2013;43(4):223–30.
- [13] Natchtigal, Nachtigal M, AI-Assaad Z, Mayer EP, Monsigny M, et al. Galectin-3 expression in human atherosclerotic lesions. 1998. p. 152.
- [14] Oyenuga A, Folsom AR, Fashanu O, Aguilar D, Ballantyne CM. Plasma galectin-3 and sonographic measures of carotid atherosclerosis in the atherosclerosis risk in communities study. Angiology 2019;70:47–55.
- [15] Bošnjak I, Bedeković D, Selthofer-Relatić K, Roguljić H, Mihaljević I, Bilić-Ćurčić I. Role of galectin-3 in diagnosis and severity assessment of epicardial artery lesions in patients with suspected coronary artery disease. BMC Cardiovasc Disord 2023; 23:268.
- [16] Maiolino G, Rossitto G, Pedon L, Cesari M, Frigo AC, Azzolini M, et al. Galectin-3 predicts long-term cardiovascular death in high-risk patients with coronary artery disease. Arterioscler Thromb Vasc Biol 2015;35(3):725–32.
- [17] Anyfanti P, Gkaliagkousi E, Gavriilaki E, Triantafyllou A, Dolgyras P, Galanopoulou V, et al. Association of galectin-3 with markers of myocardial function, atherosclerosis, and vascular fibrosis in patients with rheumatoid arthritis. Clin Cardiol 2019;42(1):62–8.
- [18] Tawakol A, Migrino RQ, Bashian GG, Bedri S, Vermylen D, Cury RC, et al. In vivo 18F-fluorodeoxyglucose positron emission tomography imaging provides a noninvasive measure of carotid plaque inflammation in patients. J Am Coll Cardiol 2006;48(9):1818–24.
- [19] Paulmier B, Duet M, Khayat R, Pierquet-Ghazzar N, Laissy JP, Maunoury C, et al. Arterial wall uptake of fluorodeoxyglucose on PET imaging in stable cancer disease patients indicates higher risk for cardiovascular events. J Nucl Cardiol 2008;15(2): 209.
- [20] Rominger A, Saam T, Wolpers S, Cyran CC, Schmidt M, Foerster S, et al. 18F-FDG PET/CT identifies patients at risk for future vascular events in an otherwise asymptomatic cohort with neoplastic disease. J Nucl Med 2009;50(10):1611–20.
- [21] Tawakol A, Fayad ZA, Mogg R, Alon A, Klimas MT, Dansky H, et al. Intensification of statin therapy results in a rapid reduction in atherosclerotic inflammation: results of a multicenter fluorodeoxyglucose-positron emission tomography/ computed tomography feasibility study. J Am Coll Cardiol 2013;62(10):909–17.
- [22] Houslay ES, Cowell SJ, Prescott RJ, Reid J, Burton J, Northridge DB, et al. Progressive coronary calcification despite intensive lipid-lowering treatment: a randomised controlled trial. Heart 2006;92(9):1207–12.
- [23] Geraldino-Pardilla L, Zartoshti A, Bag Ozbek A, Giles JT, Weinberg R, Kinkhabwala M, et al. Arterial inflammation detected with (18) Ffluorodeoxyglucose-positron emission tomography in rheumatoid arthritis. Arthritis Rheumatol 2018;70(1):30–9.
- [24] Amigues I, Russo C, Giles JT, Tugcu A, Weinberg R, Bokhari S, et al. Myocardial microvascular dysfunction in rheumatoid arthritis(quantitation by (13)N-ammonia positron emission tomography/computed tomography). Circ Cardiovasc Imaging 2019;12(1):e007495.
- [25] Winchester R, Giles JT, Nativ S, Downer K, Zhang HZ, Bag-Ozbek A, et al. Association of elevations of specific T cell and monocyte subpopulations in rheumatoid arthritis with subclinical coronary artery atherosclerosis. Arthritis Rheumatol 2016;68(1):92–102.

A. Nussdorf et al.

- [26] Geraldino-Pardilla L, Giles JT, Sokolove J, Zartoshti A, Robinson WH, Budoff M, Detrano R, Bokhari S, Bathon JM. Association of anti-citrullinated peptide antibodies with coronary artery calcification in rheumatoid arthritis. Arthritis Care Res 2017;69(8):1276–81. https://doi.org/10.1002/acr.23106. Epub 2017 Jul 10.
- [27] Amigues I, Tugcu A, Russo C, Giles JT, Morgenstein R, Zartoshti A, Schulze C, Flores R, Bokhari S, Bathon JM. Myocardial inflammation, measured using 18fluorodeoxyglucose positron emission tomography with computed tomography, is associated with disease activity in rheumatoid arthritis. Arthritis Rheumatol 2019; 71(4):496–506. https://doi.org/10.1002/art.40771. Epub 2019 Feb 28.
- [28] Aletaha D, Neogi T, Silman AJ, Funovits J, Felson DT, Bingham CO, et al. 2010 Rheumatoid arthritis classification criteria: an American College of Rheumatology/ European League Against Rheumatism collaborative initiative. Ann Rheum Dis 2010;69(9):1580–8.
- [29] Inoue E, Yamanaka H, Hara M, Tomatsu T, Kamatani N. Comparison of Disease Activity Score (DAS)28- erythrocyte sedimentation rate and DAS28- C-reactive protein threshold values. Ann Rheum Dis 2007;66(3):407–9.
- [30] Prevoo ML, van 't Hof MA, Kuper HH, van Leeuwen MA, van de Putte LB, van Riel PL. Modified disease activity scores that include twenty-eight-joint counts. Development and validation in a prospective longitudinal study of patients with rheumatoid arthritis. Arthritis Rheum 1995;38(1):44–8.
- [31] Wells G, Becker JC, Teng J, Dougados M, Schiff M, Smolen J, et al. Validation of the 28-joint Disease Activity Score (DAS28) and European League Against Rheumatism response criteria based on C-reactive protein against disease progression in patients with rheumatoid arthritis, and comparison with the DAS28 based on erythrocyte sedimentation rate. Ann Rheum Dis 2009;68(6):954–60.
- [32] Fries JF, Spitz P, Kraines RG, Holman HR. Measurement of patient outcome in arthritis. Arthritis Rheum 1980;23(2):137–45.
- [33] Rudd JH, Myers KS, Bansilal S, Machac J, Rafique A, Farkouh M, et al. (18) Fluorodeoxyglucose positron emission tomography imaging of atherosclerotic plaque inflammation is highly reproducible: implications for atherosclerosis therapy trials. J Am Coll Cardiol 2007;50(9):892–6.
- [34] Agatston AS, Janowitz WR, Hildner FJ, Zusmer NR, Viamonte Jr M, Detrano R. Quantification of coronary artery calcium using ultrafast computed tomography. J Am Coll Cardiol 1990;15(4):827–32.

- [35] Mendez-Huergo SP, Hockl PF, Stupirski JC, Maller SM, Morosi LG, Pinto NA, et al. Clinical relevance of galectin-1 and galectin-3 in rheumatoid arthritis patients: differential regulation and correlation with disease activity. Front Immunol 2018; 9:3057.
- [36] Solomon DH, Giles JT, Liao KP, Ridker PM, Rist PM, Glynn RJ, Broderick R, Lu F, Murray MT, Vanni K, Santacroce LM, Abohashem S, Robson PM, Fayad Z, Mani V, Tavakol A, Bathon J, Trial Consortium TARGET. Reducing cardiovascular risk with immunomodulators: a randomised active comparator trial among patients with rheumatoid arthritis. Ann Rheum Dis 2023;82(3):324–30.
- [37] Falcone C, Lucibello S, Mazzucchelli I, Bozzini S, D'Angelo A, Schirinzi S, et al. Galectin-3 plasma levels and coronary artery disease: a new possible biomarker of acute coronary syndrome. Int J Immunopathol Pharmacol 2011;24(4):905–13.
- [38] Criqui MH, Denenberg JO, Ix JH, McClelland RL, Wassel CL, Rifkin DE, et al. Calcium density of coronary artery plaque and risk of incident cardiovascular events. JAMA 2014;311(3):271–8.
- [39] Oyenuga A, Folsom AR, Fashanu O, Aguilar D, Ballantyne CM. Plasma galectin-3 and sonographic measures of carotid atherosclerosis in the atherosclerosis risk in communities study. Angiology 2018;70(1):47–55.
- [40] Suthahar N, Lau ES, Blaha MJ, Paniagua SM, Larson MG, Psaty BM, et al. Sexspecific associations of cardiovascular risk factors and biomarkers with incident heart failure. J Am Coll Cardiol 2020;76(12):1455–65.
- [41] McEvoy JW, Chen Y, Halushka MK, Christenson E, Ballantyne CM, Blumenthal RS, et al. Galectin-3 and risk of heart failure and death in blacks and whites. J Am Heart Assoc 2016;5(5).
- [42] Filer A, Bik M, Parsonage GN, Fitton J, Trebilcock E, Howlett K, et al. Galectin 3 induces a distinctive pattern of cytokine and chemokine production in rheumatoid synovial fibroblasts via selective signaling pathways. Arthritis Rheum 2009;60(6): 1604–14.
- [43] Stojanovic B, Milovanovic J, Arsenijevic A, Stojanovic B, Strazic Geljic I, Arsenijevic N, et al. Galectin-3 deficiency facilitates TNF-α-dependent hepatocyte death and liver inflammation in MCMV infection. Front Microbiol 2019;10:185.